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Harold Robinson A Monograph on
Foliar Anatomy of
the Genera *Connellia*,
Cottendorfia, and *Navia*
(Bromeliaceae)

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ABSTRACT

Harold Robinson. A Monograph on Foliar Anatomy of the Genera *Connellia*, *Cottendorfia*, and *Navia* (Bromeliaceae). *Smithsonian Contributions to Botany*, 2: 1-41. 1969.—The 102 presently known species of the pitcairnioid genera *Connellia*, *Cottendorfia*, and *Navia* are discussed, illustrated, and keyed on the basis of cross sections and epidermal peels of leaves. Comparison with other genera of the bromeliad subfamily Pitcairnioideae indicates *Connellia*, *Cottendorfia*, and *Navia* belong in a series with *Ayensua*, *Brocchinia*, *Fosterella*, *Pitcairnia*, and *Puya* that have sharply demarcated chlorenchyma and water-storage layers in the mesophyll. *Abromeitiella*, *Deuterocohnia*, *Dyckia*, *Encholirium*, and *Hechtia* seem to be another related series without such sharp demarcation. *Connellia* is considered related to a series of *Cottendorfia* species having elongate stomata. Many *Cottendorfia* species show abaxial water-storage tissue of a structure similar to *Puya* and some Amazonian species of *Pitcairnia*. In *Navia*, the species with paniculate or racemose inflorescences are found to represent three rather distinct groups, one of which seems close to *Cottendorfia*. Species of *Navia* with vascular bundles more completely enclosed in chlorenchyma, with scales replaced by trichomes, with sessile or glomerate inflorescences, or with connate and partially reduced sepals are considered specialized. A great number of substomatal variations are recorded in *Navia* that distinguish species or groups of species. *Navia lopezii* is shown to have a very modified epidermal structure with simple stomata and no scales, characters unlike any other bromeliad presently known.

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Harold Robinson

A Monograph on Foliar Anatomy of the Genera *Connellia*, *Cottendorfia*, and *Navia* (Bromeliaceae)

The pitcairnioid genera, *Connellia*, *Cottendorfia*, and *Navia*, are restricted almost entirely to the remote and relatively inaccessible part of South America known as the Guayana Highland. While one species of *Cottendorfia* and two species of *Navia* were described as early as 1830 and a few additional species from 1846 to 1929, the vast majority have been described during the years since 1930, mostly by Lyman B. Smith working with collections of Tate, Steyermark, Maguire, and others. The most complete treatments for the genera may be found in the recent summary of the Bromeliaceae of the Guayana Highland (Smith, 1967a). About eight species have been described since the 1967 work.

As presently accepted, the three genera, *Connellia*, *Cottendorfia*, and *Navia*, are distinguished as broad but apparently natural groups on the basis of floral and seed characters. However, rather marked characters have been observed within genera, such as the lack of a scape in *Cottendorfia navioides* (in a genus where it is normally well developed), and the presence of a long scape in various species of *Navia* (a genus in which it is usually lacking). In the single species, *Navia parvula*, the anterior sepal is totally aborted, a character that might be considered very significant in other groups of plants. One species, *Cottendorfia florida*, is unique in its distribution, being found in eastern Brazil, far from the Guayana Highlands and all other species of the group.

One of the primary concerns of this study was a

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search for anatomical characters that might support or refute suspected relationships derived from floral characters alone. In the process any evidence of natural or related groups has been sought to decrease the reliance on the artificial though workable key characters that are now prevalent. It is hoped that the series of keys and notes produced in the process will be useful in the determination of sterile or otherwise incomplete material. To these ends, Dr. Lyman B. Smith of the Smithsonian Institution staff, with his continuing interest, has provided time, material, and constant encouragement. Dr. Edward S. Ayensu, also of the Smithsonian, who has concerned himself increasingly with the anatomy of the Bromeliaceae has likewise provided much needed assistance.

A general anatomical treatment of the family Bromeliaceae is available in the work of Solereder and Meyer (1929); however, the work does not include any of the species of *Connellia*, *Cottendorfia*, or *Navia*. I have seen, in manuscript form, some parts of the treatment of the family by P. B. Tomlinson for the series on the Anatomy of the Monocotyledons edited by C. R. Metcalfe, Oxford University Press. This latter work does include a few of the species in the above complex.

Methods

Many characters are available in studies of bromeliad leaf anatomy, but the present effort emphasizes epidermal and stomatal structure along with the leaf structures seen in cross section. Scales and trichomes

are used to distinguish some species, but no critical review of variations in scale structure has been attempted.

The leaf sections and epidermal preparations for this study were all made free hand and mounted in Hoyer's Solution (Anderson, 1954). All specimens were obtained from fragments of dried herbarium material soaked in tap water. The final slides used in the study were unstained temporary mounts. Preparations kept more than two years clearly show the original limits of the chlorenchyma and the resinous deposits.

In taking epidermal strips and leaf sections, the leaf base has been strictly avoided. In the leaf base the chlorenchymatous mesophyll is poorly differentiated and air chambers are highly developed. The vascular bundles are surrounded with irregular groupings of small parenchyma cells, and the epidermal layers are greatly thickened. It is at a higher level that stomata begin to appear and finally are densely organized into definite bands underlying the smaller bundles or intervening mesophyll regions. In some species internal features at this higher level may still show some disorganization characteristic of the leaf base, and for critical observations only those sections from midleaf or above have been relied on.

The present study constitutes a preliminary effort with definite limitations. Even in the case of *Cottendorfia*, where sections of leaves have often been taken at various levels, conclusions must be drawn with care. Rarely are more than two levels sampled or more than one leaf, and few of the species are even known from more than one collection. In *Navia*, with its far greater number of species, observations are based on far less thorough sectioning, and the stomatal characters, which happen to be of particular interest in the genus, are emphasized.

For a list of specimens examined during this study, see page 19.

Generic Limitations

Of the three genera, *Navia*, *Cottendorfia*, and *Connellia*, the first, *Navia*, is distinguished almost entirely on the basis of the unappendaged seeds. *Connellia* is close to *Cottendorfia* but distinguished by the subbasifixed anthers and showier flowers.

This study has involved only the most cursory examination of flower parts, but what has been seen tends

to indicate uniformity of these structures in such genera as *Pitcairnia*, *Brocchinia*, *Cottendorfia*, and *Connellia*. Only the last, a small genus of four species, shows such uniformity in leaf anatomy. All the other genera listed show two or more distinct types of leaf structure. Some of the types that appear in more than one genus might indicate relationships, but some are probably the result of parallel development. There is not, at present, any reason to question that present genera represent natural groups, but there is considerable reason to regard them as very broad concepts that may be subject to some subdivision.

After observing transverse sections of the leaves of over twenty species selected by Dr. Lyman B. Smith to represent the variation in the genus *Pitcairnia*, and of nearly all species of *Brocchinia*, these genera seem relatively uniform with only two very distinct types in *Pitcairnia* and three types in *Brocchinia*. This is contrary to the impression that is given by the macroscopic leaf variation in *Pitcairnia* where the basic anatomical unity is obscured by broad bladed and slenderly petiolate, narrow bladed and deciduous, and heterophyllous variants. The variation in transverse sections in *Cottendorfia* seems at least as great, although there are certain types totally lacking, reflecting a basic uniformity in the genus. The genus *Navia* is, from all indications, the most richly varied in almost all structures, patterns of leaf cross section, stomata, and scales. One might conclude that although restricted to a climatic and topographic area lacking any great antiquity, *Navia* may represent one of the oldest lines of development in the subfamily.

The leaf anatomy does not clearly demarcate the existing generic concepts of the genera *Connellia*, *Cottendorfia*, and *Navia*, but it does show distinctive trends within the latter two genera. These trends are discussed in the following section but they are summarized as follows.

Characters sometimes found in *Navia*, not in *Connellia* or *Cottendorfia*: (1) Vascular bundles separated from adaxial water-storage cells by a layer of chlorenchyma; (2) substomatal pores with large or long intrusions or reduced to a slit; (3) leaves bearing radially symmetrical uniseriate trichomes.

Character sometimes found in *Cottendorfia*, not in *Connellia* or *Navia*: Vascular bundles in contact with well-developed areas of water-storage tissue both abaxially and adaxially.

253, 256, 261, 266). This is in contrast to the genera *Hechtia*, *Dyckia*, *Encholirium*, *Deuterocohnia*, and *Abromeitiella*, which have an area of poorly differentiated cells between, forming a transition zone. In dried material this can be very difficult to observe since water-storage cells can collapse and become obliterated or compacted into a layer that looks like a demarcation line. I would refer to sections lacking a sharp demarcation as a *Dyckia* type, which in its most typical form is very thick-leaved with vascular bundles remote from the abaxial surface, and the abaxial surface deeply ridged with stomata restricted to the deep furrows. There has not been much tendency to relate the South American genera of this group with the Mexican and northern Central American *Hechtia* because of their very divergent and rather remote distributions. On the basis of leaf anatomy, however, these genera together seem to form a related series well out of the *Navia*, *Cottendorfia*, *Connellia* relationship.

EXPOSURE OF VASCULAR BUNDLES.—The extent to which the chlorenchyma surrounds the vascular bundles seems to be a variable factor that can differ in closely related groups, at least in *Navia* (Figures 232, 239, 242, 243, 253, 256, 261, 266). In *Pitcairnia*, at least two species, *P. punicea* and *P. oaxacana*, have chlorenchyma covering the smaller bundles adaxially, while most of the species, including many seemingly closely related, have bundles all emergent against or into the adaxial water-storage tissue.

In spite of such variation between related species, the overall trends indicate the exposure of the vascular bundles is of considerable significance. The extreme situations are quite contrasting and are not found in related groups. Also, in some cases whole groups do show uniformity in the character. The bundles are covered adaxially with chlorenchyma in most species of *Navia* or show a tendency toward such cover with only the larger bundles being exposed (Figures 92, 100, 106, 109, 116, 117, 124, 128, 132, 133). I would refer to such sections as the *Navia* type. A marked exception in *Navia* is a group of perhaps primitive species having a paniculate inflorescence of the *N. reflexa* (Figure 70), *N. hohenbergioides* (Figure 79), *N. ramosa* (Figure 69) relationship, which all have bundles definitely exposed. This group is so marked that the few species that have paniculate inflorescences but with bundles covered with chlorenchyma adaxially seem only distantly related.

Such distinctly covered bundles appear elsewhere in

the Pitcairnioideae in the recently described genus, *Ayensua* (Ayensu, 1969; Smith, 1969), in at least two species of *Fosterella*, and in *Brocchinia vestita*, *B. maguirei*, *B. delicata*, *B. melanacra*, and *B. cowanii*. In *Brocchinia* the covered bundles are highly characteristic of the particular series, but the probable primitive type, *B. serrata*, and the series including the type, *B. paniculata*, have bundles not covered with chlorenchyma adaxially.

In what I would call the *Cottendorfia*-type cross section, the chlorenchyma may form large masses extending upward between the vascular bundles, but with rare exceptions there is a group of water-storage cells reaching the adaxial surface of each bundle (Figures 1, 4, 9, 15, 61, 64). This pattern is found in all species of *Cottendorfia* and *Connellia*. In *Cottendorfia* the appearance can become greatly accentuated with the development of large abaxial water-storage canals that also reach the vascular bundles (Figure 43). This extreme type of *Cottendorfia* cross section seems unlikely to have evolved very often, if more than once, and I find it significant that similar sections occur in *Puya* as represented by *P. santosii* and *P. sodiroana*, and in a small section of rather distinctive *Pitcairnia* species as represented by *P. nuda*, *P. bulbosa*, and *P. armata*. The latter three *Pitcairnia* species are from the part of the extensive range of their genus in which *Cottendorfia* is found.

The abaxial water-storage canals occur in a part of the leaf in which they are often replaced by a fibrous region connecting the vascular bundle to the abaxial epidermis. Such extensions of the sclerenchyma occur in three species of *Cottendorfia*—*C. florida* (Figure 4), *C. thyrsoidea* (Figure 24), and *C. minor* (Figure 25)—and in *Navia immersa* (Figure 136), *N. serrulata* (Figure 182), and various species of the *Navia reflexa* group (Figure 79). In these two genera there is no indication that any of the species are particularly closely related except for those in the *Navia reflexa* group. In neither of these genera do the fibrous bundle groups extend adaxially to reach or nearly reach the epidermis as sometimes happens in *Pitcairnia* or *Brocchinia*.

PLACEMENT OF VASCULAR BUNDLES.—In some species of *Navia* the central portion of the leaf may be rather distinctly thickened, but there is none of the marked alteration in arrangement or shape of the vascular bundles that is so prominent in the costa-like central portion of the leaves of many species of

Pitcairnia. The differentiation of the middle part of the leaf in *Navia* is so slight that there is no reason to assume any close relationship to that in *Pitcairnia*. In the latter genus this pseudocostate or what I would call the *Pitcairnia*-type arrangement has been present to some extent in most species seen, except those in the previously mentioned group including *P. nuda*, *P. bulbosa*, and *P. armata*. I have seen nothing I would call a *Pitcairnia*-type section in any other genus, and it seems to have evolved completely within the genus *Pitcairnia*.

The review of the various Pitcairnioideae has left the impression that the *Cottendorfia*-type leaf section, at least in its less specialized form, is primitive within the group. In *Brocchinia*, *Pitcairnia*, and *Navia* the less specialized species all or mostly seem to show such sections, and the *Navia*-type section seems to be only in obviously derived groups of species.

STOMATA.—The guard cells and subsidiary cells, except for those of the very distinctive *Navia lopezii*, are relatively uniform. They vary in relative width in both *Navia* and *Cottendorfia*. In *Cottendorfia* the genus seems divisible into natural groups on this basis (Figures 5, 7, 10, 13, 16, 19, vs. 22, 27, 29, 31, 34, 37, 41, 44, 46). No such pattern is evident in *Navia* though obviously related species tend to be alike.

In *Navia lopezii* the guard cells are very narrow and have similarly shaped lateral subsidiary cells that are broadly exposed on the outer surface (Figures 275, 277). This pattern of narrow guard cells and exposed subsidiary cells is not seen elsewhere in *Navia*, but resembles the amazingly uniform stomatal type found in most species of *Pitcairnia*. In *Pitcairnia*, only three species observed, *P. nuda*, *P. bulbosa*, and *P. armata*, which have other cottendorfioid characters, tend to have broader guard cells and more included subsidiary cells. Variation between stomata of the *Navia lopezii* type with only the two lateral subsidiary cells (paracytic) and those such as seen in other bromeliads with both lateral and polar subsidiary cells (tetracytic) has been noted by Stebbins and Khush (1961) in various families, but not previously between species of a single genus.

It is notable that the developmental influence of the stomata of most bromeliads extends laterally to the cells of the adjacent rows to the same extent it does to the polar cells, usually resulting in a true total of six subsidiary cells. In some cases the true inner lateral subsidiary cells seem to become completely included

(Figures 215, 216, 218, 219), and the outer laterals directly contact the guard cells becoming subsidiary cells in the strictest sense of the term.

SUBSTOMATAL INTRUSIONS.—The pore immediately inside the stoma is usually rounded or only slightly misshapen in most genera of the Pitcairnioideae. It has slight intrusions in some *Hechtia* or *Pitcairnia* species and is rather reduced in *Cottendorfia florida* (Figure 6). It is in the genus *Navia*, however, that the pore is subject to incredibly rich variation in shape. One totally unique variant is the special case of *N. lopezii* in which the pore does not occur because an inner layer of subepidermal cells is not formed (Figure 275). Other variations in *Navia* concern the types of intrusions projecting inward from surrounding cells. These variations include *N. breweri* (Figure 236) and *N. involucreta* (Figure 108) types in which the surrounding cells are thick-walled and intrude leaving only a small slitlike opening or no opening that can be seen from face view. In the *N. reflexa* type (Figures 72, 75) hooked thick-walled intrusions reduce the opening to a double-anchor shape. The *N. subpetiolata* type (Figure 204) shows similar but thin-walled projections. The *N. cretacea* type (Figure 161) is similar to that of *N. subpetiolata* but bears many extra lobes or papillae. The *N. parvula* type (Figure 102) consists of four separate intrusions that reduce the opening to a cruciate form. The *N. abysmophila* type (Figure 153) has the opening divided into a number of separate round pores. There are in addition a large number of species of *Navia* such as *N. myriantha* (Figure 264) with the simple structure most often found in other genera.

In *Navia* there are strong indications that stomatal types are stable not only in species but species groups. The stomatal type in *N. reflexa* is rather characteristic of the group of species, and the *Navia subpetiolata* group seems to be a natural one showing stomatal uniformity. In most of the species related to *N. myriantha* the substomatal pore is simple. The distinctive substomatal pore of *N. parvula* is correlated with the distinctive aborted sepal in the flower but *N. lepidota* with a reduced sepal has a stomatal type that is as near as any seen to that in *N. parvula*. Still, a case of obvious failure of the character is in the unmodified opening of *N. ramosa*, a species rather obviously related to *N. reflexa* and its relatives with their otherwise distinctive pore type. The similarity of the substomatal pore in *N. breweri* and *N. cardonae* might indicate

close relationship, but here the character is in conflict with the occurrence of trichomes, and in the present treatment I have given the latter character priority.

In many species (Figures 6, 141, 166, 252) I have observed a series of stomata in a row in which an extra cell is interposed in the substomatal layer that forms a bridge across the pore. The fact that this seems to occur erratically without regard to species and that it appears in a series of six to eight stomata in direct line, indicates that it is a common aberration that acts in the manner of a small chimera.

EPIDERMAL CELLS.—On the basis of material examined the epidermal cells of the Pitcairnioideae seem to hold amazingly to a normal type with oblong shape and sinuous walls. The walls tend to be straighter around the stomata and more sinuous in the part of each wall nearer the outer surface. There is considerable variation in size as can be seen in the various figures which, except for *Navia lopezii*, are drawn to the same scale. Among the smallest cells are those of *N. abyssophila* (Figure 152) while the largest epidermal cells of normal structure occur in *Navia intermedia* (Figure 134).

It is remarkable that all marked departures from this basic pattern, even though stable within a species, are presently each known only from a single species. In *Cottendorfia florida* only, the walls of the epidermal cells appear to be straight throughout. In a species outside the immediate study, *Brocchinia paniculata*, one end of each epidermal cell is much enlarged and thick-walled and sinuous walls are mostly lacking. In this species a very distinctive pattern of interlocking fish-shaped cells is formed. Other closely related species of *Brocchinia* lack this pattern.

The most distinctive form of epidermal cell is that of the unique species, *Navia lopezii* (Figures 275–277). Here the cells are extremely large, individual cells reaching nearly 1 mm in length. These cells are visible under a dissecting microscope. Similar large epidermal cells are seen in the outer floral bracts and the character is reflected to some extent in the carpel wall in the fruit. I have concluded on the basis of the epidermal structure that these cells are the result of the failure of protoderm cells to subdivide fully before maturing. As a result whole layers of tissues fail to differentiate. As illustrated in surface view (Stebbins and Khush, 1961), members of some genera in other families, *Juncus* (Juncaceae) and *Anigosanthus* (Haemodora-ceae), have similar structure and may have arisen in

a similar way. A similar process was probably responsible for the peculiar sporophyte development in the moss genus *Archidium*, where there are only eight large spores and there is no central columella. The evolution of fern sporangia and of leaves of Hymenophyllaceae ferns have also involved a reduction in numbers of cells, but these examples involve more long term trends than the abrupt development of *Navia lopezii*.

A central silica body is characteristic of the epidermal cells of most Bromeliaceae as well as some other monocot families. As seen in the Pitcairnioideae this is generally distinct in such genera as *Dyckia*, *Hechtia*, and *Navia*, and it tends to be weaker or lacking in species of *Cottendorfia*, *Pitcairnia*, and *Brocchinia*. This is only a trend, however, with many exceptions. In both *Navia lopezii* (Figures 276, 277) and *N. cataractarum* (Figure 95) the silica body seems to be entirely lacking.

SCALES.—The scales are one of the primary characteristics of the family Bromeliaceae, being found somewhere on the plant in all but one species. The most common type is similar to those seen in *Cottendorfia argentea* (Figure 40), *Navia serrata* (Figure 183), and *N. caulescens* (Figure 194), platelike with elongate cells at least toward the margin. The scales may be reduced to a plug as seen in *N. garcia-barrigae* (Figure 89) where it sits in the center of a pit covering a circle of stomata. Vestigial scales as shown in *Cottendorfia savannensis* (Figure 58) are undoubtedly common in the family, but only evident under the compound microscope. The scale type in *Navia arida* with its thin-walled rounded cells and partially resorbed margin of small cells is unique among the species in the three-genus complex, but somewhat similar ones have been seen in *Pitcairnia* and *Brocchinia*.

The most distinctive type of emergence encountered in the study is the trichome (Figures 244, 262, 265, 271, 274) seen only in certain species of *Navia*. The radially symmetrical uniseriate structure seems very distinct from all scales seen, but might represent a single branch of some of the deeply cleft types occurring in a few other species. For the present it seems reasonable to treat species so armed as a related complex, but like other characters this may vary in expression within the part of the genus where it occurs.

Even in species with leaves that seem glabrous, microscopic examination will usually reveal vestigial scales with distinct basal plugs set in the leaf. In one

species, *Navia lopezii*, the highly modified epidermis of the leaf has resulted in the elimination of all traces of scales. This species is the one bromeliad now known in which there seems to be no trace of scales on any part of the plant.

Comparative Evaluation of Floral Characters

Some floral characters used in previous keys, such as shape of sepals, have been admittedly artificial. Three characters used by Smith (1967a) in his treatment of the genus *Navia*, however, are worthy of special consideration: (1) connate versus free sepals; (2) inferior versus superior ovary; and (3) paniculate versus glomerate inflorescence.

Connate sepals, as seen in the Smith key, are characteristic of a series of Colombian species, including *Navia heliophila* and *N. graminifolia*, a fact called to my attention by Dr. Smith. In spite of some differences in general aspect, all these Colombian species show similar basic anatomy and three, *N. heliophila*, *N. acaulis*, and *N. bicolor*, are unquestionably closely related. The connate sepals prove to be characteristic of another group of closely related species, the *Navia subpetiolata* group. Here, the cross section, substomatal intrusions, and general aspect all agree and close relationship is unquestionable. Only *N. caricifolia* of this series seems particularly different and it differs only by the adaxial exposure of the vascular bundle and the somewhat more sinuous walls of the epidermal cells. There is no reason now to believe all species of *Navia* with connate sepals are closely related, but I have nevertheless grouped them together in my survey of the genus.

The inferior ovary is an important character used to distinguish *Pitcairnia* and *Brocchinia* from most other genera of the Pitcairnioideae. The character varies considerably, however, in the genus *Navia*. The species of *Navia* having inferior or partially inferior ovaries include a diverse lot of which only *N. parvula* and *N. lepidota* seem particularly closely related. Others include *N. aloifolia* which has open substomatal pores and trichomes, *N. arida* which has open substomatal pores and very distinctive scales, *N. immersa* with closed pores and vascular bundles fused to the abaxial epidermal layers, and *N. scopulorum* which has closed pores and free bundles. *Navia fontoides* has closed pores, but, like only *N. parvula* and *N. lepidota* of the series, has chlorenchyma completely covering the vas-

cular bundles adaxially. In addition, the degree of inferiority of the ovary in the group varies. No basic unity is evident between the species involved, and I have been inclined to relegate them to various groups on the basis of other characters. It is significant, of course, that while these species show no close relationship among themselves, neither do they show close relationship to any other species in the genus. They are an assemblage of some of the most singularly distinctive species in the genus.

The paniculate inflorescence is certainly primitive in the Pitcairnioideae compared to the condensed and usually sessile inflorescence of most *Navia* species. One of the basic aims of this study involved the analysis of the few *Navia* species having a paniculate inflorescence. As indicated in the discussion of the chlorenchyma distribution, these species of *Navia* are divided into two probably not too closely related groups. The group including *N. ramosa*, *N. reflexa*, *N. hohenbergioides*, and *N. garcia-barrigae* has *Cottendorfia*-type cross sections and no tendency toward the *Navia* type. This group seems to be a side branch of *Navia* evolution that, but for the unappendaged seeds, would be called *Cottendorfia*. An additional character suggesting closer relationship to *Navia* is the development of marked substomatal intrusions in many but not all of the species. In the remaining species of *Navia* having paniculate inflorescences, the cross sections are of the *Navia* type. This series of three species, *N. lindmanioides*, *N. rupestris*, and *N. cataractarum*, may represent a more primitive group near the main line of *Navia* development.

Systematic Review

Floral characters in the following generic descriptions are taken partially from Smith (1967a).

Genus *Connellia*

Flowers showy; sepals free, much shorter than the petals; petals free with a narrow claw, the blades suborbicular, brightly colored, spreading at anthesis, not twisted together afterward; stamens included, free, the anthers subbasifixed; ovary superior, glabrous, the style slender, elongate; placentae elongate, nearly the height of the locule; capsule septical; seeds narrow, bicaudate.

Leaves thin with reduced scales (except in *C. quel-*

chii), margins incurved; stomata distinctly longer than wide with narrow guard cells; vascular bundles contiguous with adaxial water-storage tissue, sometimes with poorly differentiated abaxial water-storage canals.

As presently recognized the genus contains four species which occur in the eastern Guayana Highlands from Ptari-tepui in Venezuela to Roraima.

The leaf anatomy of the species of *Connellia* appears quite uniform (Figures 1-3) and seems basically indistinguishable from the group of *Cottendorfia* spe-

cies having elongate stomata. Among these latter species epidermal scales are generally reduced, abaxial water-storage canals are usually shallow and often indistinct, and leaves in some species have distinct cartilaginous margins, all these characters being found among the species of *Connellia*. One species, *Cottendorfia longipes*, from the group with longer stomata, is possibly a *Connellia*. This species is known only from fruiting material, and flowers will be necessary for final generic placement.

Vegetative Key to Species of *Connellia*

1. Upper leaf margins with a dense fringe of scales *C. quelchii*
1. Leaf margins without noticeable scales.
 2. Leaf blade with prominent white cartilaginous margins *C. caricifolia*
 2. Leaf blade without differentiated white cartilaginous margins . . . *C. angustae*, *C. nutans*

I have not discovered any foliar characters distinguishing *C. angustae* and *C. nutans*. The abaxial water-storage canals are most highly developed in the distal part of the leaves of *C. quelchii* (Figures 1-3). They are most rudimentary in *C. caricifolia*. In all species the vascular bundles are covered by chlorenchyma abaxially in at least the lower part of the leaf.

Genus *Cottendorfia*

Floral bracts minute; flowers small; petals free with a narrow claw, the blades elliptical, white, separate and not twisted after anthesis; anthers versatile; filaments usually free; ovary superior, glabrous, the style slender; placentae short, subbasal; capsule septicidal; seeds narrow, bicaudate.

Leaves thin to rather thickened, often with dense cover of scales, margins incurved or recurved, sometimes essentially flat; stomata elongate with narrow guard cells or wider than long with broad guard cells; vascular bundles contiguous with adaxial water-storage tissue, sometimes contiguous with abaxial water-storage canals.

As presently recognized the genus contains 24 species, mostly in the eastern Guayana highlands. *Cottendorfia thyrsoides* and some of the *Lindmania* group extend into central and southern Venezuela.

The majority of *Cottendorfia* species show the abaxial water-storage canals in the leaf. In one large group of species these canals are highly developed and the vascular bundles very high and narrow giving a very characteristic appearance to the leaf cross section not seen in *Navia* or *Connellia* (Figure 43). These species have short broad stomata (Figures 22, 27, 29, 31, 34, 37, 41, 44, 46, 48, 50, 52, 54, 56, 59) and often have a dense cover of epidermal scales on the leaf surface. Another group of species with less distinct water-storage canals, more rounded vascular bundles, more elongate stomata (Figures 5, 7, 10, 13, 16, 19), without obvious scales, and in a few cases bearing a narrow pale border, shows a strong relationship to *Connellia*. The two other very divergent elements in the genus show, each in its own way, resemblance to *Navia*, *Cottendorfia florida* by the partially closed substomatal orifice (Figure 6), and *C. navioides* by the lack of a scape in the inflorescence.

Vegetative Key to Species of *Cottendorfia*

1. Epidermal cells of leaf with straight walls (Figure 5); stomata having inner opening usually narrow, partially closed by outgrowth of surrounding cells (Figure 6) *C. florida*
1. Epidermal cells of leaf with sinuous walls (Figure 7); stomata with inner pore oval or circular, fully or almost fully open (Figure 8).
 2. Leafy stems elongate, usually much longer than leaves; leaves in cross section with upper surface contoured over the massive subdermal cells (Figure 64); leaves often glaucous *C. navioides*

2. Stems short; leaves in section with upper surface showing contours of epidermal cells only (Figure 61); leaves not glaucous.
3. Stomata elongate, longer than wide (Figures 16, 19); leaf surface never covered with obvious scales; leaves often with narrow cartilaginous border.
 4. Leaves strongly involute *C. steyermarkii*
 4. Leaves mostly plane or narrowly revolute.
 5. Leaves with a layer of resinous cells internally, at least adaxially (Figure 12).
 6. Vascular bundles deeply recessed in the chlorenchyma (Figure 9) . . . *C. longipes*
 6. Adaxial surface of major vascular bundles even with upper level of chlorenchyma (Figure 12) *C. dyckioides*
 5. Leaves without layers of resinous cells (Figure 15).
 7. Leaves distinctly toothed near base *C. wurdackii*
 7. Leaves nearly or completely entire margined.
 8. Most leaves 2 cm or more broad at the base, apex broadly acute *C. brachyphylla*
 8. Most leaves about 1 cm broad at the base, apex very narrowly acute *C. tillandsioides*
 3. Stomata short, as wide as long or wider (Figures 22, 27, 29); leaf surface often densely covered with scales; leaves without distinct border.
 9. Vascular bundles (as seen in cross section) continuous with abaxial epidermal layers or separated by a single row of thinner walled cells (Figures 24, 25).
 10. Leaves mostly over 20 cm long, leaf margins plane or recurved . . . *C. thyrsoidea*
 10. Leaves less than 20 cm long, margins slightly incurved *C. minor*
 9. Vascular bundles separated from lower epidermis by at least two rows of enlarged or thin-walled cells (Figures 33, 36, 39, 43).
 11. Vascular bundles (as seen in cross section) covered by chlorenchyma abaxially (Figures 33, 36).
 12. Leaves completely without abaxial water-storage canal (Figure 36); upper leaf margin with few or no teeth *C. subsimplex*
 12. Leaves with small abaxial water-storage canals (Figure 33); leaf margins distinctly and closely serrate throughout *C. serrulata, C. gracillima*
 11. Abaxial water-storage canals reaching the vascular bundles, the bundles in contact with water-storage tissue both abaxially and adaxially (Figure 43).
 13. Leaf margins incurved *C. guianensis, C. stenophylla*
 13. Leaf margins mostly recurved.
 14. Leaves with prominent scales on the adaxial surface *C. paludosa*
 14. Leaves without distinct scales, adaxially, scales often prominent on abaxial surface *C. argentea, C. cylindrostachya, C. dendritica, C. geniculata, C. maguirei, C. nubigena, C. phelpisiae, and C. savannensis*

The genus *Cottendorfia* can be subdivided into the following four groups and eight subgroups.

GROUP A: Inflorescence scapose; epidermal cells with straight walls; stomata elongate with narrow guard cells; substomatal orifices partially closed; occurring in northeastern Brazil.

Subgroup I: One species with leaf margins entire or nearly entire; vascular bundles connected to abaxial epidermis; sepals cochleate (anterior sepal included).

Cottendorfia florida (Figures 4–6), the type species of the genus, seems unique in the group studied by the straight walls of the epidermal cells. These are often difficult to see but can be demonstrated best by viewing torn edges of the epidermis. The cochleate sepals and

the partially closed substomatal pores are reminiscent of *Navia* where there are a few species with scapose inflorescence and half-buttressed vascular bundles as in *C. florida*. Cochleate sepals and partially buttressed vascular bundles are found elsewhere in the genus *Cottendorfia*, but not combined as in the paniculate *Navia* species. It is on the basis of the appendaged seeds that the present species is most clearly distinct from *Navia*.

GROUP B: Inflorescence scapose; epidermal cells with sinuous walls; stomata elongate with narrow guard cells; substomatal orifices oval or rounded; occurring in the eastern Guayana Highlands.

Subgroup II: One species with leaf margins involute, not cartilaginous, entire; vascular bundles

not buttressed abaxially, separated by chlorenchyma from abaxial water-storage canals; sepal imbrication not determined.

In anatomical structure *C. steyermarkii* is very similar to *C. tillandsioides* (Figures 13–15) but the relatively short, incurved, and involute leaves seem very distinct. *Cottendorfia tillandsioides* along with other species of the following subgroup seem very consistent in their flattened or revolute leaves.

Subgroup III: Five species with leaf margins usually plane or reflexed, often cartilaginous, entire or sometimes toothed near the base; vascular bundles not buttressed abaxially, separated by chlorenchyma from abaxial water-storage canals; sepals where determined are convolute.

The five species share many characters with the genus *Connellia* and close relationship seems likely. The generic status of one species, *Cottendorfia longipes* (Figures 7–9), remains uncertain until flowering material is discovered. All species of the group including *C. longipes*, *C. dyckiioides* (Figures 10–12), *C. tillandsioides* (Figures 13–15), *C. brachyphylla* (Figures 16–18), and *C. wurdackii* (Figures 19–21) have distinct abaxial water-storage canals.

GROUP C: Inflorescence scapose; epidermal cells with sinuous walls; stomata as wide as long or wider, with broad guard cells; substomatal orifices oval or rounded; occurring in the central and eastern Guayana Highlands.

Subgroup IV: Two species with leaf margins often distinctly but remotely serrate; major vascular bundles buttressed abaxially; sepal arrangement not determined.

Though *Cottendorfia thyrsioidea* (Figures 22–24) and *C. minor* (Figures 25–28) share the half-buttressed vascular bundles, they may not be particularly closely related. This condition occurs also in *C. florida* and in a few species of *Navia*, species of rather remote relationship. In *C. minor* the half-buttressed condition seems to be present only in the upper half of the leaf. In other species showing the character, it seems less restricted.

Subgroup V: Two species with leaf margins closely and finely serrate; vascular bundles covered by chlorenchyma abaxially; abaxial water-storage canals present; sepal arrangement not determined.

There can be no question of the close relationship

of *C. serrulata* (Figures 29–30) and *C. gracillima* (Figures 31–32). They seem to agree in all structures other than size of inflorescence parts. In addition, they seem closely related to the members of the following two subgroups of *Cottendorfia*, being distinguished by the arrangement of chlorenchyma and water-storage tissue in the abaxial half of the leaf.

Subgroup VI: One species with leaf margins nearly entire; vascular bundles completely enclosed in chlorenchyma abaxially; without abaxial water-storage canals; sepal arrangement not determined.

The single species, *C. subsimplex* (Figures 34–36), has all the characters of the following subgroup except for the dense and continuous chlorenchyma found in the abaxial half of the leaf.

Subgroup VII: Eleven species with leaf margins entire or with teeth restricted to near the base; vascular bundles contiguous abaxially with well-developed water-storage canals; sepal imbrication where determined usually convolute, rarely cochleate.

The subgroup contains *Cottendorfia guianensis* (Figures 44–45), type-species of the old genus *Lindmania*. This widely distributed and variable species, along with *C. argentea* (Figures 40–43), *C. cylindrostachya* (Figures 48–49), *C. dendritica* (Figures 46–47), *C. geniculata* (Figures 50–51), *C. maguirei* (Figures 52–53), *C. nubigena* (Figures 54–55), *C. paludosa* (Figures 59–60), *C. phelpsiae* (Figures 61–63), *C. savanensis* (Figures 56–58), and *C. stenophylla* (Figures 37–39), have the rather thick leaves with high contiguous abaxial water-storage canals (Figure 43) that I have come to think of as most characteristic of *Cottendorfia*. I have not seen such sections in either *Navia* or *Connellia*, though they occur in those *Puya* species I have seen, and in a small group of *Pitcairnia* species. The species of this subgroup seem very closely related and most of them cannot be distinguished from each other by vegetative characters.

GROUP D: Inflorescence nonscapose, corymbose; epidermal cells with sinuous walls; stomata as wide as long or wider, with broad guard cells; substomatal orifices oval or rounded; occurring in the eastern Guayana Highlands.

Subgroup VIII. One species with leaf margins entire; vascular bundles somewhat buttressed abaxially; sepal arrangement not determined.

Cottendorfia navioides (Figures 64–66) is named for the *Navia*-like short, nonscapose inflorescence, a structure not otherwise known in *Cottendorfia*. The species is distinguished from *Navia* by the presence of appendaged seeds. The species seems unique and can easily be determined by the contour of the adaxial leaf surface as seen in cross section.

Genus *Navia*

Inflorescence of one or more distinct or agglomerated heads or in a few species paniculate; flowers large or small, sessile or pedicellate; sepals cochlear, the anterior covered by the two posterior or rarely the anterior wholly aborted; petals free with a claw, broad, naked, connate in a slender tube, blades spreading at anthesis, often cucullate with an inflexed acute apex; anthers linear, filaments usually free; ovary usually superior, sometimes partially or wholly inferior; style slender; placentae linear; ovules naked or nearly so; capsule septicidal; seeds naked or with a vestigial wing, reticulate or corrugated.

Leaves thin to much thickened, often with dense cover of scales or trichomes, margins incurved, recurved or flat, entire to strongly toothed; stomata with narrow or broad guard cells, substomatal pore often with intrusions from surrounding cells; vascular bundles either exposed to adaxial water-storage tissue or partially to completely covered by chlorenchyma, abaxial water-storage canals sometimes present but not reaching vascular bundles.

As presently recognized the genus contains 74 species, and occurs throughout the Guayana Highlands.

Many of the species have vascular bundles covered with chlorenchyma abaxially (Figures 92, 100, 106, 109, 116, 117, 124, 128, 132, 133), a character not seen in *Cottendorfia* or *Connellia*. The character seems somewhat variable in its development, but it is completely lacking from the species that seem most like *Cottendorfia* (Figures 69, 70, 79, 88), and is well developed in almost all species with connate sepals or trichomes. The species lacking the character include most of those with a paniculate inflorescence. Also common in the genus but not found in *Cottendorfia* or *Connellia* is a rich variation in the structure of the substomatal pore (Figures 72, 75, 102, 108, 153, 161, 204, 236, 264). The types are itemized in the morphological discussion. Distinctive trichomes (Figures 265, 271, 274) are found in one series of species, most of which have large rounded substomatal pores and vascular bundles covered adaxially by chlorenchyma. I have not seen such trichomes in other bromeliad genera. *Navia* is otherwise notable for a great variety of individual species or small species groups that apparently all represent diverse rather unrelated lines of development. The most distinctive species, *N. lopezii* (Figures 275–277), could not be assigned to the genus or the family on the basis of leaf anatomy alone, having an outer surface of extremely large epidermal cells, no scales, simple paracytic stomata, and a relatively undifferentiated mesophyll.

Vegetative Key to Species of *Navia*

1. Epidermal cells extremely large, up to 800 μ long, without separate subepidermal layers (Figures 275–277) *N. lopezii*
1. Epidermal cells small, usually near size of stomata, rarely over 50 μ long, subepidermal layers distinct.
 2. Leaves 50 cm or more long, at least 2 cm and usually 3 cm or more wide near the base (inflorescence paniculate).
 3. Substomatal openings large and rounded (Figure 68) *N. ramosa*
 3. Substomatal openings nearly closed by large thick-wall intrusions (Figures 75, 83).
 4. Vascular bundles separated from the abaxial epidermis by many rows of thin-walled cells (Figure 70).
 5. Adaxial leaf surface bearing many stomata (Figures 71–73) *N. reflexa*
 5. Adaxial leaf surface with few or no stomata *N. hechtioides*
 4. Vascular bundles connected or nearly connected to abaxial epidermis by a region of fibers (Figure 79) *N. brocchinioides*, *N. diffusa*, *N. gracilis*, *N. hohenbergioides*
 2. Leaves distinctly less than 50 cm long or less than 2 cm wide.
 6. Stomata clustered in pits, each cluster protected by a small rather conical scale (Figure 90) (inflorescence paniculate) *N. garcia-barrigae*
 6. Stomata not grouped in pits with protective scale.

Vegetative Key to Species of *Navia*—Continued

7. Leaves bearing distinct erect stiff trichomes, at least on the abaxial surface (Figures 265, 271, 274).
8. Leaves with only small cells abaxial to the vascular bundles (Figures 239, 242, 243); substomatal openings partially to almost completely closed (Figures 236, 238, 241).
9. Substomatal openings divided into many small pores (Figure 238) . *N. colorata*
9. Substomatal openings not completely divided or at most divided in two (Figures 236, 241).
10. Chlorenchyma covering the adaxial surface of the vascular bundles (Figure 232) *N. navicularis*, *N. breweri*
10. Larger vascular bundles exposed to adaxial water-storage tissue (Figures 242, 243) *N. pauciflora*, *N. steyermarkii*, *N. wurdackii*, *N. xyridiflora*
8. Leaves with large cells forming water-storage canals abaxial to the vascular bundles (Figures 253, 256, 261); substomatal openings large and rounded (Figures 252, 255, 258).
11. Larger vascular bundles exposed to water-storage tissue adaxially (Figure 253); leaves succulent in appearance *N. aloifolia*
11. Chlorenchyma covering adaxial surface of vascular bundles (Figures 256, 261); leaves usually rather thin.
12. Leaves with only vestigial marginal teeth.
13. Lower leaf margins densely pubescent *N. lanigera*
13. Lower leaf margin with only scattered hairs or teeth *N. myriantha*
12. Leaves with strongly serrate margins.
14. Leaf margins strongly reflexed *N. phelpisiae*
14. Leaf margins plane.
15. Leaves up to 2 cm wide, with long erect spinose marginal teeth *N. stenodonta*
15. Leaves 1 cm or less wide.
16. Leaves stiff, with prominent tufts of trichomes in axils of all the marginal teeth *N. trichodonta*
16. Leaves lax, without prominent tufts of trichomes in axils of the teeth *N. crispa*, *N. viridis*
7. Leaves with branching, platelike, or vestigial scales (Figures 58, 183, 184, 194).
17. Substomatal opening cruciate (Figure 102) or 4-lobed (Figure 104); anterior sepal of flower much reduced or aborted.
18. Substomatal opening cruciate (Figure 102); leaves with only small scales abaxially; anterior sepal aborted *N. parvula*
18. Substomatal opening with 4 broad lobes (Figure 104); leaves with large round scales abaxially; anterior sepal only reduced *N. lepidota*
17. Substomatal opening not cruciate or 4-lobed; anterior sepal not reduced or aborted.
19. Substomatal openings single, rather rounded, without obvious intrusions, usually large (Figures 186, 189, 192, 217, 220, 226).
20. Leaves with large cells abaxial to the vascular bundles (Figures 193, 218, 222, 227).
21. Abaxial subepidermal cells very thick-walled (Figure 217); large vascular bundles exposed to adaxial water-storage tissue (Figure 218) *N. mima*
21. Abaxial subepidermal cells with relatively thin walls (Figures 226, 229, 231); vascular bundles covered by chlorenchyma adaxially (Figures 221, 222, 227).
22. Leaves entire, narrowed in basal half; epidermal cells thin-walled, without silica bodies (Figure 95); inflorescence racemose *N. cataractarum*
22. Leaves at least slightly toothed, not narrowed below; epidermal cells with silica bodies (Figures 191, 225, 228); inflorescence not racemose.
23. Abaxial water-storage canals with one layer of very high vertical cells (Figure 193); leaf bearing prominent asymmetric scales (Figure 194); inflorescence long scapose *N. caulescens*
23. Abaxial water-storage canals with 2 or 3 layers of equal sized cells (Figures 222, 227); scales various; inflorescence not long scapose.

24. Leaves less than 5 cm long *N. schultesiana*
24. Leaves mostly over 5 cm long.
25. Leaves 1–3 mm wide; abaxial water-storage cells usually in 3 layers (Figure 222) *N. graminifolia*
25. Leaves 7–15 mm wide; abaxial water-storage cells usually in 2 layers (Figure 227) *N. acaulis*, *N. bicolor*, *N. heliophila*
20. Leaves with only small cells abaxial to the vascular bundles (Figures 99, 187, 190).
26. Leaves bearing large scales with small rather isodiametric cells (Figure 184); vascular bundles narrowly fused to abaxial epidermis (Figure 187) *N. arida*
26. Leaf scales vestigial or with elongate cells (Figure 183); vascular bundles variously shaped.
27. Vascular bundles broadly fused to abaxial epidermis (Figure 182) *N. serrulata*
27. Vascular bundles separated from the abaxial epidermis by distinct layers of small chlorenchymatous cells (Figures 99, 179, 190).
28. Adaxial water-storage tissue with 1 or 2 layers of cells (Figure 99); inflorescence paniculate *N. lindmanioides*
28. Adaxial water-storage tissue composed of 3 to many layers of large cells (Figures 179, 190); inflorescence glomerate, sessile.
29. Chlorenchyma continuous over adaxial surface of vascular bundles (Figure 190) *N. saxicola*
29. Larger vascular bundles exposed to adaxial water-storage tissue (Figure 179) *N. umbratilis*
19. Substomatal openings reduced, multiple, or with obvious intrusions (Figures 94, 126, 135, 153, 163, 173, 176, 214).
30. Substomatal opening multiple, with many separate scattered pores (Figures 94, 153).
31. Chlorenchyma covering adaxial surface of vascular bundles (Figure 92); inflorescence racemose *N. rupestris*
31. Vascular bundles mostly exposed to adaxial water-storage tissue (Figures 151, 156); inflorescence not racemose.
32. Stomata about 20 μ wide; subepidermal cells mostly 10–15 μ wide (Figure 152) *N. abysmophila*
32. Stomata about 25 μ wide; subepidermal cells mostly 15–20 μ wide (Figure 154) *N. duidae*
30. Substomatal opening not multiple, sometimes divided in the middle or completely obscured (Figures 126, 135, 163, 173, 176, 202, 214).
33. Substomatal opening large though sometimes divided in half, intrusions usually thin-walled (Figures 163, 173, 176, 202, 214).
34. Abaxial subepidermal cells and the projections into the substomatal cavity thick-walled (Figure 214) *N. pungens*
34. Abaxial subepidermal cells and the projections into the substomatal cavity rather thin-walled (Figures 163, 173, 176, 202).
35. Projections into the substomatal cavity multifid, bearing many papillae (Figures 163, 168).
36. Adaxial water-storage tissue composed of 3 or more layers of large cells, leaves thick (Figure 156) *N. brachyphylla*, *N. glauca*
36. Adaxial water-storage tissue composed of 1 or 2 layers of large cells (Figure 157); leaves thin, broad and flat.
37. Leaves narrowed near the base, not chalky below *N. nubicola*
37. Leaves not noticeably narrowed near base, often chalky on abaxial surface *N. cretacea*, *N. latifolia*
35. Projections into substomatal cavity usually paired, smooth (Figures 176, 202, 212).
38. Stomata as broad as long or broader (Figures 169, 172, 175); sepals separate.

Vegetative Key to Species of *Navia*—Continued

39. Leaves 3–5 mm wide; larger vascular bundles exposed to adaxial water-storage tissue (Figure 171); stomata about 20μ wide; distinct rounded scales present *N. semiserrata*
39. Leaves about 1 cm wide; adaxial surface of vascular bundles covered by chlorenchyma (Figure 174); stomata 25μ wide or more; scales much dissected or vestigial.
40. Leaf margins with teeth erect to antrorse *N. octopoides*
40. Leaf margins with teeth close-set, mostly retrorse *N. caurensis*
38. Stomata longer than broad (Figures 205, 207, 209, 211, 213); sepals connate.
41. Larger vascular bundles exposed to adaxial water-storage tissue (Figure 197); abaxial epidermal cells with extremely sinuous walls (Figure 195) *N. caricifolia*
41. Vascular bundles usually covered adaxially by chlorenchyma (Figure 200); walls of abaxial epidermal cells strongly to laxly sinuous (Figures 205, 207, 209, 211, 213).
42. Leaf margins entire or with mostly vestigial teeth *N. affinis*,
N. connata, *N. gleasonii*, *N. maguirei*, *N. subpetiolata*
42. Leaf margins sharply serrate.
43. Leaf blade very narrow, about 2.5 mm wide, with a short broad base *N. ocellata*
43. Leaf blade up to 1 cm or more wide, base not markedly wider *N. sandwithii*
33. Substomatal opening obscure, or reduced to a narrow slit or dumbbell-shaped pore by thick-walled intrusions (Figures 108, 126, 135).
44. Adaxial water-storage tissue composed of 1 or 2 layers of large cells (Figure 150) *N. angustifolia*
44. Adaxial water-storage tissue composed of many layers of large cells (Figures 139, 156).
45. Larger vascular bundles exposed to adaxial water-storage tissue (Figures 133, 136, 139).
46. Vascular bundles broadly fused to abaxial epidermis (Figure 136) *N. immersa*
46. Vascular bundles separated from lower epidermis by distinct layers of small chlorenchyma cells (Figure 139).
47. Stomata $40\text{--}50\mu$ long, about 30μ wide; walls of abaxial epidermal cells extremely sinuous (Figure 134) *N. intermedia*
47. Stomata $20\text{--}30\mu$ long; walls of abaxial epidermal cells strongly to weakly sinuous (Figures 140, 142).
48. Stomata as wide as long, about 30μ by 30μ (Figure 146). *N. scopulorum*
48. Stomata longer than wide, about 25μ long by 20μ wide (Figures 140, 142).
49. Leaves less than 5 cm long *N. pulvinata*
49. Leaves over 5 cm long *N. cucullata*
45. Vascular bundles usually covered adaxially by chlorenchyma (Figures 128, 132).
50. Substomatal opening completely obscured from face view (Figure 108); leaf with branching scales covering adaxial surface. *N. involuocrata*
50. Substomatal opening distinct as a narrow slit (Figures 121, 123, 126); scales on adaxial surface of leaf sparse or vestigial.
51. Abaxial leaf surface densely covered with branching scales.
52. Scales not completely obscuring abaxial leaf surface; leaf margins strongly incurved with sharp teeth; leaf interior without obvious resinous cells (Figure 117) *N. robinsonii*

- 52. Scales of abaxial leaf surface forming a dense woolly cover; leaves very stiff with erect margins and vestigial teeth; leaf interior with numerous resinous cells (Figures 127, 128, 131, 132).
- 53. Leaves broadly lanceolate, with many layers of narrow thick-walled resinous cells adaxially (Figure 131) *N. incrassata*
- 53. Leaves narrowly lanceolate, with 2 or 3 layers of rounded thick-walled resinous cells adaxially (Figure 127) *N. lasiantha*
- 51. Scales of abaxial leaf surface sparse or vestigial.
- 54. Leaves narrow with very large teeth at the base; ovary almost wholly inferior *N. fontoides*
- 54. Leaves without larger teeth at the base, ovary superior.
- 55. Leaves broad without broader base; marginal teeth very erect and close, about as long as the distance between them.
- 56. Abaxial subepidermal cells with very thick walls, lumens very small (Figure 111) *N. barbellata*
- 56. Abaxial subepidermal cells with rather thin walls (Figure 113) *N. splendens*
- 55. Leaves narrow with a very broad base; marginal teeth short, shorter than the distance between them.
- 57. Lower teeth of leaf margins with prominent tufts of trichomes in axils; interior of leaf with many layers of thick-walled subepidermal cells abaxially (Figure 124) *N. aurea*
- 57. Lower teeth of leaf margins without tufts of prominent trichomes; interior of leaf with only 1 or 2 layers of distinct subepidermal cells abaxially (Figure 117) *N. cardonae*

The genus *Navia* can be subdivided into the following 7 groups and 26 subgroups.

GROUP A: Inflorescence paniculate; vascular bundles not covered adaxially by chlorenchyma; occurring in the western and central Guayana Highlands of Colombia and southern Venezuela.

Subgroup I: One species with substomatal pore large, oval or rounded; vascular bundles free from abaxial epidermis.

The single species, *N. ramosa* (Figures 67–69), can be distinguished from *Cottendorfia* only by the unappended seeds.

Subgroup II: Six species with the substomatal pore reduced to a narrow double-anchor shape by pairs of hooked thick-walled intrusions.

In habit, the six species are like the preceding *N. ramosa* and like many species of *Cottendorfia*, but they can be distinguished by the shape of the substomatal pore. As presently known, *N. reflexa* (Figures 70–76) and *N. hechtioides* (Figures 77–78) have free vascular bundles while those of *N. hohenbergioides* (Figures 79–81), *N. gracilis* (Figures 82–83), *N. brocchiniooides* (Figures 84–85), and *N. diffusa* (Figures 86–87) are connected to the lower epidermis by areas

of sclerenchyma. Stomata seem rather common on the adaxial leaf surface of *N. reflexa* where the surrounding cells of the third subepidermal layer show a rather distinctive structure (Figure 73). A few adaxial stomata have been observed in *N. gracilis*, but they have not been observed in any other bromeliads examined.

Subgroup III: One species with stomata clustered in pits around thick pluglike scales; substomatal pores irregularly rounded to partially closed by short thick-walled intrusions.

The arrangement of stomata in the single species, *N. garcia-barrigae* (Figures 88–91), seems distinctive. The single specimen available has only a few separate leaves, which may not be representative of the fully vegetative parts of the plant.

GROUP B: Inflorescence racemose to paniculate; vascular bundles nearly or completely covered adaxially by chlorenchyma.

Subgroup IV: Two small species with racemose inflorescence; leaves rather thick; silica bodies small or lacking; occurring in the eastern Guayana Highlands in British Guiana.

In *N. rupestris* (Figures 92–94) the substomatal openings are usually divided into three or more small rounded pores. In *N. cataractarum* (Figures 95–96) the pores are large and simple.

Subgroup V: One rather large species with large panicle bearing racemose branches; leaves thin; silica bodies distinct; occurring in central Venezuela.

Navia lindmanioides (Figures 97–99) has simple rounded substomatal pores. The species is quite distinct from others with paniculate or racemose inflorescences, and vegetatively resembles many having glomerate inflorescences.

GROUP C: Inflorescence glomerate; vascular bundles covered adaxially by chlorenchyma; anterior sepal reduced or absent; scales platelike or branching.

Subgroup VI: Two species with substomatal pore cruciate or 4-lobed; ovary partly inferior; occurring on Cerro Neblina in southern Venezuela.

Navia parvula (Figures 100–102) shows the extreme conditions of the anterior sepal absent and the substomatal pore cruciate. In *N. lepidota* (Figures 103–106) the anterior sepal is present in reduced form and the substomatal pore has only slight intrusions from each of the four corners. In both mentioned characters the latter species seems transitional, but the species is very distinct in other characters such as its large rounded scales.

GROUP D: Inflorescence glomerate; sepals equal, free; scales platelike, branching, or vestigial.

Subgroup VII: One species with substomatal opening completely obscured from face view; vascular bundles covered adaxially by chlorenchyma; occurring in the Guayana Highlands of central Venezuela.

The intrusions into the substomatal pore of *N. involucrata* (Figures 107–108, section as in 106) are so complex and thick-walled that it is impossible to determine any opening or any pattern. The broad involute leaves with a grayish cover of adaxial scales are also rather distinctive.

Subgroup VIII: One species with substomatal pore reduced to a short narrow slit; vascular bundles covered adaxially by chlorenchyma; leaves with a few very large teeth near the base; ovary

almost wholly inferior; occurring in the Guayana Highlands of Colombia.

Navia fontoides (Figures 114–116) seems readily distinguishable by its narrow leaves bearing large teeth near the base. The species also seems rather distinct in its inferior ovary and in its geographical distribution.

Subgroup IX: Two species with substomatal pore reduced to a short narrow slit; vascular bundles covered adaxially by chlorenchyma; leaves broad and smooth with closely and finely serrulate margins; ovary superior; occurring in the eastern Guayana Highlands of British Guiana and adjacent Venezuela.

Navia barbellata (Figures 109–111) and *N. splendens* (Figures 112–113, section as in 109) are similar in most vegetative characters. The material seen of *N. barbellata* has much thicker walled subepidermal cells.

Subgroup X: Five species with substomatal pore reduced to a short narrow slit; vascular bundles covered adaxially by chlorenchyma; leaves mostly with evident scales on abaxial surface; ovary superior; occurring in the Jáua-Duida area of south-central Venezuela.

Navia lasiantha (Figures 125–128) and *N. incrassata* (Figures 129–132) both have nearly entire-margined leaves with a very dense abaxial cover of scales. The three remaining species of the group have less prominent scales. Of these three, *N. robinsonii* (Figures 120–121, section as in 117) has broader involute leaves while *N. cardonae* (Figures 117–119) and *N. aurea* (Figures 122–124) both have narrow blades born on broad sheathing bases. *Navia aurea* is quite distinct in the many layers of thick-walled convoluted cells adjacent to the lower epidermis.

Subgroup XI: One species with substomatal pore reduced to a pair of small hidden pores; epidermal cells large with very sinuous walls; vascular bundles not all covered adaxially by chlorenchyma; ovary superior; occurring on Serranía Jáua in south-central Venezuela.

The epidermal cells of *N. intermedia* (Figures 133–135) are the largest in the genus that are of normal structure. The substomatal structure is also rather distinctive, but is very closely related to that in the immediately preceding and following groups. The leaves are very narrow but thick and nearly entire-margined, unlike any others seen in the genus.

Subgroup XII: Five species with substomatal pore reduced to a short narrow slit; vascular bundles not completely covered by chlorenchyma adaxially; occurring in the Guayana Highlands of British Guiana and Venezuela.

Three of the species, *N. cucullata* (Figures 139–141), *N. pulvinata* (Figures 142–144), and *N. angustifolia* (Figures 148–150), have superior ovaries. Two, *N. scopulorum* (Figures 145–147) and *N. immersa* (Figures 136–138), have partially inferior ovaries. *Navia immersa* is further distinguished by the vascular bundles being broadly fused to the lower epidermis. The subgroup represents the greatest concentration of the leaf type in which the blade is very narrow and stiff from a broad rounded sheathing base. Such leaves are also seen in *N. cardonae* and *N. aurea* of Subgroup X, *N. saxicola* of Subgroup XVI, and *N. graminifolia* of Subgroup XX.

Subgroup XIII: Two species with the substomatal opening divided into few or many small rounded pores; vascular bundles not completely covered adaxially by chlorenchyma; occurring in southern Venezuela.

The two species, *N. abyssophila* (Figures 151–153) and *N. duidae* (Figures 154–156), have slightly different habits but very similar basic anatomy. The epidermal cells are among the smallest seen in the genus.

Subgroup XIV: Five species with substomatal intrusions short but distinct, thin-walled, much divided or bearing many papillae; vascular bundles usually covered adaxially by chlorenchyma; occurring in the Duida-Neblina area of southern Venezuela.

Two of the species, *N. brachyphylla* (Figures 165–166, section as in 156) and *N. glauca* (Figures 167–168, section as in 156), have rather thick leaves. In *N. nubicola* (Figures 157–159), *N. cretacea* (Figures 160–161, section as in 157), and *N. latifolia* (Figures 162–164, section as in 157) the leaves are very thin. *Navia cretacea* and *N. latifolia* are obviously closely related and cannot be distinguished vegetatively.

Subgroup XV: Three species with substomatal intrusions rather short but distinct, thin-walled, simple; vascular bundles partially to completely covered adaxially by chlorenchyma; occurring in

the Rio Caura-Rio Ventuari-Duida area of south-central Venezuela.

The subgroup seems unnatural with species differing somewhat in both habit and anatomy. *Navia semiserata* (Figures 169–171) differs by its stiff leaves and vascular bundles not completely covered by chlorenchyma. In *N. caurensis* (Figures 172–174) and *N. octopoides* (Figures 175–176, section as in 157) the vascular bundles are covered by chlorenchyma adaxially, but the former has broad relatively short thick leaves and the latter has much longer and thinner leaves.

Subgroup XVI: Five species with substomatal pores rounded or oval, without distinct intrusions; occurring in various parts of the Guayana Highlands from British Guiana to Colombia.

The five species included in the subgroup are sufficiently distinct that each could constitute a separate subgroup of its own. *Navia umbratilis* (Figures 177–179), of Cerro Yutaje in south-central Venezuela, has long slender nearly entire-margined leaf blades from a rounded sheathing base. The substomatal pores are slightly narrowed, and the larger vascular bundles are not completely covered adaxially by chlorenchyma. The substomatal pores of the other species are not at all obstructed. In both *N. serrulata* (Figures 180–183), of southern Venezuela, and *N. arida* (Figures 184–187), of British Guiana, the vascular bundles are not completely covered with chlorenchyma. In the former species the leaves are narrow from a broad base, but are strongly serrulate, and the scales have oblong cells. In the latter species the leaves are broad and strap-shaped, the scales are rounded with very distinctive small rounded cells and an eroded margin, and the ovary, unlike other species in the subgroup, is partly to wholly inferior. In both *N. saxicola* (Figures 188–190), of southern Venezuela, and *N. caulescens* (Figures 191–194), of Colombia, the vascular bundles are completely covered adaxially by chlorenchyma. The former species has stiff narrow leaf blades from rounded sheathing bases, while the latter has broad strap-shaped leaves. *Navia caulescens*, the type-species of the genus, is most distinct by the elongate inflorescence with several moniliform heads born on a scape.

GROUP E: Inflorescence glomerate; sepals connate; scales platelike, branching, or vestigial.

Subgroup XVII: Eight species with substomatal pores partly closed by long, paired, thin-walled intrusions; vascular bundles covered adaxially by

chlorenchyma in all but one species; abaxial water-storage canals absent; occurring in the eastern half of the Guayana Highlands from Cerro Sipapo and Neblina in Venezuela to Tafelberg in Surinam.

One species, *N. caricifolia* (Figures 195–197), is distinct in the vascular bundles not being covered adaxially by chlorenchyma, and in the more sinuous walls of the epidermal cells. In other characters such as the long strap-shaped usually nearly entire leaves having somewhat sheathing bases and the structure of the substomatal pore, *N. caricifolia* is like *N. ocellata* (Figures 198–200), *N. maguirei* (Figures 201–202, section as in 200), *N. subpetiolata* (Figures 203–204, section as in 200), *N. gleasonii* (Figures 205–206, section as in 200), *N. sandwithii* (Figures 207–208, section as in 200), *N. connata* (Figures 209–210, section as in 200), and *N. affinis* (Figures 211–212, section as in 200).

Subgroup XVIII: One species with substomatal pores partly closed by short, paired, thick-walled intrusions; vascular bundles covered adaxially by chlorenchyma, abaxial water-storage canals absent; occurring on Cerro Guanay and Cerro Yutaje in south-central Venezuela.

The single species, *N. pungens* (Figures 213–215), is distinct among *Navia* species with a sessile inflorescence by the small but distinct thick-walled intrusions. The leaves are thick and stiff, and the plant closely resembles *N. mima* of the following subgroup in habit.

Subgroup XIX: One species with substomatal pore only slightly obstructed; vascular bundles not completely covered adaxially by chlorenchyma; with distinct large cells forming water-storage canals abaxially; occurring on Cerro Sipapo in south-central Venezuela.

The one species, *N. mima* (Figures 216–218), resembles the preceding, *N. pungens*, in habit, but it has the substomatal pores only slightly obstructed by the surrounding thin-walled resinous cells.

Subgroup XX: Five species with substomatal pore rounded, not obstructed; vascular bundles completely covered adaxially by chlorenchyma; distinct large cells forming abaxial water-storage canals; occurring in the western Guayana Highlands in Colombia.

The five species, *N. acaulis* (Figures 230–231, section as in 227), *N. bicolor* (Figures 228–229, section

as in 227), *N. heliophila* (Figures 225–227), *N. graminifolia* (Figures 222–224), and *N. schultesiana* (Figures 219–221), all share the same basic anatomy and the same rather distinctive geographic distribution. Still, the habits of the various species in the subgroup are quite diverse. The first three species are much alike with long, broad, strap-shaped leaves having no distinct central thickening. These three species cannot be distinguished vegetatively. *Navia graminifolia* in contrast has the very different slender-leaved form found most often in Subgroup XII. *Navia schultesiana* has distinctive small rosettes of serrate stiffly spreading leaves, bearing little superficial resemblance to the preceding types. In this study of *Navia*, most attempts to use vegetative habit as an indication of relationship have failed. Most subgroups established on the basis of anatomy have contained many variations in habit. In this subgroup there is some basis to suspect real relationship and therefore potentially good evidence of the erratic nature of the vegetative habit.

GROUP F: Inflorescence glomerate; sepals equal, separate; scales lacking, large erect trichomes present.

Subgroup XXI: Two species with substomatal pores reduced to a short narrow slit; vascular bundles covered adaxially by chlorenchyma; without distinct water-storage canals abaxially; ovary superior; occurring in the Jáua-Ichún area of south-central Venezuela.

The two species, *N. navicularis* (Figures 232–234) and *N. breweri* (Figures 235–236, section as in 232), seem indistinguishable vegetatively. They and the five species in the following two subgroups, and *N. viridis* and *N. crispa* of Subgroup XXV, show a long, thin, strap-shaped leaf often with a distinct central thickening. While this rather distinctive leaf type is most common among species with trichomes, it is also found in *N. octopoides* of Subgroup XV and *N. lindmanioides* of Subgroup V. Some species like *N. arida* of Subgroup XVI and *N. nubicola* of Subgroup XIV are similar but lack the distinct central thickening.

Subgroup XXII: One species with substomatal opening divided into many small usually rounded pores; with few or no distinct water-storage cells abaxially; vascular bundles covered adaxially by chlorenchyma; ovary superior; occurring in the Rio Ventuari area of south-central Venezuela.

The species, *N. colorata* (Figures 237–239), agrees in habit and in most anatomical characters with the species of the preceding subgroup, but it differs markedly in the thin-walled substomatal cells which form multiple openings. This species is probably most closely related to *N. viridis* and *N. crispa* of Subgroup XXV.

Subgroup XXIII: Four species with substomatal opening obstructed by distinct, long, thin-walled, paired intrusions; vascular bundles not completely covered adaxially by chlorenchyma; with few distinct water-storage cells abaxially; ovary superior; occurring in the Chimanta-Marahuaca-Duida areas of eastern and south-central Venezuela.

The four species, *N. pauciflora* (Figures 247–248, section as in 243), *N. steyermarkii* (Figures 249–250, section as in 243), *N. wurdackii* (Figures 243–246), and *N. xyridiflora* (Figures 240–242), seem indistinguishable vegetatively. The failure of the chlorenchyma to completely cover the adaxial surface of the vascular bundles in this series of species seems to be of relatively little significance, since the organization of the tissues is similar to that in the related subgroups that have covered bundles.

Subgroup XXIV: One species with substomatal pore large and unobstructed; vascular bundles mostly not covered adaxially by chlorenchyma; large cells forming distinct water-storage canals abaxially; ovary inferior; occurring on Cerro Neblina in southern Venezuela.

The species, *N. aloifolia* (Figures 251–253), seems very distinct in the habit, thick leaves, vascular bundles distinctly exposed to the adaxial water-storage tissue, and the inferior ovary. The trichomes on the backs of the leaves are perhaps the best examples of such structures in the genus.

Subgroup XXV: Seven species with substomatal pores large and unobstructed; vascular bundles covered adaxially by chlorenchyma; large cells usually forming distinct water-storage canals abaxially; ovary superior; occurring in south-central and southern Venezuela and adjacent Brazil.

Two species, *N. viridis* (Figures 254–256) and *N. crispa* (Figures 257–258, section as in 256), have the appearance of the species in Subgroups XXI, XXII, and XXIII, but in addition to differences in stomata they show a greater development of water-storage cells abaxially. The remaining five species, *N. stenodonta* (Figures 259–262), *N. myriantha* (Figures 263–266), *N. trichodonta* (Figures 267–268, section as in 266), *N. lanigera* (Figures 269–271), and *N. phelpsi* (Figures 272–274), have rather diverse leaf types, but all have rather well-developed water-storage canals abaxially.

GROUP G: Inflorescence glomerate; sepals equal and separate; scales completely absent from leaves.

Subgroup XXVI: One species with broad entire-margined leaves; epidermal cells extremely large, subepidermal layers absent; stomata paracytic; mesophyll not forming distinct layers; occurring in the central and western Guayana Highlands of southern Venezuela and adjacent Brazil and Colombia.

All the most distinctive features of *N. lopezii* (Figures 275–277) could be related to the lack of subdivision of initial cells. This is most pronounced in the epidermal cells of the leaves and floral bracts, but is also somewhat reflected in the structure of the walls of the fruits. The modified epidermis has resulted in complete elimination of scales, a condition that is in exception to the generally accepted definition of the family.

List of Specimens Examined

(Specimens used for illustrations are marked with an asterisk.)

Species	Collector and Number	Locality	Herbarium
<i>Abromeitiella brevifolia</i> (Griseb.) Castellanos	Cabrera 16333	Argentina: Prov. Jujuy: Tumbaya	US
<i>Ayensua uaiapanensis</i> (Maguire) L. B. Smith	Steyermark 93940	Venezuela: Bolívar: Auyán-tepuf	US
<i>Brocchinia acuminata</i> L. B. Smith	Maguire 32785	Venezuela: Bolívar: Cerro Guai-quinima	US
<i>Brocchinia cowanii</i> L. B. Smith	Maguire, Cowan, & Wurdack 30924	Venezuela: Amazonas: Cerro Moriche, Río Ventuari	US holotype

List of Specimens Examined—Continued

<i>Species</i>	<i>Collector and Number</i>	<i>Locality</i>	<i>Herbarium</i>
<i>Brocchinia cryptantha</i> L. B. Smith	Maguire, Cowan & Wurdack 30709	Venezuela: Amazonas: Cerro Yapacana, Río Orinoco	US holotype
<i>Brocchinia delicatula</i> L. B. Smith	Maguire, Wurdack & Maguire 42207	Venezuela: Amazonas: Cerro Neblina, Río Yatua	US holotype
<i>Brocchinia hechtiioides</i> Mez	Maguire & Politi 27603	Venezuela: Amazonas: Cerro Sipapo	US
<i>Brocchinia hitchcockii</i> L. B. Smith	Phelps & Hitchcock 522	Venezuela: Amazonas: Serranía Parú, Río Ventuari	NY holotype
<i>Brocchinia maguirei</i> L. B. Smith	Maguire & Politi 27950	Venezuela: Amazonas: Cerro Sipapo	US
<i>Brocchinia melanaera</i> L. B. Smith	Maguire, Phelps et al. 31691	Venezuela: Amazonas: Río Manapiare, Río Ventuari	US
<i>Brocchinia micrantha</i> (Baker) Mez	Steyermark 89564	Venezuela: Bolívar: Sierra de Lema, Cabeceras de Río Chicanán	US
<i>Brocchinia paniculata</i> Schult. f.	Schultes & Cabrera 17131	Colombia: Vaupés: Río Piraparaná	US
<i>Brocchinia prismatica</i> L. B. Smith	Maguire, Cowan, & Wurdack 30596	Venezuela: Amazonas: Cerro Yapacana, Río Orinoco	US
<i>Brocchinia reducta</i> Baker	Maguire 33185	Venezuela: Bolívar: Ilu-tepuí	US
<i>Brocchinia secunda</i> L. B. Smith	Steyermark 59863	Venezuela: Bolívar: Ptari-tepuí	F isotype
<i>Brocchinia serrata</i> L. B. Smith	García Barriga 15096	Colombia: Vaupés: Río Kubiyú	US
<i>Brocchinia steyermarkii</i> L. B. Smith	Maguire, Tillett, & Tillett 43808	British Guiana: Upper Mazaruni	US
<i>Brocchinia tatei</i> L. B. Smith	Maguire 32874	Venezuela: Bolívar: Cerro Guaiquinima, Río Paragua	US
<i>Brocchinia vestita</i> L. B. Smith	Maguire, Wurdack, etc. 42168	Venezuela: Amazonas: Cerro Neblina, Río Yatua	US
<i>Connellia angustae</i> (Rich. & Schombrock) N. E. Brown	Steyermark & Nilsson 745	Venezuela: Bolívar: Cerro Venamo	US
<i>Connellia caricifolia</i> L. B. Smith	Steyermark 58846	Venezuela: Bolívar: Mt. Roraima	F holotype
<i>Connellia nutans</i> L. B. Smith	Steyermark 59933	Venezuela: Bolívar: Ptari-tepuí	F holotype
<i>Connellia quelchii</i> N. E. Brown	*Im Thurn 315	Venezuela: Bolívar: Mt. Roraima	US isotype
<i>Cottendorfia argentea</i> (L. B. Smith) L. B. Smith	*Steyermark & Wurdack 677	Venezuela: Bolívar: Chimantá-tepuí	US holotype
<i>Cottendorfia brachyphylla</i> (L. B. Smith) L. B. Smith	*Wurdack 34134-B	Venezuela: Bolívar: Sarvén-tepuí	US holotype
<i>Cottendorfia cylindrostachya</i> (L. B. Smith) L. B. Smith	*Maguire 35280	Venezuela: Amazonas: Serranía Yutaje, Río Manapiare	US holotype
<i>Cottendorfia dendritica</i> L. B. Smith	*Maguire, Pires, & Maguire 60543	Brazil: Amazonas: Serra da Neblina	NY holotype
<i>Cottendorfia dyckioides</i> L. B. Smith	Steyermark 93186	Venezuela: Bolívar: Auyán-tepuí	US
	*Steyermark 93188	Venezuela: Bolívar: Auyán-tepuí	US holotype
<i>Cottendorfia florida</i> Schultes f.	*Martius 1938	Brazil: Bahia: Serra do Sincorá	M holotype
<i>Cottendorfia geniculata</i> (L. B. Smith) L. B. Smith	*Steyermark 75774	Venezuela: Bolívar: Chimantá-tepuí	US holotype
<i>Cottendorfia gracillima</i> L. B. Smith	*Steyermark & Nilsson 436	Venezuela: Bolívar: Cerro Venamo	US holotype
<i>Cottendorfia guianensis</i> (Beer) Klotzsch ex Baker var. <i>vestita</i> L. B. Smith	*Steyermark & Nilsson 567	Venezuela: Bolívar: Uarama-tepuí	US holotype
<i>Cottendorfia longipes</i> L. B. Smith	*Steyermark 93790	Venezuela: Bolívar: Auyán-tepuí	US holotype
<i>Cottendorfia maguirei</i> L. B. Smith	*Maguire, Pires, & Maguire 60478	Brazil: Amazonas: Serra da Neblina	US holotype
<i>Cottendorfia minor</i> (L. B. Smith) L. B. Smith	*Steyermark & Wurdack 678-A	Venezuela: Bolívar: Chimantá-tepuí	US
<i>Cottendorfia naviooides</i> (L. B. Smith) L. B. Smith	*Steyermark & Wurdack 677	Venezuela: Bolívar: Chimantá-tepuí	US holotype
<i>Cottendorfia nubigena</i> L. B. Smith	*Maguire, Wurdack, & Maguire 42151	Venezuela: Amazonas: Cerro de la Neblina, Río Yatua	US holotype
<i>Cottendorfia paludosa</i> (L. B. Smith) L. B. Smith	*Maguire 33034	Venezuela: Bolívar: Cerro Guaiquinima Río Paragua	US holotype

List of Specimens Examined—Continued

Species	Collector and Number	Locality	Herbarium
<i>Cottendorfia phelpsiae</i> (L. B. Smith) L. B. Smith	*Phelps & Hitchcock 501	Venezuela: Bolívar: Cerro Parú	NY holotype
<i>Cottendorfia savannensis</i> L. B. Smith	*Maguire, Wurdack, & Maguire 42167	Venezuela: Bolívar: Cerro de la Neblina, Río Yatua	US holotype
<i>Cottendorfia serrulata</i> (L. B. Smith) L. B. Smith	Cardona 1587	Venezuela: Bolívar: Cerro Apacara, Río Caroní	US holotype
<i>Cottendorfia serrulata</i> var. <i>reducta</i> (L. B. Smith) L. B. Smith	*Steiermark & Wurdack 1167	Venezuela: Bolívar: Chimantá-tepuí	US holotype
<i>Cottendorfia stenophylla</i> (L. B. Smith) L. B. Smith	*Steiermark & Wurdack 951	Venezuela: Bolívar: Chimantá- tepuí	US holotype
<i>Cottendorfia steiermarkii</i> (L. B. Smith) L. B. Smith	Steiermark & Wurdack 827	Venezuela: Bolívar: Chimantá Massif	NY holotype
<i>Cottendorfia subsimplex</i> (L. B. Smith) L. B. Smith	*Steiermark 75924	Venezuela: Bolívar: Chimantá Massif	US holotype
<i>Cottendorfia thyrsoidea</i> (L. B. Smith) L. B. Smith	Maguire & Maguire 35385	Venezuela: Amazonas: Serranía Yutaje	NY holotype
<i>Cottendorfia tillandsioides</i> (L. B. Smith) L. B. Smith	*Steiermark 74863	Venezuela: Bolívar: Abácapa- tepuí	US holotype
<i>Cottendorfia wurdackii</i> (L. B. Smith) L. B. Smith	*Maguire, Cowan, & Wurdack 29637	Venezuela: Amazonas: Cerro Duida	US holotype
<i>Deuterocohnia longipetala</i> (Baker) Mez	Castellanos 26/448	Argentina: San Juan: Quebrada del Zonda	US
<i>Dyckia brevifolia</i> Baker	Reitz 3979	Brazil: Santa Catarina: Salto Blumenau	US
<i>Dyckia choristaminea</i> Mez	Rambos s. n. (PACA 48832)	Brazil: Rio Grande do Sul: Pôrto Alegre	US
<i>Dyckia duckei</i> L. B. Smith	Eiten & Eiten 4659	Brazil: Maranhão: Mun. de Lorêto	US
<i>Dyckia maritima</i> Baker	L. B. Smith & Reitz 5824	Brazil: Rio Grande do Sul: Mun. Tórres	US
<i>Dyckia reitzii</i> L. B. Smith	Reitz & Klein 2690	Brazil: Santa Catarina: Serra do Oratório S. Joaquim	US holotype
<i>Encholirium densiflorum</i> Ule	Foster 2474	Brazil: Bahia: Amargosa, Milagres	US
<i>Fosterella penduliflora</i> (C. H. Wright) Stapf.	Venturi 5474	Argentina: Jujuy: Sierra de Santa Barbara	US
<i>Fosterella schidosperma</i> (Baker) L. B. Smith	Killip & Smith 25326	Peru: Junín: Río Paucartambo Valley	US
<i>Hechtia integerrima</i> M. B. Foster	Foster 3072	Mexico? cultivated	US holotype
<i>Hechtia lundelliorum</i> L. B. Smith	Van Hying 6066	Mexico: San Luis Potosí Tamazunchale	US
<i>Hechtia montana</i> Brand.	Ferris 8747	Mexico: Sonora: San Carlos Bay, near Guayamas	US
<i>Hechtia tillandsioides</i> (Andrè) L. B. Smith	Foster June 1950	Mexico: cultivated	US
<i>Navia abysmophila</i> L. B. Smith	*Maguire, Wurdack, & Maguire 42471	Venezuela: Amazonas: Cerro de la Neblina, Río Yatua	US holotype
<i>Navia acaulis</i> Martius ex Schultes f.	*Idrobo & Schultes 644	Colombia: Vaupés: Mesa la Lindosa, Ajaju River	US
<i>Navia affinis</i> L. B. Smith	*Maguire, Wurdack, & Maguire 42470	Venezuela: Amazonas: Cerro de la Neblina, Río Yatua	US holotype
<i>Navia aloifolia</i> L. B. Smith	*Maguire, Wurdack, & Bunting 37146	Venezuela: Amazonas: Cerro de la Neblina, Río Yatua	US holotype
<i>Navia angustifolia</i> (Baker) Mez	*Appun 1055	British Guiana: Marima	K holotype
<i>Navia arida</i> L. B. Smith & Steiermark	*Steiermark 89698	Venezuela: Bolívar: Sierra de Lema	US holotype
<i>Navia aurea</i> L. B. Smith	*Steiermark 58219	Venezuela: Amazonas: Cerro Duida	F holotype
<i>Navia barbellata</i> L. B. Smith	*Tillett 45557	British Guiana: Essequibo, Karowtupo, Mazaruni River	US isotype

List of Specimens Examined—Continued

Species	Collector and Number	Locality	Herbarium
<i>Navia bicolor</i> L. B. Smith	*Schultes 5444	Colombia: Vaupés: Cerro Chiribiquete, Río Macaya	US
<i>Navia brachyphylla</i> L. B. Smith	*Tate 571	Venezuela: Amazonas: Cerro Duida	NY holotype
<i>Navia breweri</i> L. B. Smith & Steyermark	*Brewer 248	Venezuela: Bolívar: Cerro Jáua, Río Caura	US holotype
<i>Navia brocchinioides</i> L. B. Smith	*Maguire & Politi 28787	Venezuela: Amazonas: Cerro Sipapo	US holotype
<i>Navia cardonae</i> L. B. Smith	Cardona 349	Venezuela: Bolívar: Río Canaracuni	US holotype fragment
	*Brewer 249	Venezuela: Bolívar: Cerro Jáua, Río Caura	US
<i>Navia caricifolia</i> L. B. Smith	*Maguire & Maguire 35437	Venezuela: Amazonas: Serranía Yutaje, Río Manapiare	US holotype
<i>Navia cataractarum</i> Sandwith	*Whitton 51	British Guiana: Essequibo, Amatum Falls, Potaro River	K holotype
<i>Navia caulescens</i> Martius ex Schultes f.	*Schultes 5859	Colombia: Amazonas: Cerro de Cupatí, Río Caqueta	US
<i>Navia caurensis</i> L. B. Smith	*Cardona 2985	Venezuela: Bolívar: Cerro Pauo, Río Caura	US holotype
<i>Navia colorata</i> L. B. Smith	*Cowan & Wurdack 31417	Venezuela: Amazonas: Serranía Parú, Río Ventuari	US holotype
<i>Navia connata</i> L. B. Smith & Steyermark	*Steyermark 90223	Venezuela: Bolívar: Sierra Ichún	US isotype
<i>Navia cretacea</i> L. B. Smith	*Maguire, Cowan, & Wurdack 29882	Venezuela: Amazonas: Cerro Huachamacari	US holotype
<i>Navia crispa</i> L. B. Smith	*Maguire, Cowan, & Wurdack 30966	Venezuela: Amazonas: Cerro Moriche, Río Ventuari	US holotype
<i>Navia cucullata</i> L. B. Smith	*Maguire 32766	Venezuela: Bolívar: Cerro Guaiquinima, Río Paragua	US holotype
<i>Navia diffusa</i> L. B. Smith	*Maguire, Wurdack, & Maguire 42537-E	Venezuela: Amazonas: Cerro de la Neblina, Río Yatua	US holotype
<i>Navia duidae</i> L. B. Smith	*Tate 404	Venezuela: Amazonas: Cerro Duida	NY holotype
<i>Navia fontoides</i> L. B. Smith	*Schultes & Cabrera 15391	Colombia: Amazonas: Rauda Yayacopi, Río Apaporis	US holotype
<i>Navia garcia-barrigae</i> L. B. Smith	*Garcia Barriga & Schultes 14137	Colombia: Amazonas: Araracuara, Río Caqueta	US holotype
<i>Navia glauca</i> L. B. Smith	*Steyermark 58320	Venezuela: Amazonas: Cerro Duida	F holotype
<i>Navia gleasonii</i> L. B. Smith	*Maguire & Fanshawe 32389	British Guiana: Essequibo	US
<i>Navia gracilis</i> L. B. Smith	*Wurdack & Adderley 43275	Venezuela: Amazonas: Maroa, Río Guainia	US holotype
<i>Navia graminifolia</i> L. B. Smith	*Schultes 5492	Colombia: Vaupés: Cerro Chiribiquete, Río Macaya	US isotype
<i>Navia hechtioides</i> L. B. Smith	*Maguire & Politi 27574	Venezuela: Amazonas: Cerro Sipapo	US holotype
<i>Navia heliophila</i> L. B. Smith	*Schultes 12085	Colombia: Amazonas: Raudel de Jirijirimo, Río Apaporis	US holotype
<i>Navia hohenbergioides</i> L. B. Smith	*Maguire Cowan, & Wurdack 30970	Venezuela: Amazonas: Cerro Moriche, Río Ventuari	US holotype
<i>Navia immersa</i> L. B. Smith	*Maguire, Cowan, & Wurdack 30325	Venezuela: Amazonas: Cerro Huachamacari, Río Cunucunuma	US holotype
<i>Navia incrassata</i> L. B. Smith & Steyermark	*Steyermark 97881	Venezuela: Bolívar: Meseta de Jáua, Río Caura	US holotype
<i>Navia intermedia</i> L. B. Smith & Steyermark	*Steyermark 97865	Venezuela: Bolívar: Cerro Jáua, Río Caura	US holotype
<i>Navia involucreta</i> L. B. Smith	*Cowan & Wurdack 31359	Venezuela: Amazonas: Serranía Parú, Río Ventuari	US holotype
<i>Navia lanigera</i> L. B. Smith	*Maguire et al. 31794	Venezuela: Amazonas: Cerro Camani, Río Ventuari	US holotype

List of Specimens Examined—Continued

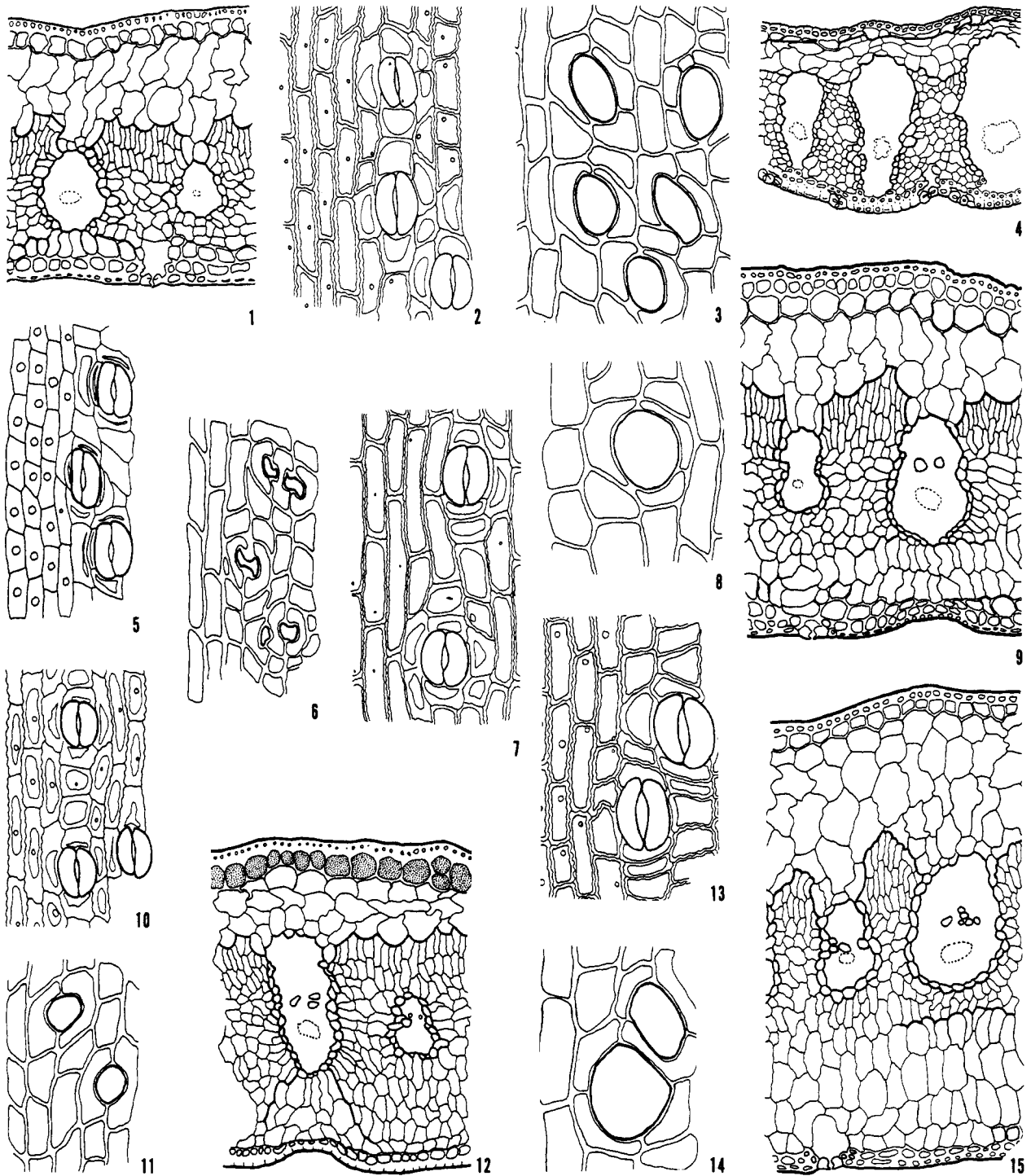
Species	Collector and Number	Locality	Herbarium
<i>Navia lasiantha</i> L. B. Smith & Steyermark	*Steyermark 97865-A	Venezuela: Bolívar: Cerro Jáua, Rfo Caura	US holotype
<i>Navia latifolia</i> L. B. Smith	*Maguire, Cowan, & Wurdack 29642	Venezuela: Amazonas: Cerro Duida	US holotype
<i>Navia lepidota</i> L. B. Smith	*Maguire, Wurdack, & Bunting 37288	Venezuela: Amazonas: Cerro de la Neblina, Rfo Yatua	US holotype
<i>Navia lindmanioides</i> L. B. Smith	*Maguire, Cowan, & Wurdack 30650	Venezuela: Amazonas: Cerro Yapacana, Rfo Orinoco	US holotype
<i>Navia lopezii</i> L. B. Smith	*Schultes & Lopez 9956 Maguire & Politi 27497	Brazil: Amazonas: Serra Dimiti Venezuela: Amazonas: Cerro Sipapo	US holotype US
<i>Navia maguirei</i> L. B. Smith	Wurdack & Adderly 43564 *Maguire 24575	Venezuela: Amazonas: Rfo Siapa Surinam: Tafelberg	US US isotype
<i>Navia maguirei</i> var. <i>minor</i> L. B. Smith	*Maguire 24334	Surinam: Tafelberg	US isotype
<i>Navia mima</i> L. B. Smith	*Maguire & Politi 27590	Venezuela: Amazonas: Cerro Sipapo	US holotype
<i>Navia myriantha</i> L. B. Smith	*Schultes & Lopez 9955	Brazil: Amazonas: Serra Dimiti	US holotype
<i>Navia navicularis</i> L. B. Smith & Steyermark	*Steyermark 90199	Venezuela: Bolívar: Sierra Ichún	US isotype
<i>Navia nubicola</i> L. B. Smith	*Maguire, Wurdack, & Bunting 37392	Venezuela: Amazonas: Cerro de la Neblina, Rfo Yatua	US holotype
<i>Navia ocellata</i> L. B. Smith	*Maguire & Politi 27500	Venezuela: Amazonas: Cerro Sipapo	US holotype
<i>Navia octopoides</i> L. B. Smith	*Maguire & Maguire 29151	Venezuela: Amazonas: Cerro Duida	US holotype
<i>Navia parvula</i> L. B. Smith	*Maguire, Wurdack, & Bunting 37326	Venezuela: Amazonas: Cerro de la Neblina, Rfo Yatua	US holotype
<i>Navia parvula</i> var. <i>expansa</i> L. B. Smith	Maguire, Wurdack, & Maguire 42246	Venezuela: Amazonas: Cerro de la Neblina, Rfo Yatua	US holotype
<i>Navia pauciflora</i> L. B. Smith	*Maguire & Maguire 29154	Venezuela: Amazonas: Cerro Marahuaca, Rfo Cunucunuma	US holotype
<i>Navia phelpsiae</i> L. B. Smith	*Maguire, Phelps et al. 31762	Venezuela: Amazonas: Cerro Guanay, Rfo Ventuari	US isotype
<i>Navia pulvinata</i> L. B. Smith	*Maguire 29111	Venezuela: Amazonas: Cerro Duida	US holotype
<i>Navia pungens</i> L. B. Smith	*Maguire et al. 31652	Venezuela: Amazonas: Cerro Guanay, Rfo Ventuari	US holotype
<i>Navia ramosa</i> L. B. Smith	*Cardona 2997	Venezuela: Bolívar: Cerro Pauo, Rfo Caura	US holotype
<i>Navia reflexa</i> L. B. Smith	*Schultes & Lopez 9321	Colombia: Vaupés: San Felipe (El Castillo) Rfo Negro	US holotype
<i>Navia robinsonii</i> L. B. Smith	*Brewer 250	Venezuela: Bolívar: Cerro Jáua, Rfo Caura	US holotype
<i>Navia rupestris</i> (Gleason) Sandwith	*Maguire & Fanshawe 32360	British Guiana: Essequibo Macreba Falls, Kurupung River	US isotype of <i>N. fluvialilis</i>
<i>Navia sandwithii</i> L. B. Smith	*Maguire & Fanshawe 23435	British Guiana: Kaieteur Falls	US
<i>Navia saxicola</i> L. B. Smith	*Maguire, Cowan, & Wurdack 30680	Venezuela: Amazonas: Cerro Yapacana, Rfo Orinoco	US holotype
<i>Navia schultesiana</i> L. B. Smith	*Schultes 5655	Colombia: Vaupés: Cerro del Castillo, Rfo Apaporis	US isotype
<i>Navia scopulorum</i> L. B. Smith	*Steyermark 75613	Venezuela: Bolívar: Chimantá Massif	US holotype
<i>Navia semiserrata</i> L. B. Smith	*Maguire, Cowan, & Wurdack 30959	Venezuela: Amazonas: Cerro Moriche, Rfo Ventuari	US holotype
<i>Navia serrulata</i> L. B. Smith	*Maguire, Cowan, & Wurdack 30521	Venezuela: Amazonas: Cerro Yapacana, Rfo Orinoco	US holotype

List of Specimens Examined—Continued

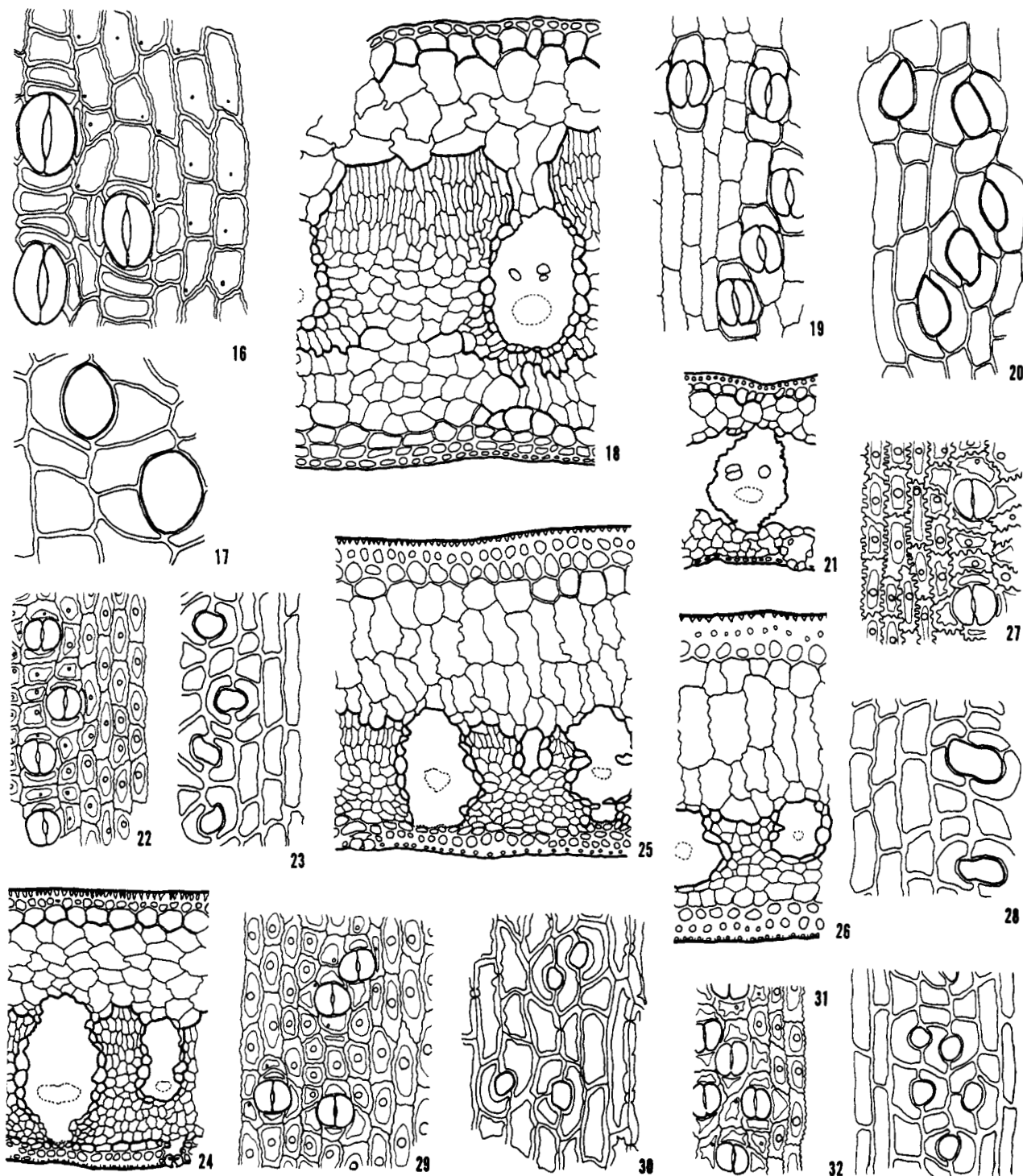
Species	Collector and Number	Locality	Herbarium
<i>Navia splendens</i> L. B. Smith	*Tillett 45841	British Guiana: Essequibo, Utschi Falls, Kamarang River	US isotype
<i>Navia stenodonta</i> L. B. Smith	*Cowan & Wurdack 31357	Venezuela: Amazonas: Serranía Parú, Río Ventuari	US holotype
<i>Navia steyermarkii</i> L. B. Smith	*Steyermark 58012	Venezuela: Amazonas: Cerro Duida	F holotype
<i>Navia subpetiolata</i> L. B. Smith	*Cowan & Wurdack 31368	Venezuela: Amazonas: Serranía Parú Río Ventuari	US holotype
<i>Navia trichodonta</i> L. B. Smith	*Maguire, Cowan, & Wurdack 30228	Venezuela: Amazonas: Cerro Huachamacari, Río Cunucunuma	US holotype
<i>Navia umbratilis</i> L. B. Smith	*Maguire & Maguire 35406	Venezuela: Amazonas: Serranía Yutaje, Río Manapiare	US holotype
<i>Navia viridis</i> L. B. Smith	*Maguire & Maguire 29078	Venezuela: Amazonas: Cerro Duida	US holotype
<i>Navia wurdackii</i> L. B. Smith	*Wurdack 34324	Venezuela: Bolívar: Sarvén-tepuí	US holotype
<i>Navia xyridiflora</i> L. B. Smith	*Steyermark 58152	Venezuela: Amazonas: Cerro Duida	F isotype
<i>Pitcairnia altensteinii</i> (Link ex Kl. & Otto) Lem.	Degado 45	Venezuela	US
<i>Pitcairnia angustifolia</i> Soland.	Sargent 443	Puerto Rico: Maricao	US
<i>Pitcairnia archeri</i> L. B. Smith	Killip 35255	Colombia: Chocó: Upper Río San Juan	US
<i>Pitcairnia armata</i> Maury	Maguire, Wurdack, & Bunting 36094	Venezuela: Amazonas: Great Rapids of Orinoco, Palomal	US
<i>Pitcairnia atrorubens</i> (Beer) Baker	Moore & Bunting 8704	Mexico: Nayarit: Tepic to Jalcocotán	US
<i>Pitcairnia brunnescens</i> L. B. Smith	Asplund 19060	Ecuador: Pichincha: El Paso to El Volante	US
<i>Pitcairnia brongniartiana</i> André	Killip 34835	Colombia: Valle: Río Engaña	US
<i>Pitcairnia brittoniana</i> Mez	Steyermark & Aristeguieta 15	Venezuela: Bolívar: El Dorado to Luepa	US
<i>Pitcairnia bulbosa</i> L. B. Smith	Maguire, Wurdack, & Bunting 36083	Venezuela: Amazonas: Great Rapids of Orinoco, Puerto Ayacucho	US
<i>Pitcairnia caricifolia</i> Martius ex Schultes f.	Cuatrecasas 7520	Colombia: Vaupés: Río Guayabero	US
<i>Pitcairnia gracilis</i> Mez	Stehlé 1804	Guadeloupe: Ste. Rose, near Callas	US
<i>Pitcairnia heterophylla</i> (Lindl.) Beer	Wilbur & Wilbur 2295	Mexico: Jalisco: Autlán	US
<i>Pitcairnia macrochlamys</i> Mez	Breedlove 6359	Mexico: Chiapas: Mun. Tenejapa, Tanaté River	US
<i>Pitcairnia micheliana</i> André	H. E. Moore 8165	Mexico: Jalisco: Colima to Puente San Pedro	US
<i>Pitcairnia multiflora</i> L. B. Smith	Cuatrecasas 23948	Colombia: Valle: Hoya del Río Digua, Río San Juan	US
<i>Pitcairnia nigra</i> (Carr.) André	Asplund 20040	Ecuador: Tungurahua: Río Verde Grande	US
<i>Pitcairnia nuda</i> Baker	A. C. Smith 3644	British Guiana: Mt. Iramaikpang, Kanuka Mts.	US
<i>Pitcairnia oaxacana</i> L. B. Smith	Moore & Bunting 8763	Mexico: Michoacan: Arteaga	US
<i>Pitcairnia paniculata</i> (R. & P.) R. & P.	Macbride 5114	Peru: Huánuco: Pampayacu	US
<i>Pitcairnia pungens</i> H. B. K.	Pittier 1028	Colombia: Cauca: Tacueyó, Río Palo Valley	US
<i>Pitcairnia punicea</i> Scheidw.	Breedlove & Raven 13569	Mexico: Chiapas: Ocozocoautla	US
<i>Pitcairnia ringens</i> Kl. ex Link	Van Hyming 599	Mexico: Veracruz: La Joya	US
<i>Pitcairnia valerii</i> Standley	Allen 2414	Panama: Panama: Cerro Campana	US
<i>Puya santosii</i> Cuatrecasas	Cuatrecasas & Jaramillo 12042	Colombia: Cundinamarca: Sabana de Bogotá	US
<i>Puya sodiroana</i> Mez	Foster 2605	Ecuador: Azuay: Cuenca to Oña	US

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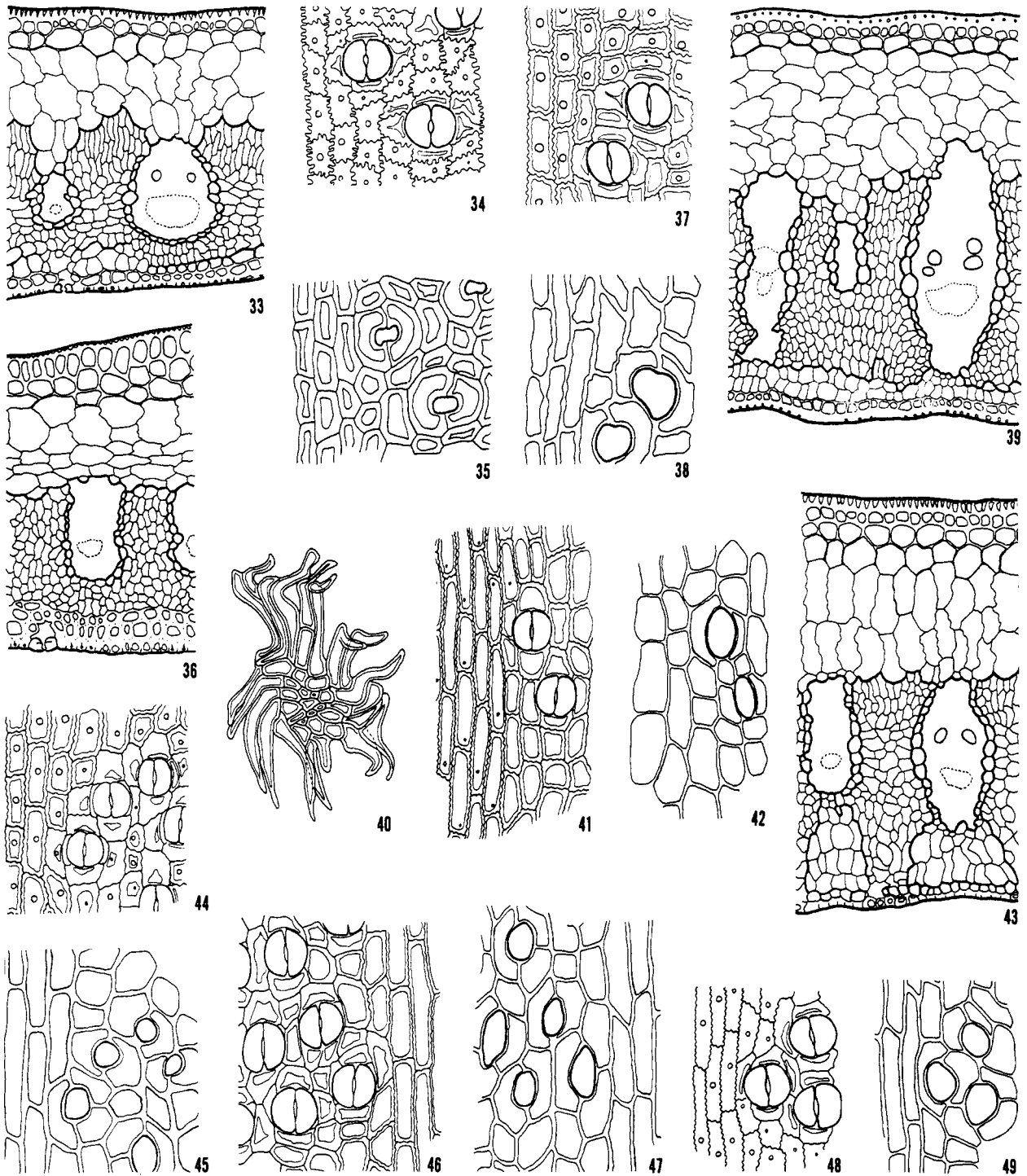
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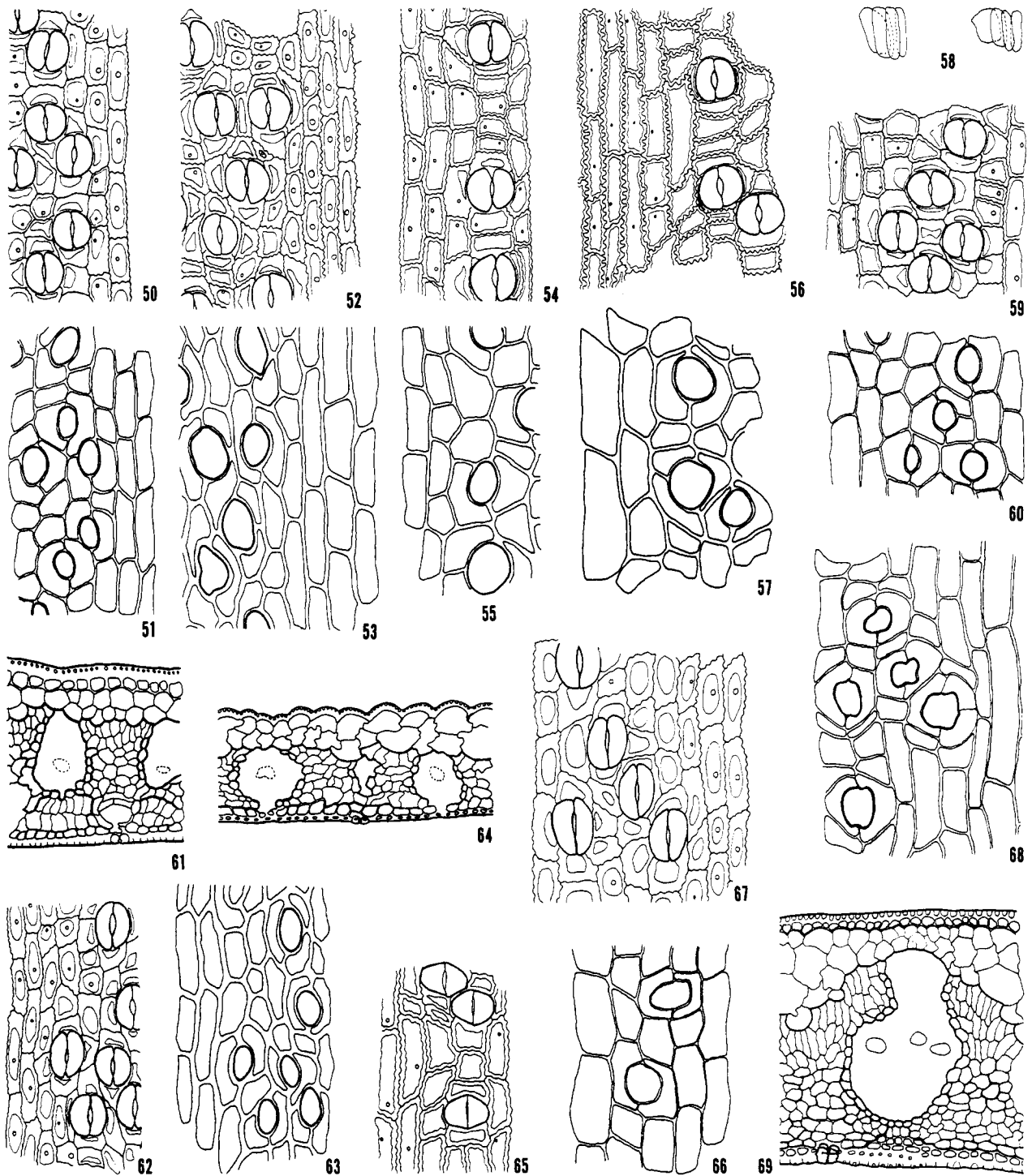
FIGURES 1-15.—*Connellia* and *Cottendorfia*, leaf anatomy. *Connellia guelchii*: 1, cross section, $\times 133$; 2, abaxial epidermis, $\times 267$; 3, abaxial subepidermis, $\times 267$. *Cottendorfia florida*: 4, cross section, $\times 133$; 5, abaxial epidermis, $\times 267$; 6, abaxial subepidermis, $\times 267$. *C. longipes*: 7, abaxial epidermis, $\times 267$; 8, abaxial subepidermis, $\times 267$; 9, cross section, $\times 133$. *C. dyckioides*: 10, abaxial epidermis, $\times 267$; 11, abaxial subepidermis, $\times 267$; 12, cross section, $\times 133$. *C. tillandsioides*: 13, abaxial epidermis, $\times 267$; 14, abaxial subepidermis, $\times 267$; 15, cross section, $\times 133$.



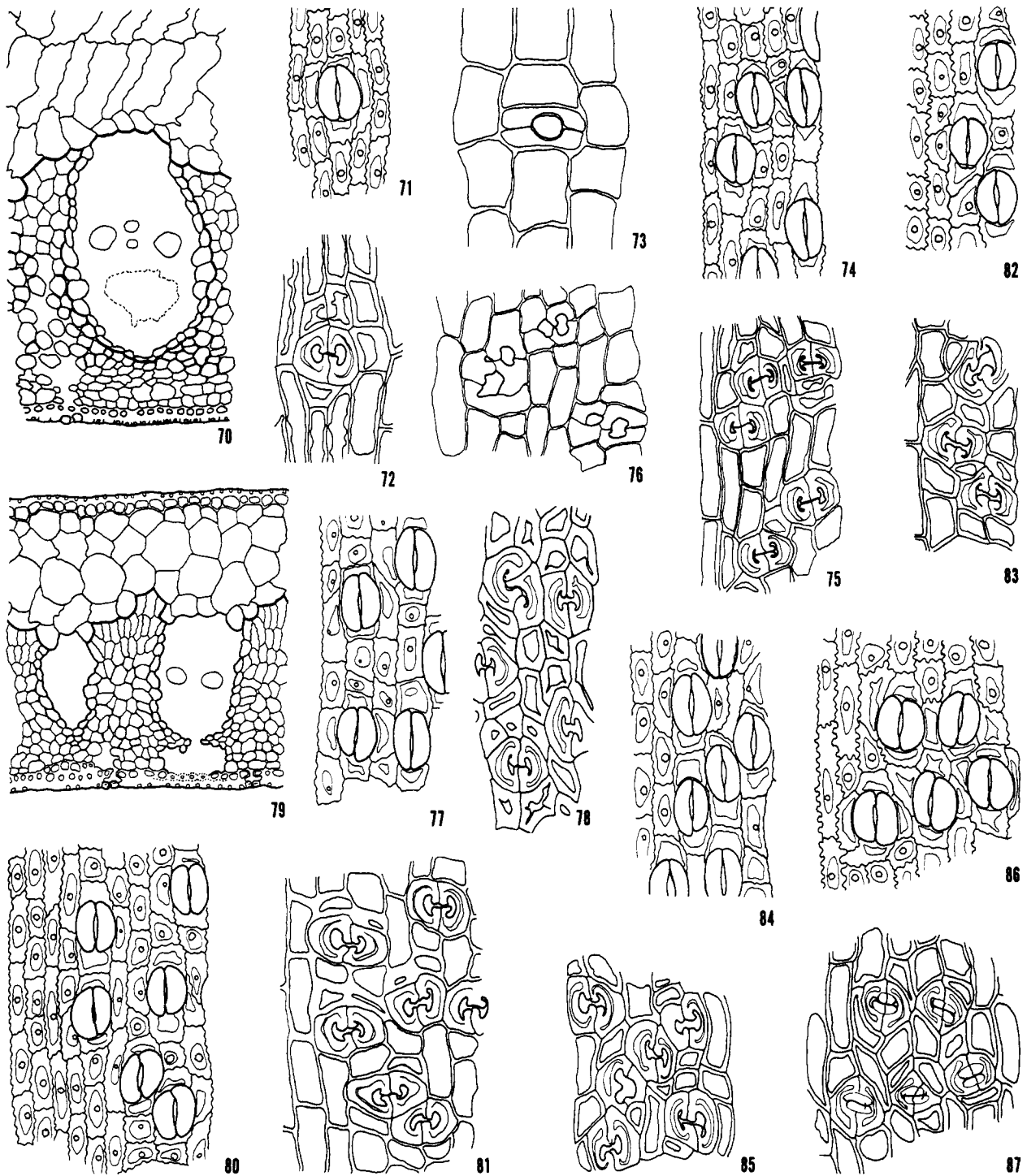
FIGURES 16–32.—*Cottendorfia*, leaf anatomy. *C. brachyphylla*: 16, abaxial epidermis, $\times 267$; 17, abaxial subepidermis, $\times 267$; 18, cross section, $\times 133$. *C. wurdackii*: 19, abaxial epidermis, $\times 267$; 20, abaxial subepidermis, $\times 267$; 21, cross section, $\times 133$. *C. thyrsoides*: 22, abaxial epidermis, $\times 267$; 23, abaxial subepidermis, $\times 267$; 24, cross section, $\times 133$. *C. minor*: 25, cross section from near midleaf, $\times 133$; 26, cross section from near lower fourth, $\times 133$; 27, abaxial epidermis, $\times 267$; 28, abaxial subepidermis, $\times 267$. *C. serrulata* var. *reducta*: 29, abaxial epidermis, $\times 267$; 30, abaxial subepidermis, $\times 267$. *C. gracillima*: 31, abaxial epidermis, $\times 267$; 32, abaxial subepidermis, $\times 267$.



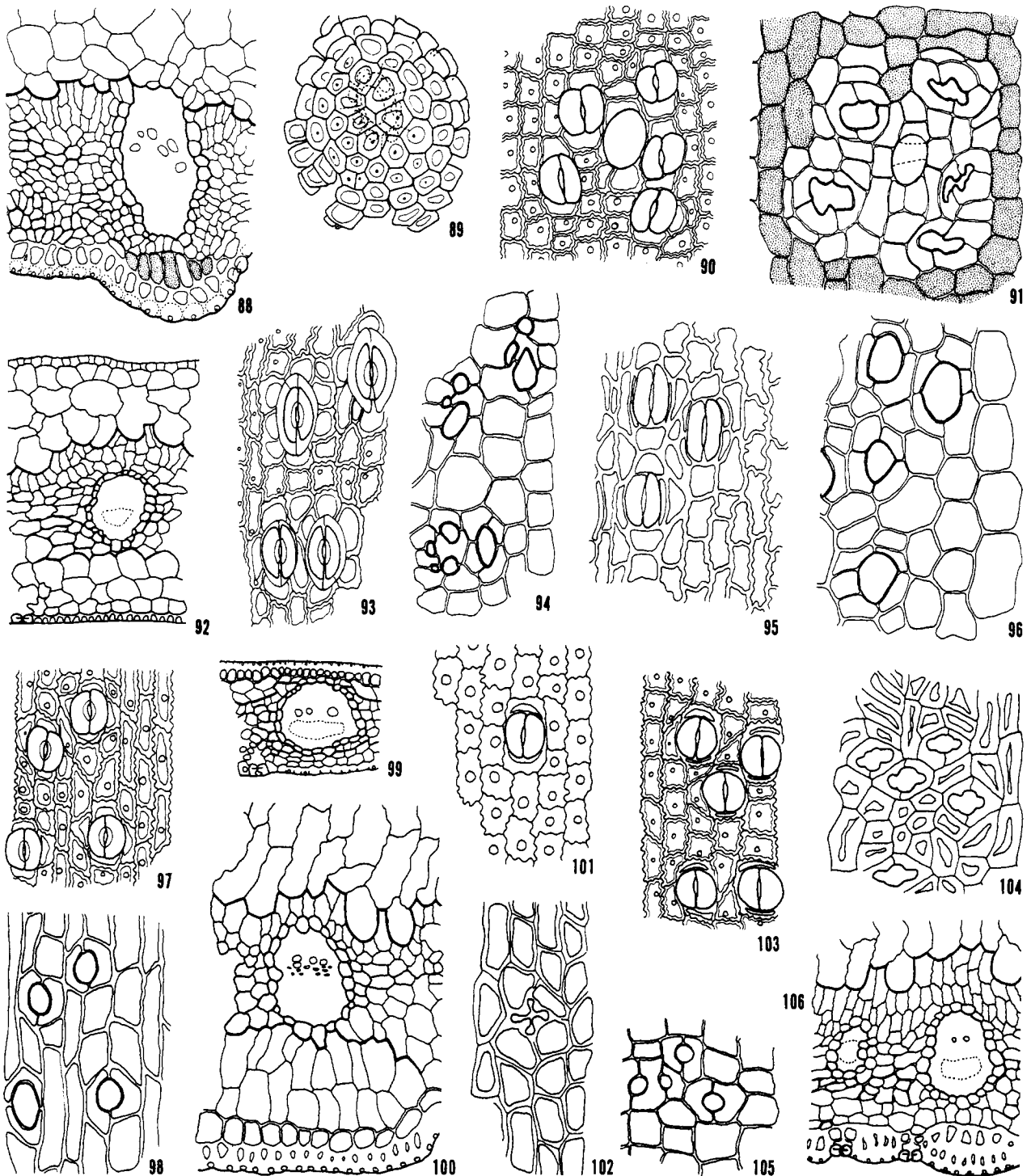
FIGURES 33-49.—*Cottendorfia*, leaf anatomy. *C. serrulata* var. *reducta*: 33, cross section, $\times 133$. *C. subsimplex*: 34, abaxial epidermis, $\times 267$; 35, abaxial subepidermis, $\times 267$; 36, cross section, $\times 133$. *C. stenophylla*: 37, abaxial epidermis, $\times 267$; 38, abaxial subepidermis, $\times 267$; 39, cross section, $\times 133$. *C. argentea*: 40, scale, $\times 267$; 41, abaxial epidermis, $\times 267$; 42, abaxial subepidermis, $\times 267$; 43, cross section, $\times 133$. *C. guianensis*: 44, abaxial epidermis, $\times 267$; 45, abaxial subepidermis, $\times 267$. *C. dendritica*: 46, abaxial epidermis, $\times 267$; 47, abaxial subepidermis, $\times 267$. *C. cylindrostachya*: 48, abaxial epidermis, $\times 267$; 49, abaxial subepidermis, $\times 267$.



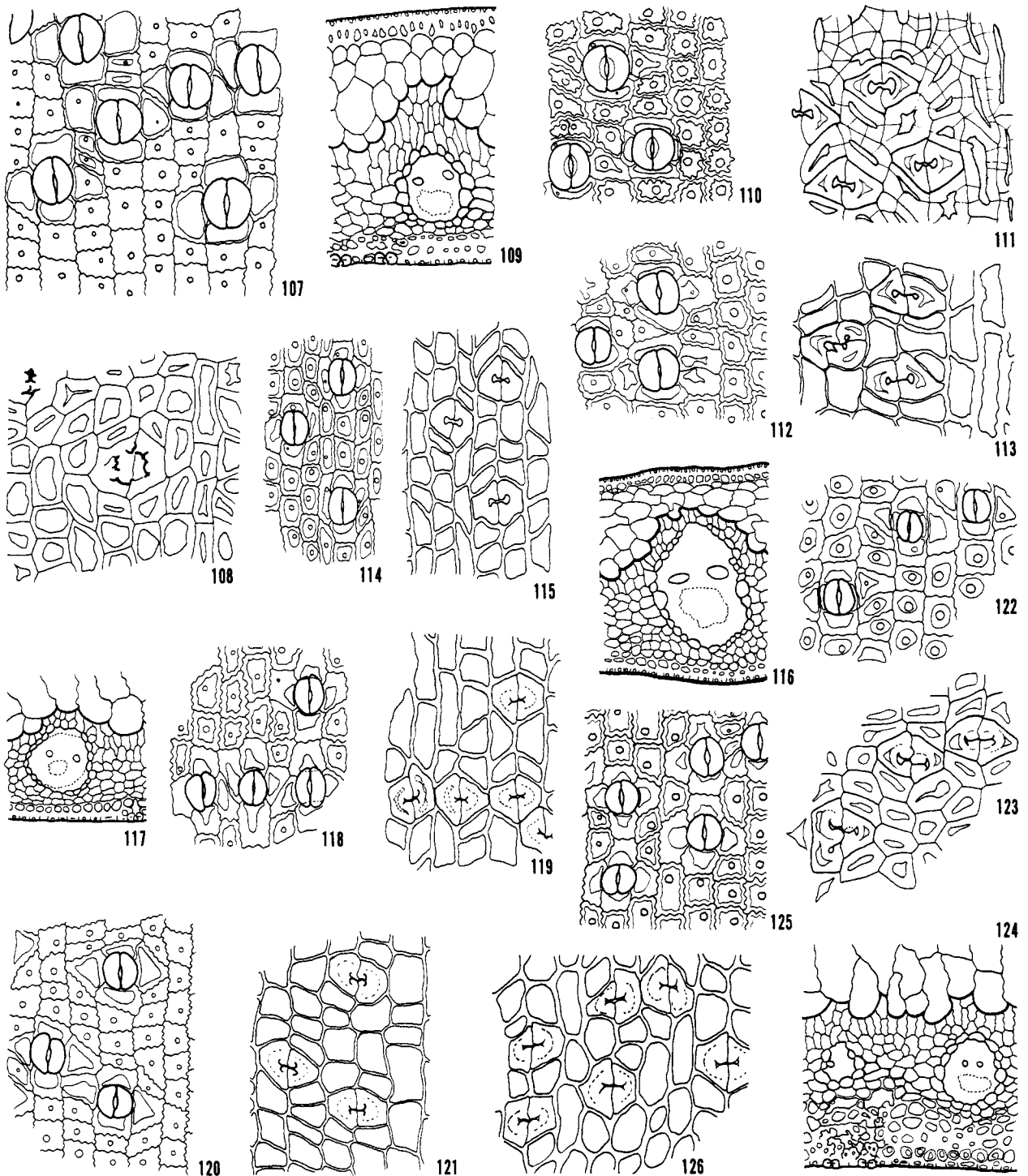
FIGURES 50-69.—*Cottendorfia* and *Navia*, leaf anatomy. *Cottendorfia geniculata*: 50, abaxial epidermis, $\times 267$; 51, abaxial subepidermis, $\times 267$. *C. maguirei*: 52, abaxial epidermis, $\times 267$; 53, abaxial subepidermis, $\times 267$. *C. nubigena*: 54, abaxial epidermis, $\times 267$; 55, abaxial subepidermis, $\times 267$. *C. savannensis*: 56, abaxial epidermis, $\times 267$; 57, abaxial subepidermis, $\times 267$; 58, scales, $\times 267$. *C. paludosa*: 59, abaxial epidermis, $\times 267$; 60, abaxial subepidermis, $\times 267$. *C. phelpsi*: 61, cross section, $\times 133$; 62, abaxial epidermis, $\times 267$; 63, abaxial subepidermis, $\times 267$. *C. navioides*: 64, cross section, $\times 133$; 65, abaxial epidermis, $\times 267$; 66, abaxial subepidermis, $\times 267$. *Navia ramosa*: 67, abaxial epidermis, $\times 267$; 68, abaxial subepidermis, $\times 267$; 69 cross section, $\times 133$.



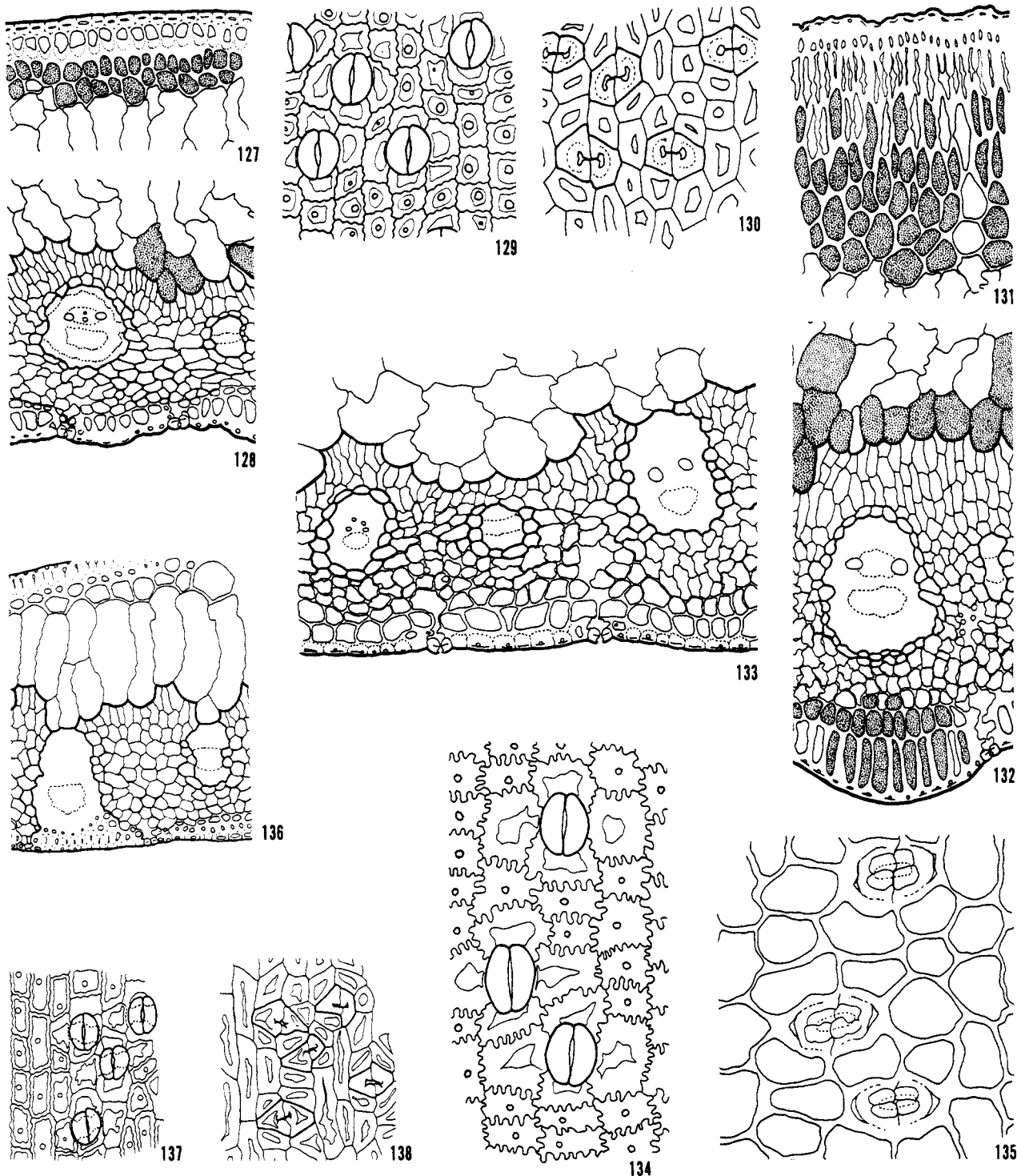
FIGURES 70-87.—*Navia*, leaf anatomy. *N. reflexa*: 70, cross section, $\times 133$; 71, adaxial epidermis, $\times 267$; 72, adaxial subepidermis, $\times 267$; 73, adaxial third cell layer, $\times 267$; 74, abaxial epidermis, $\times 267$; 75, abaxial subepidermis, $\times 267$; 76, abaxial third cell layer, $\times 267$. *N. hechtioides*: 77, abaxial epidermis, $\times 267$; 78, abaxial subepidermis, $\times 267$. *N. hohenbergioides*: 79, cross section, $\times 133$; 80, abaxial epidermis, $\times 267$; 81, abaxial subepidermis, $\times 267$. *N. gracilis*: 82, abaxial epidermis, $\times 267$; 83, abaxial subepidermis, $\times 267$. *N. brocchinioides*: 84, abaxial epidermis, $\times 267$; 85, abaxial subepidermis, $\times 267$. *N. diffusa*: 86, abaxial epidermis, $\times 267$; 87, abaxial subepidermis, $\times 267$.



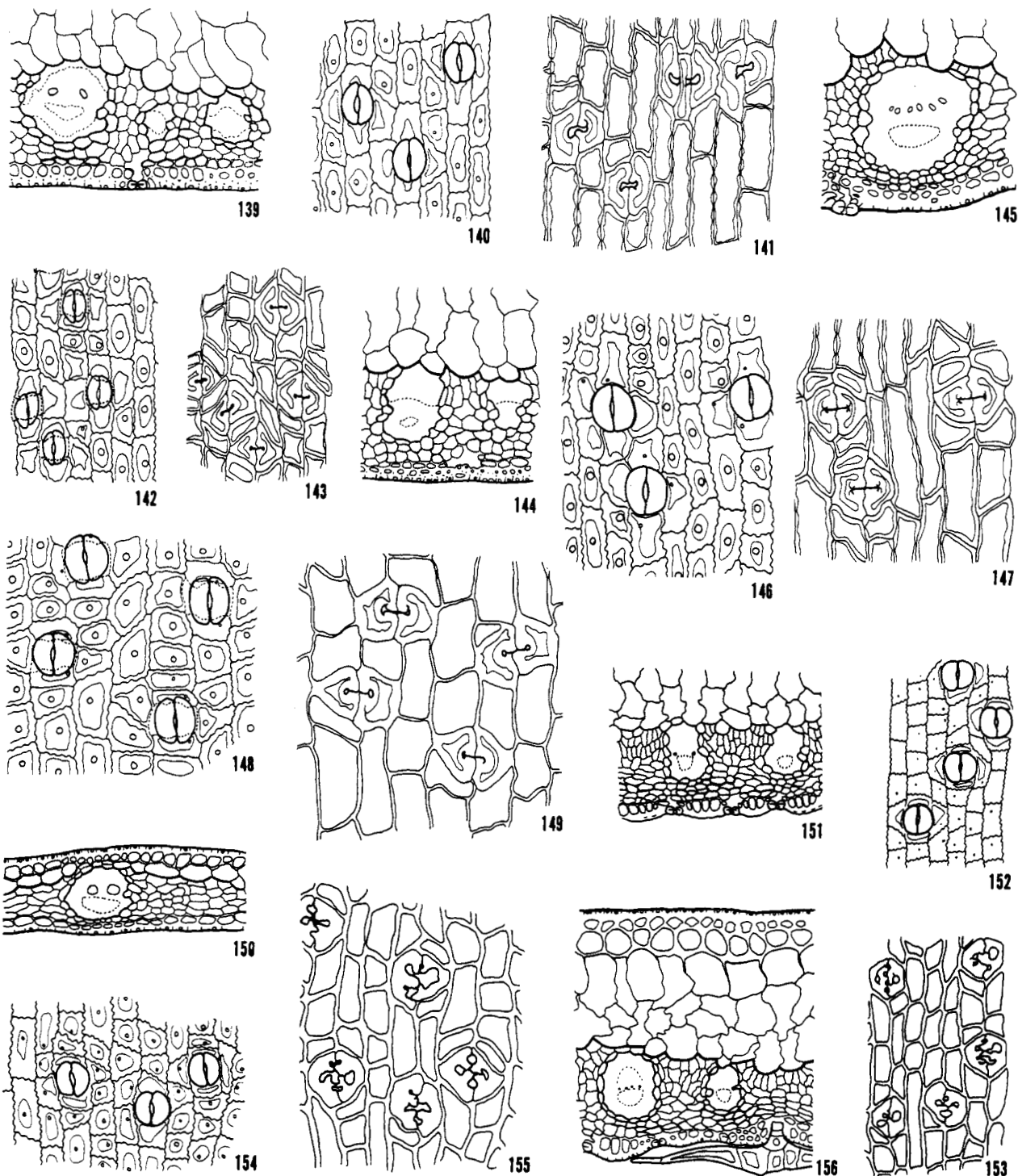
FIGURES 88-106.—*Navia*, leaf anatomy. *N. garcia-barrigae*: 88, cross section, $\times 133$; 89, scale, $\times 133$; 90, abaxial epidermis, $\times 267$; 91, abaxial subepidermis, $\times 267$. *N. rupestris*: 92, cross section, $\times 133$; 93, abaxial epidermis, $\times 267$; 94, abaxial subepidermis, $\times 267$. *N. cataractarum*: 95, abaxial epidermis, $\times 267$; 96, abaxial subepidermis, $\times 267$. *N. lindmanioides*: 97, abaxial epidermis, $\times 267$; 98, abaxial subepidermis, $\times 267$; 99, cross section, $\times 133$. *N. parvula*: 100, cross section, $\times 133$; 101, abaxial epidermis, $\times 267$; 102, abaxial subepidermis, $\times 267$. *N. lepidota*: 103, abaxial epidermis, $\times 267$; 104, abaxial subepidermis, $\times 267$; 105, abaxial third cell layer, $\times 267$; 106, cross section, $\times 133$.



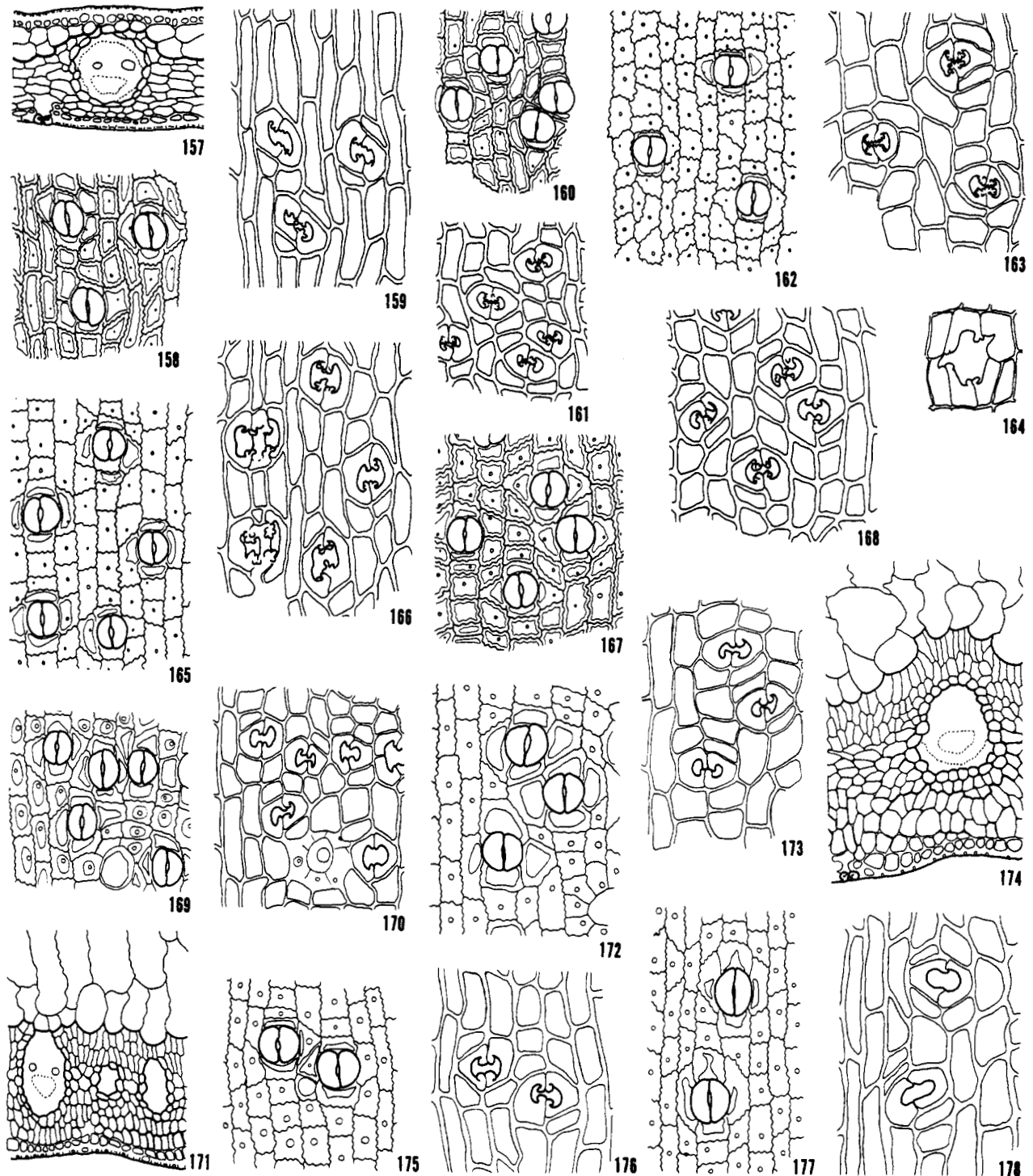
FIGURES 107-126.—*Navia*, leaf anatomy. *N. involucrata*: 107, abaxial epidermis, $\times 267$; 108, abaxial subepidermis, $\times 267$. *N. barbellata*: 109, cross section, $\times 133$; 110, abaxial epidermis, $\times 267$; 111, abaxial subepidermis, $\times 267$. *N. splendens*: 112, abaxial epidermis, $\times 267$; 113, abaxial subepidermis, $\times 267$. *N. fontoides*: 114, abaxial epidermis, $\times 267$; 115, abaxial subepidermis, $\times 267$; 116, cross section, $\times 133$. *N. cardonae*: 117, cross section, $\times 133$; 118, abaxial epidermis, $\times 267$; 119, abaxial subepidermis, $\times 267$. *N. robinsonii*: 120, abaxial epidermis, $\times 267$; 121, abaxial subepidermis, $\times 267$. *N. aurea*: 122, abaxial epidermis, $\times 267$; 123, abaxial subepidermis, $\times 167$; 124, cross section, $\times 133$. *N. lasiantha*: 125, abaxial epidermis, $\times 267$; 126, abaxial subepidermis, $\times 267$.



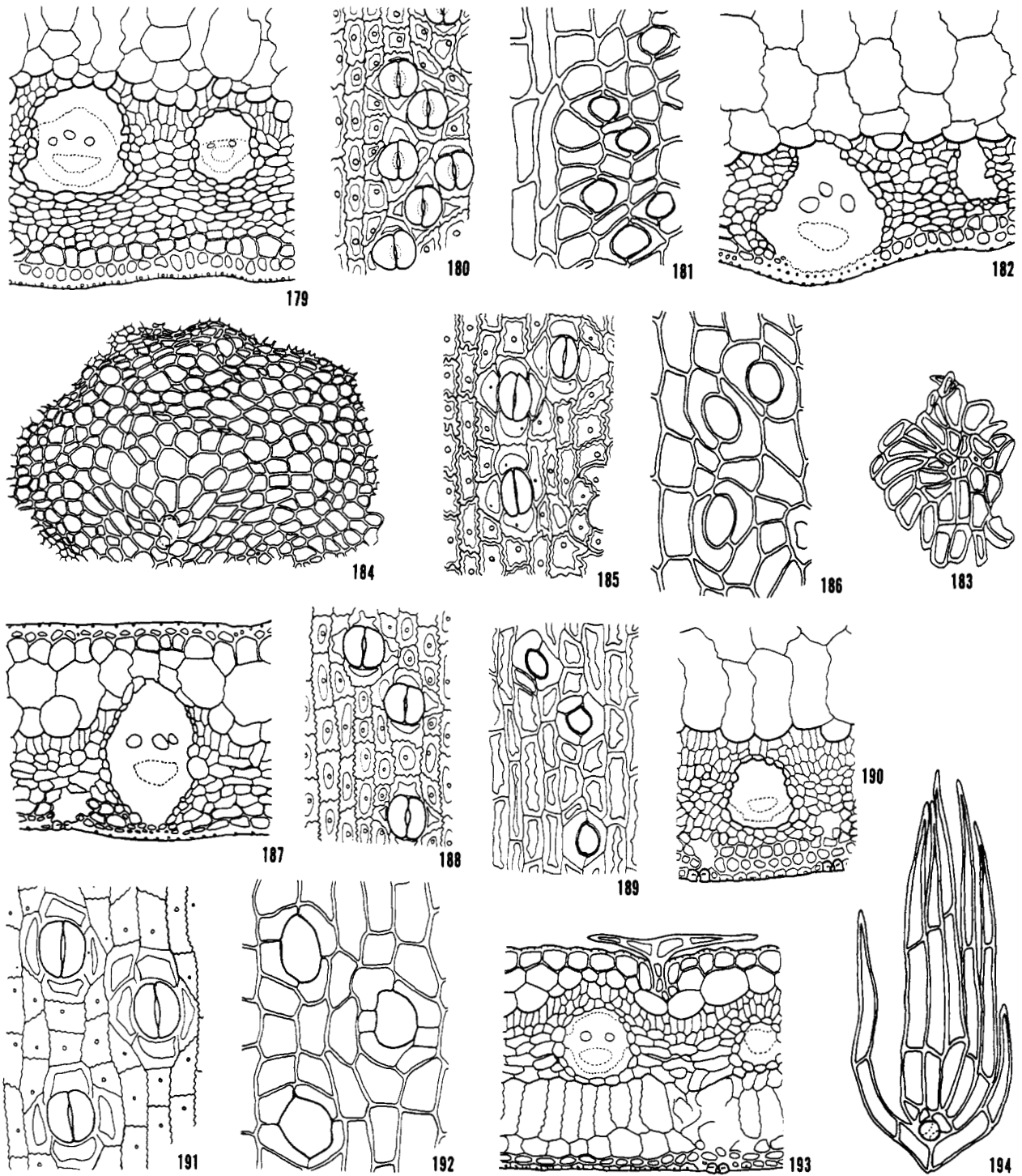
FIGURES 127-138.—*Navia*, leaf anatomy. *N. lasiantha*: 127, cross section, adaxial surface, $\times 133$; 128, cross section, abaxial surface, $\times 133$. *N. incrassata*: 129, abaxial epidermis, $\times 267$; 130, abaxial subepidermis, $\times 167$; 131, cross section, adaxial surface, $\times 133$; 132, cross section, abaxial surface, $\times 133$. *N. intermedia*: 133, cross section, $\times 133$; 134, abaxial epidermis, $\times 267$; 135, abaxial subepidermis, $\times 267$. *N. immersa*: 136, cross section, $\times 133$; 137, abaxial epidermis, $\times 267$; 138, abaxial subepidermis, $\times 267$.



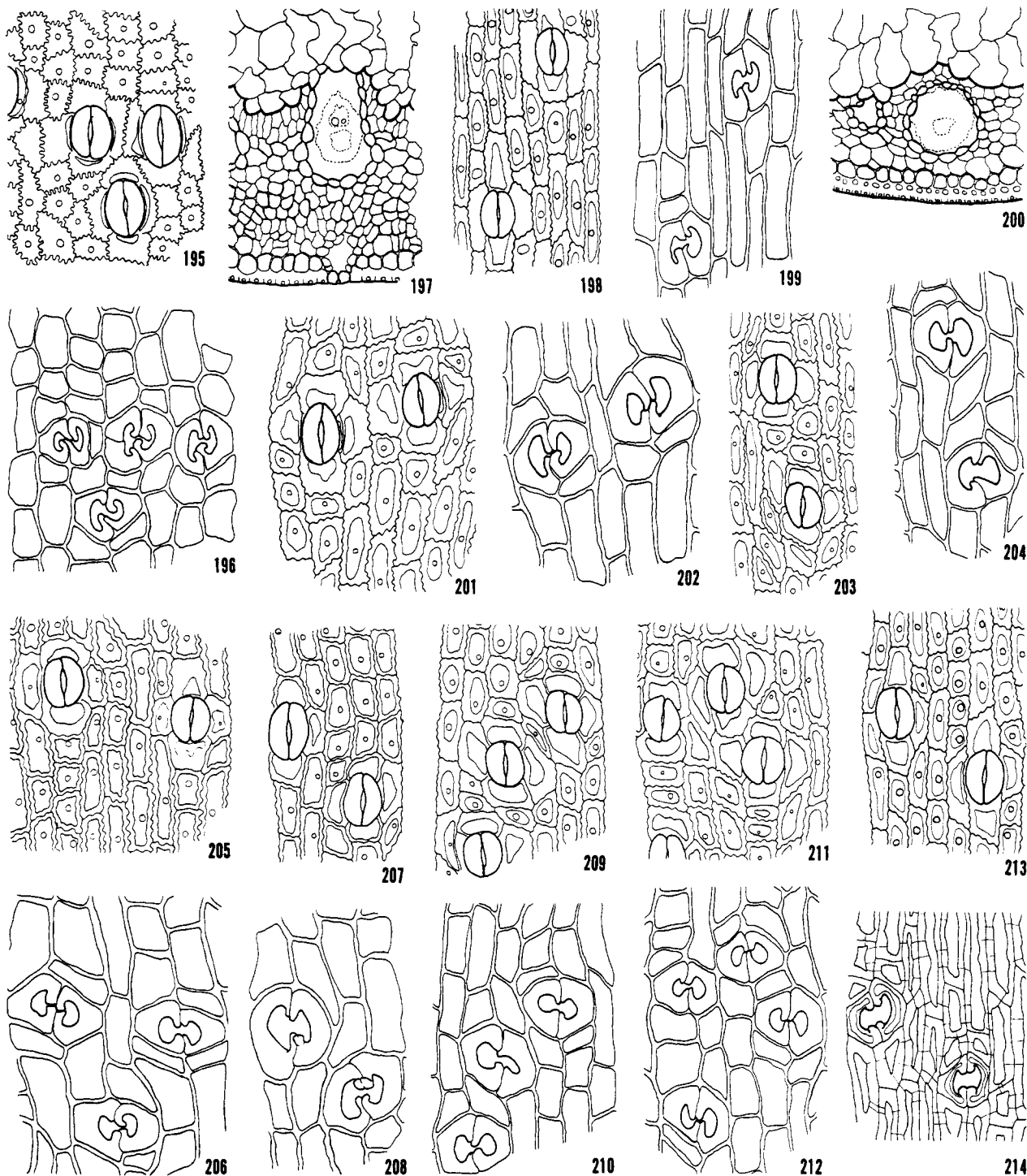
FIGURES 139–156.—*Navia*, leaf anatomy. *N. cucullata*: 139, cross section, $\times 133$; 140, abaxial epidermis, $\times 267$; 141, abaxial subepidermis, $\times 267$. *N. pulvinata*: 142, abaxial epidermis, $\times 267$; 143, abaxial subepidermis, $\times 267$; 144, cross section, $\times 133$. *N. scopulorum*: 145, cross section, $\times 133$; 146, abaxial epidermis, $\times 267$; 147, abaxial subepidermis, $\times 267$. *N. angustifolia*: 148, abaxial epidermis, $\times 267$; 149, abaxial subepidermis, $\times 267$; 150, cross section, $\times 133$. *N. abyssophila*: 151, cross section, $\times 133$; 152, abaxial epidermis, $\times 267$; 153, abaxial subepidermis, $\times 267$. *N. duidae*: 154, abaxial epidermis, $\times 267$; 155, abaxial subepidermis, $\times 267$; 156, cross section, $\times 133$.



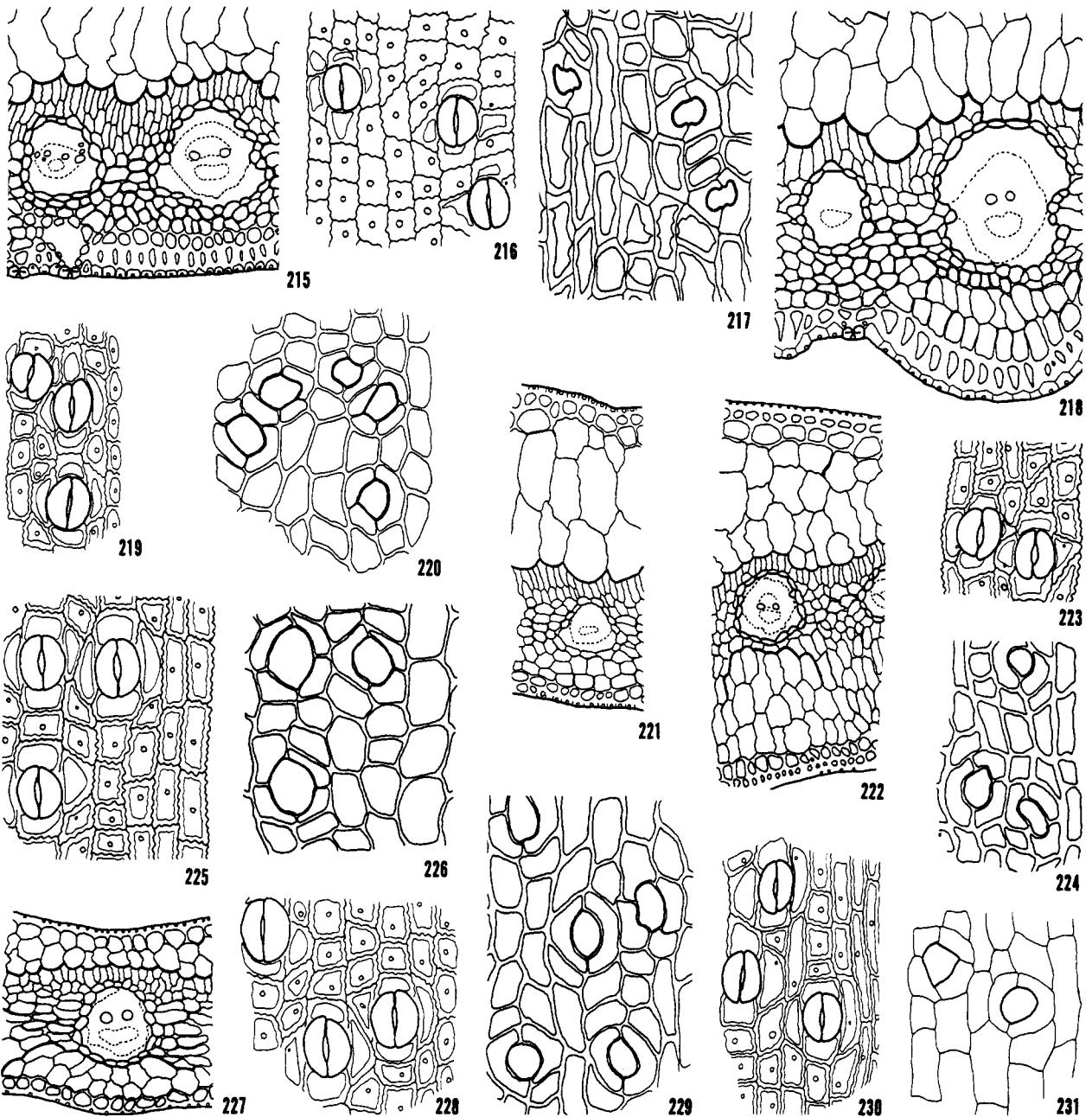
FIGURES 157-178.—*Navia*, leaf anatomy. *N. nubicola*: 157, cross section, $\times 133$; 158, abaxial subepidermis, $\times 267$. *N. cretacea*: 160, abaxial epidermis, $\times 267$; 161, abaxial subepidermis, $\times 267$. *N. latifolia*: 162, abaxial epidermis, $\times 267$; 163, abaxial subepidermis, $\times 267$; 164, abaxial third cell layer, $\times 267$. *N. brachyphylla*: 165, abaxial epidermis, $\times 267$; 166, abaxial subepidermis, $\times 267$. *N. glauca*: 167, abaxial epidermis, $\times 267$; 168, abaxial subepidermis, $\times 267$. *N. semiserrata*: 169, abaxial epidermis, $\times 267$; 170, abaxial subepidermis, $\times 267$; 171, cross section, $\times 133$. *N. caurensis*: 172, abaxial epidermis, $\times 267$; 173, abaxial subepidermis, $\times 267$; 174, cross section, $\times 133$. *N. octopoides*: 175, abaxial epidermis, $\times 267$; 176, abaxial subepidermis, $\times 267$. *N. umbratilis*: 177, abaxial epidermis, $\times 267$; 178, abaxial subepidermis, $\times 267$.



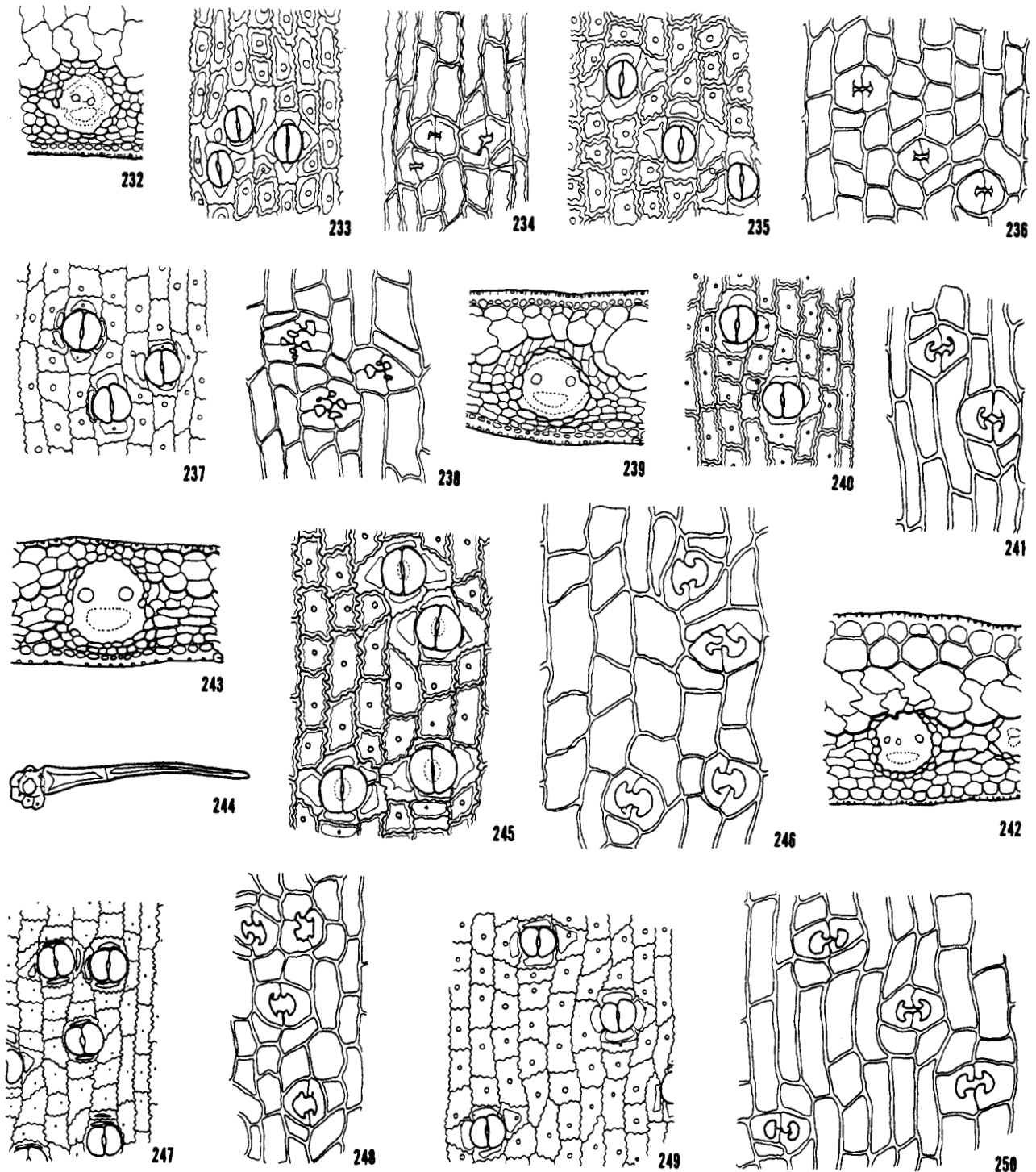
FIGURES 179-194.—*Navia*, leaf anatomy. *N. umbratilis*: 179, cross section, $\times 133$. *N. serrulata*: 180, abaxial epidermis, $\times 267$; 181, abaxial subepidermis, $\times 267$; 182, cross section, $\times 133$; 183, scale, $\times 133$. *N. arida*: 184, scale, $\times 133$; 185, abaxial epidermis, $\times 267$; 186, abaxial subepidermis, $\times 267$; 187, cross section, $\times 133$. *N. saxicola*: 188, abaxial epidermis, $\times 267$; 189, abaxial subepidermis, $\times 267$; 190, cross section, $\times 133$. *N. caulescens*: 191, abaxial epidermis, $\times 267$; 192, abaxial subepidermis, $\times 267$; 193, cross section, $\times 133$; 194, scale, $\times 133$.



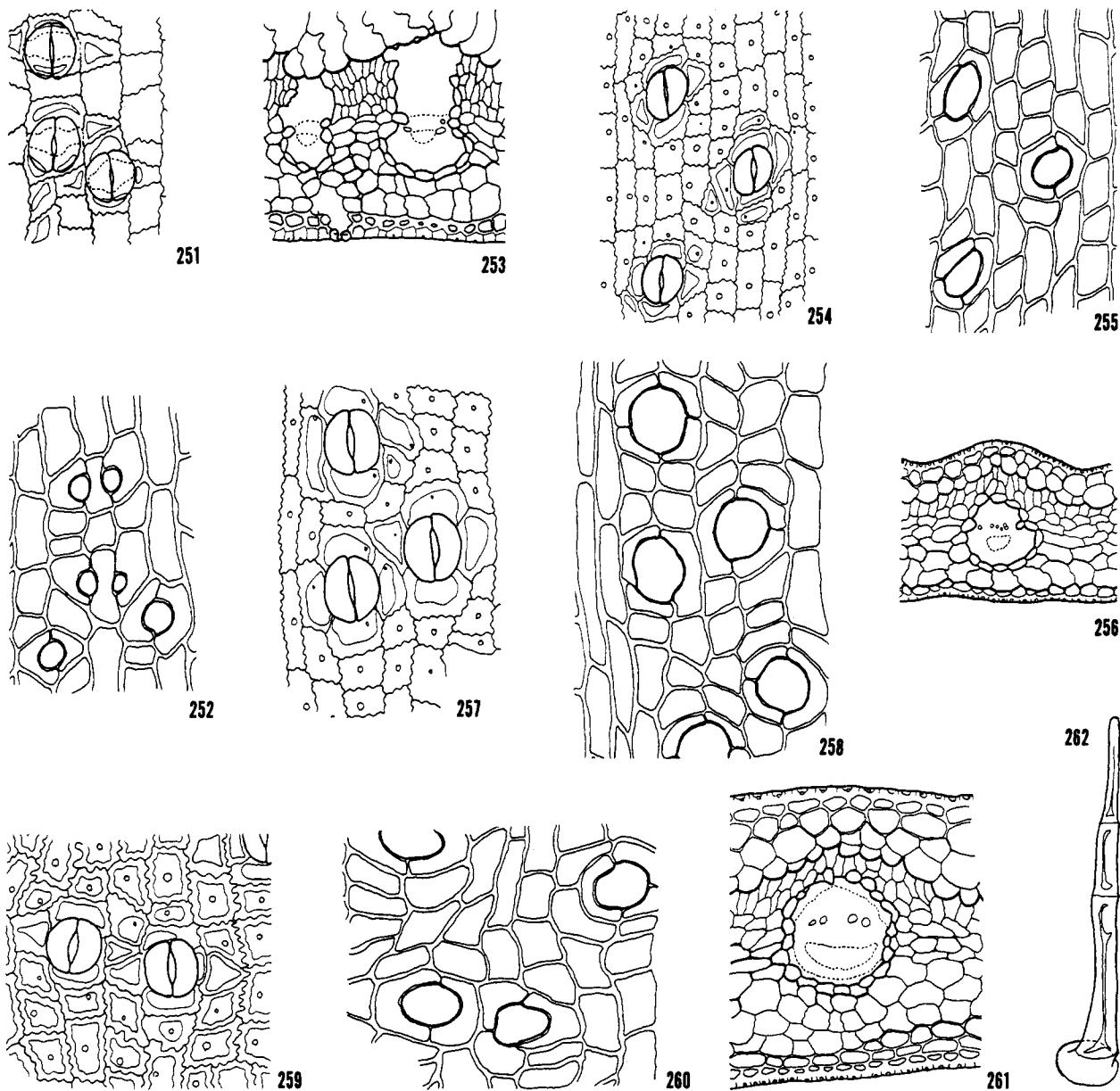
FIGURES 195-214.—*Navia*, leaf anatomy. *N. caricifolia*: 195, abaxial epidermis, $\times 267$; 196, abaxial subepidermis, $\times 267$; 197, cross section, $\times 133$. *N. ocellata*: 198, abaxial epidermis, $\times 267$; 199, abaxial subepidermis, $\times 267$; 200, cross section, $\times 133$. *N. maguirei*: 201, abaxial epidermis, $\times 267$; 202, abaxial subepidermis, $\times 267$. *N. subpetiolata*: 203, abaxial epidermis, $\times 267$; 204, abaxial subepidermis, $\times 267$. *N. gleasonii*: 205, abaxial epidermis, $\times 267$; 206, abaxial subepidermis, $\times 267$. *N. sandwithii*: 207, abaxial epidermis, $\times 267$; 208, abaxial subepidermis, $\times 267$. *N. connata*: 209, abaxial epidermis, $\times 267$; 210, abaxial subepidermis, $\times 267$. *N. affinis*: 211, abaxial epidermis, $\times 267$; 212, abaxial subepidermis, $\times 267$. *N. pungens*: 213, abaxial epidermis, $\times 267$; 214, abaxial subepidermis, $\times 267$.



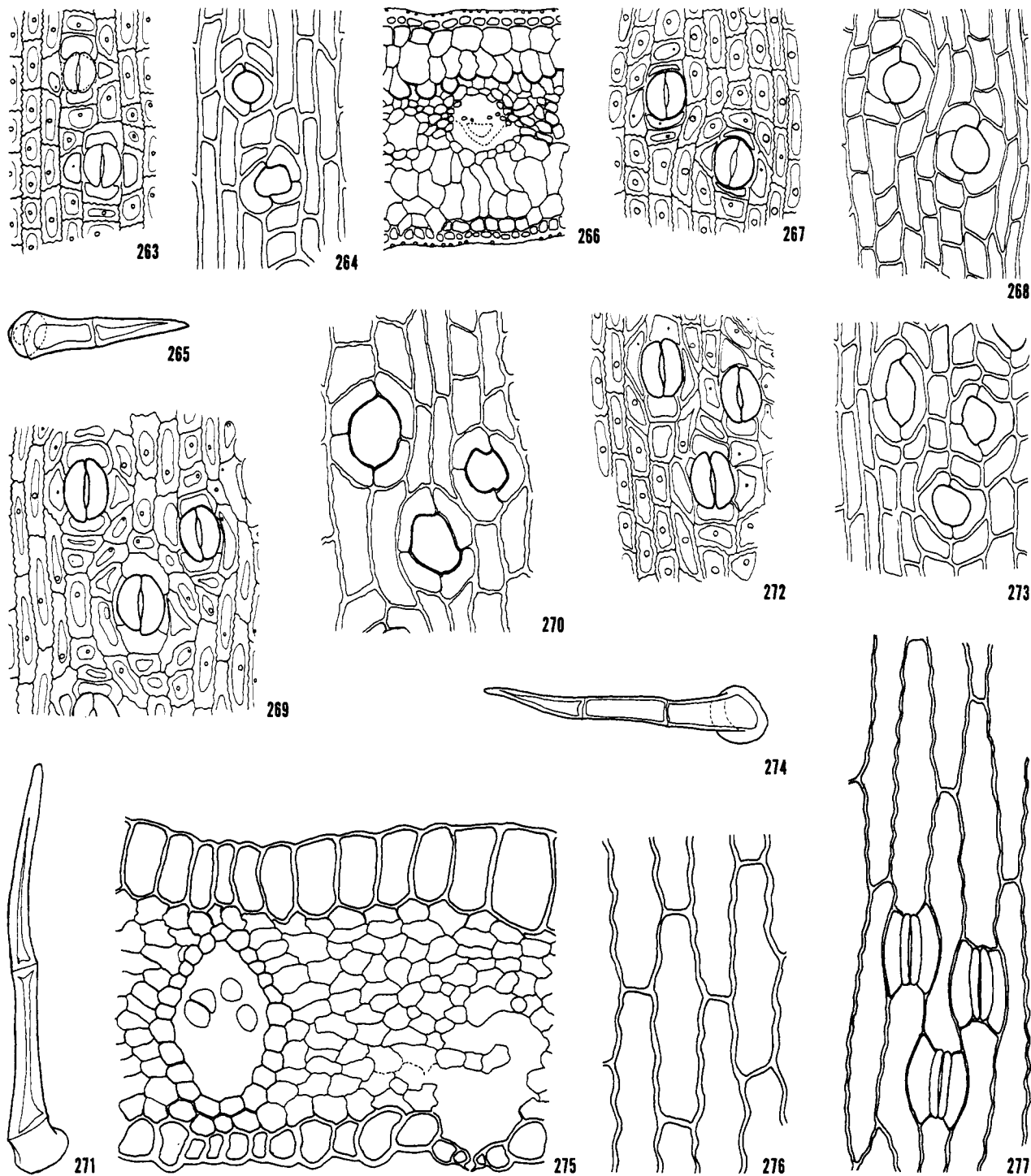
FIGURES 215-231.—*Navia*, leaf anatomy. *N. pungens*: 215, cross section, $\times 133$. *N. mima*: 216, abaxial epidermis, $\times 267$; 217, abaxial subepidermis, $\times 267$; 218, cross section, $\times 133$. *N. schultesiana*: 219, abaxial epidermis, $\times 267$; 220, abaxial subepidermis, $\times 267$; 221, cross section, $\times 133$. *N. graminifolia*: 222, cross section, $\times 133$; 223, abaxial epidermis, $\times 267$; 224, abaxial subepidermis, $\times 267$. *N. heliophila*: 225, abaxial epidermis, $\times 267$; 226, abaxial subepidermis, $\times 267$; 227, cross section, $\times 133$. *N. bicolor*: 228, abaxial epidermis, $\times 267$; 229, abaxial subepidermis, $\times 267$. *N. acaulis*: 230, abaxial epidermis, $\times 267$; 231, abaxial subepidermis, $\times 267$.



FIGURES 232-250.—*Navia*, leaf anatomy. *N. navicularis*: 232, cross section, $\times 133$; 233, abaxial epidermis, $\times 267$; 234, abaxial subepidermis, $\times 267$. *N. breweri*: 235, abaxial epidermis, $\times 267$; 236, abaxial subepidermis, $\times 267$. *N. colorata*: 237, abaxial epidermis, $\times 267$; 238, abaxial subepidermis, $\times 267$; 239, cross section, $\times 133$. *N. xyridiflora*: 240, abaxial epidermis, $\times 267$; 241, abaxial subepidermis, $\times 267$; 242, cross section, $\times 133$. *N. wurdackii*: 243, cross section, $\times 133$; 244, trichome, $\times 133$; 245, abaxial epidermis, $\times 267$; 246, abaxial subepidermis, $\times 267$. *N. pauciflora*: 247, abaxial epidermis, $\times 267$; 248, abaxial subepidermis, $\times 267$. *N. steyermarkii*: 249, abaxial epidermis, $\times 267$; 250, abaxial subepidermis, $\times 267$.



FIGURES 251–262.—*Navia*, leaf anatomy. *N. aloifolia*: 251, abaxial epidermis, $\times 267$; 252, abaxial subepidermis, $\times 267$; 253, cross section, $\times 133$. *N. viridis*: 254, abaxial epidermis, $\times 267$; 255, abaxial subepidermis, $\times 267$; 256, cross section, $\times 133$. *N. crispa*: 257, abaxial epidermis, $\times 267$; 258, abaxial subepidermis, $\times 267$. *N. stenodonta*: 259, abaxial epidermis, $\times 267$; 260, abaxial subepidermis, $\times 267$; 261 cross section, $\times 133$; 262, trichome, $\times 133$.



FIGURES 263-277.—*Navia*, leaf anatomy. *N. myriantha*: 263, abaxial epidermis, $\times 267$; 264, abaxial subepidermis, $\times 267$; 265, trichome, $\times 133$; 266, cross section, $\times 133$. *N. trichodonta*: 267, abaxial epidermis, $\times 267$; 268, abaxial subepidermis, $\times 267$. *N. lanigera*: 269, abaxial epidermis, $\times 267$; 270, abaxial subepidermis, $\times 267$; 271, trichome, $\times 133$. *N. phelpsiae*: 272, abaxial epidermis, $\times 267$; 273, abaxial subepidermis, $\times 267$; 274, trichome, $\times 133$. *N. lopezii*: 275, cross section, $\times 133$; 276, abaxial epidermis, $\times 133$; 277, abaxial epidermis, $\times 133$.