

**TOXICOLOGICAL PROFILE FOR
2,4- and 2,6-DINITROTOLUENE**

U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES
Public Health Service
Agency for Toxic Substances and Disease Registry

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UPDATE STATEMENT

A Toxicological Profile for 2,4- and 2,6-dinitrotoluene was released in September 1997. This edition supersedes any previously released draft or final profile.

Toxicological profiles are revised and republished as necessary, but no less than once every three years. For information regarding the update status of previously released profiles, contact ATSDR at:

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FOREWORD

This toxicological profile is prepared in accordance with guidelines* developed by the Agency for Toxic Substances and Disease Registry (ATSDR) and the Environmental Protection Agency (EPA). The original guidelines were published in the *Federal Register* on April 17, 1987. Each profile will be revised and republished as necessary.

The ATSDR toxicological profile succinctly characterizes the toxicologic and adverse health effects information for the hazardous substance described therein. Each peer-reviewed profile identifies and reviews the key literature that describes a hazardous substance's toxicologic properties. Other pertinent literature is also presented, but is described in less detail than the key studies. The profile is not intended to be an exhaustive document; however, more comprehensive sources of specialty information are referenced.

The focus of the profiles is on health and toxicologic information; therefore, each toxicological profile begins with a public health statement that describes, in nontechnical language, a substance's relevant toxicological properties. Following the public health statement is information concerning levels of significant human exposure and, where known, significant health effects. The adequacy of information to determine a substance's health effects is described in a health effects summary. Data needs that are of significance to protection of public health are identified by ATSDR and EPA.

Each profile includes the following:

- (A) The examination, summary, and interpretation of available toxicologic information and epidemiologic evaluations on a hazardous substance to ascertain the levels of significant human exposure for the substance and the associated acute, subacute, and chronic health effects;
- (B) A determination of whether adequate information on the health effects of each substance is available or in the process of development to determine levels of exposure that present a significant risk to human health of acute, subacute, and chronic health effects; and
- (C) Where appropriate, identification of toxicologic testing needed to identify the types or levels of exposure that may present significant risk of adverse health effects in humans.

The principal audiences for the toxicological profiles are health professionals at the Federal, State, and local levels; interested private sector organizations and groups; and members of the public.

This profile reflects ATSDR's assessment of all relevant toxicologic testing and information that has been peer-reviewed. Staff of the Centers for Disease Control and Prevention and other Federal scientists have also reviewed the profile. In addition, this profile has been peer-reviewed by a nongovernmental panel and was made available for public review. Final responsibility for the contents and views expressed in this toxicological profile resides with ATSDR.



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*Legislative Background

The toxicological profiles are developed in response to the Super-fund Amendments and Reauthorization Act (SARA) of 1986 (Public Law 99-499) which amended the Comprehensive Environmental Response, Compensation, and Liability Act of 1980 (CERCLA or Superfund). This public law directed ATSDR to prepare toxicological profiles for hazardous substances most commonly found at facilities on the CERCLA National Priorities List and that pose the most significant potential threat to human health, as determined by ATSDR and the EPA. The availability of the revised priority list of 275 hazardous substances was announced in the Federal Register on November 17, 1997 (62 FR 61332). For prior versions of the list of substances, see Federal Register notices dated April 29, 1996 (61 FR 18744); April 17, 1987 (52 FR 12866); October 20, 1988 (53 FR 41280); October 26, 1989 (54 FR 43619); October 17, 1990 (55 FR 42067); October 17, 1991 (56 FR 52166); October 28, 1992 (57 FR 48801); and February 28, 1994 (59 FR 9486). Section 104(i)(3) of CERCLA, as amended, directs the Administrator of ATSDR to prepare a toxicological profile for each substance on the list.

QUICK REFERENCE FOR HEALTH CARE PROVIDERS

Toxicological Profiles are a unique compilation of toxicological information on a given hazardous substance. Each profile reflects a comprehensive and extensive evaluation, summary, and interpretation of available toxicologic and epidemiologic information on a substance. Health care providers treating patients potentially exposed to hazardous substances will find the following information helpful for fast answers to often-asked questions.

Primary Chapters/Sections of Interest

Chapter 1: Public Health Statement: The Public Health Statement can be a useful tool for educating patients about possible exposure to a hazardous substance. It explains a substance's relevant toxicologic properties in a nontechnical, question-and-answer format, and it includes a review of the general health effects observed following exposure.

Chapter 2: Health Effects: Specific health effects of a given hazardous compound are reported by *route of exposure*, by *type of health effect* (death, systemic, immunologic, reproductive), and by *length of exposure* (acute, intermediate, and chronic). In addition, both human and animal studies are reported in this section.

NOTE: Not all health effects reported in this section are necessarily observed in the clinical setting. Please refer to the Public Health Statement to identify general health effects observed following exposure.

Pediatrics: Four new sections have been added to each Toxicological Profile to address child health issues:

Section 1.6 How Can (Chemical X) Affect Children?

Section 1.7 How Can Families Reduce the Risk of Exposure to (Chemical X)?

Section 2.6 Children's Susceptibility

Section 5.6 Exposures of Children

Other Sections of Interest:

Section 2.7 Biomarkers of Exposure and Effect

Section 2.10 Methods for Reducing Toxic Effects

ATSDR Information Center

Phone: 1-800-447-1544 (to be replaced by 1-888-42-ATSDR in 1999)

or 404-639-6357 **Fax:** 404-639-6359

E-mail: atsdric@cdc.gov **Internet:** <http://atsdr1.atsdr.cdc.gov:8080>

The following additional material can be ordered through the ATSDR Information Center:

Case Studies in Environmental Medicine: Taking an Exposure History-The importance of taking an exposure history and how to conduct one are described, and an example of a thorough exposure history is provided. Other case studies of interest include *Reproductive and Developmental Hazards; Skin Lesions and Environmental Exposures; Cholinesterase-Inhibiting Pesticide Toxicity*; and numerous chemical-specific case studies.

Managing Hazardous Materials Incidents is a three-volume set of recommendations for on-scene (prehospital) and hospital medical management of patients exposed during a hazardous materials incident. Volumes I and II are planning guides to assist first responders and hospital emergency department personnel in planning for incidents that involve hazardous materials. Volume III-Medical *Management Guidelines for Acute Chemical Exposures*-is a guide for health care professionals treating patients exposed to hazardous materials.

Fact Sheets (ToxFAQs) provide answers to frequently asked questions about toxic substances.

Other Agencies and Organizations

The National Center for Environmental Health (NCEH) focuses on preventing or controlling disease, injury, and disability related to the interactions between people and their environment outside the workplace. *Contact:* NCEH, Mailstop F-29,4770 Buford Highway, NE, Atlanta, GA 30341-3724 | Phone: 770-488-7000 | FAX: 770-488-7015.

The National Institute for Occupational Safety and Health (NIOSH) conducts research on occupational diseases and injuries, responds to requests for assistance by investigating problems of health and safety in the workplace, recommends standards to the Occupational Safety and Health Administration (OSHA) and the Mine Safety and Health Administration (MSHA), and trains professionals in occupational safety and health. *Contact:* NIOSH, 200 Independence Avenue, SW, Washington, DC 20201 | Phone: 800-356-4674 or NIOSH Technical Information Branch, Robert A. Taft Laboratory, Mailstop C-19,4676 Columbia Parkway, Cincinnati, OH 45226-1998 | Phone: 800-35-NIOSH.

The National Institute of Environmental Health Sciences (NIEHS) is the principal federal agency for biomedical research on the effects of chemical, physical, and biologic environmental agents on human health and well-being. *Contact:* NIEHS, PO Box 12233,104 T.W. Alexander Drive, Research Triangle Park, NC 27709 | Phone: 919-541-3212.

Referrals

The Association of Occupational and Environmental Clinics (AOEC) has developed a network of clinics in the United States to provide expertise in occupational and environmental issues. *Contact:* AOEC, 1010 Vermont Avenue, NW, #513, Washington, DC 20005 | Phone: 202-347-4976 | FAX: 202-347-4950 | e-mail: aoec@dgs.dgsys.com | AOEC Clinic Director: <http://occ-envmed.mc.duke.edu/oem/aoec.htm>.

The American College of Occupational and Environmental Medicine (ACOEM) is an association of physicians and other health care providers specializing in the field of occupational and environmental medicine. *Contact:* ACOEM, 55 West Seegers Road, Arlington Heights, IL 60005 | Phone: 847-228-6850 | FAX: 847-228-1856.

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THE PROFILE HAS UNDERGONE THE FOLLOWING ATSDR INTERNAL REVIEWS:

1. Health Effects Review. The Health Effects Review Committee examines the health effects chapter of each profile for consistency and accuracy in interpreting health effects and classifying end points.
2. Minimal Risk Level Review. The Minimal Risk Level Workgroup considers issues relevant to substance-specific minimal risk levels (MRLs), reviews the health effects database of each profile, and makes recommendations for derivation of MRLs.
3. Data Needs Review. The Research Implementation Branch reviews data needs sections to assure consistency across profiles and adherence to instructions in the Guidance.

PEER REVIEW

A peer review panel was assembled for 2,4- and 2,6-dinitrotoluene. The panel consisted of the following members:

- 1 . G.A. Shakeel Ansari, Ph.D., Professor, Department of Human Biological Chemistry and Genetics, and Department of Pathology, University of Texas Medical Branch, Galveston, TX.
- 2 . James Klaunig, Ph.D., Professor and Director of Toxicology, Department of Pharmacology and Toxicology, Division of Toxicology, School of Medicine, Indiana University, Indianapolis, IN.
- 3 . Gary Stoner, Ph.D., Professor and Chair, Division of Environmental Health Sciences, School of Public Health, Ohio State University, Columbus, OH.
- 4 . William J. George, Ph.D., Professor and Director of Toxicology, Tulane University School of Medicine, New Orleans, LA.
- 5 . Gerald Kennedy, DuPont/Haskell Lab., Newark, DE.

These experts collectively have knowledge of 2,4- and 2,6-dinitrotoluene's physical and chemical properties, toxicokinetics, key health end points, mechanisms of action, human and animal exposure, and quantification of risk to humans. All reviewers were selected in conformity with the conditions for peer review specified in Section 104(i)(13) of the Comprehensive Environmental Response, Compensation, and Liability Act, as amended.

Scientists from the Agency for Toxic Substances and Disease Registry (ATSDR) have reviewed the peer reviewers' comments and determined which comments will be included in the profile. A listing of the peer reviewers' comments not incorporated in the profile, with a brief explanation of the rationale for their exclusion, exists as part of the administrative record for this compound. A list of databases reviewed and a list of unpublished documents cited are also included in the administrative record.

The citation of the peer review panel should not be understood to imply its approval of the profile's final content. The responsibility for the content of this profile lies with the ATSDR.

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1. PUBLIC HEALTH STATEMENT

This public health statement tells you about 2,4-dinitrotoluene (2,4-DNT) and 2,6-dinitrotoluene (2,6-DNT) and the effects of exposure.

The Environmental Protection Agency (EPA) identifies the most serious hazardous waste sites in the nation. These sites make up the National Priorities List (NPL) and are the sites targeted for long-term federal cleanup activities. 2,4-DNT and 2,6-DNT have been found in at least 122 of the 1,467 current or former NPL sites. However, the total number of NPL sites evaluated for this substance is not known. As more sites are evaluated, the sites at which 2,4-DNT and 2,6-DNT is found may increase. This information is important because exposure to this substance may harm you and because these sites may be sources of exposure.

When a substance is released from a large area, such as an industrial plant, or from a container, such as a drum or bottle, it enters the environment. This release does not always lead to exposure. You are exposed to a substance only when you come in contact with it. You may be exposed by breathing, eating, or drinking the substance or by skin contact.

If you are exposed to 2,4-DNT or 2,6-DNT, many factors determine whether you'll be harmed. These factors include the dose (how much), the duration (how long), and how you come in contact with it. You must also consider the other chemicals you're exposed to and your age, sex, diet, family traits, lifestyle, and state of health.

1.1 WHAT ARE 2,4- AND 2,6-DINITROTOLUENE?

2,4-DNT and 2,6-DNT are pale yellow solids with a slight odor and are two of the six forms of the chemical called dinitrotoluene (DNT). The other four forms (2,3-DNT, 2,5-DNT, 3,4-DNT, and 3,5-DNT) only make up about 5% of the technical grade DNT. DNT is not a natural substance but rather is usually made by reacting toluene (a solvent) with mixed nitric and sulfuric

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acids, which are strong acids. DNT is used to produce flexible polyurethane foams used in the bedding and furniture industry. DNT is also used to produce ammunition and explosives and to make dyes. It is also used in the air bags of automobiles. It has been found in the soil, surface water, and groundwater of at least 122 hazardous waste sites that contain buried ammunition wastes and wastes from manufacturing facilities that release DNT. DNT does not usually evaporate and is found in the air only in manufacturing plants. DNT also does not usually remain in the environment for a long time because it is broken down by sunlight and bacteria into substances such as carbon dioxide, water, and nitric acid. More detailed information on the chemical and physical properties of DNT and its uses is provided in Chapters 3 and 4.

1.2 WHAT HAPPENS TO 2,4- AND 2,6-DINITROTOLUENE WHEN THEY ENTER THE ENVIRONMENT?

DNT can be found in air, surface water, groundwater, and soil. Releases to the air are usually in the form of dusts or aerosols from manufacturing plants. Evaporation from water containing DNT is not a likely means of release to the air. DNT is thought to break down in air by a variety of chemical reactions that take place upon exposure to sunlight.

In water, DNT can be broken down by sunlight. Under conditions without oxygen or without light, DNT may be broken down by biological degradation, whereby microbes utilize the chemical as a source of energy and convert it into chemicals such as carbon dioxide and water. DNT in surface water from rivers and streams and groundwater from wells can result from releases of waste water from trinitrotoluene (TNT) manufacturing facilities and from buried munition wastes.

No information was located regarding the changing of DNT to other chemical substances in soil. DNT is unlikely to build up in animal tissues after animals are exposed by eating impacted soil, water, or vegetation, or by inhaling contaminated air. However, since DNT is quite soluble in water, it can be transferred to plants via root uptake from soil or irrigation with contaminated water, although no direct measurements have been found. It is, however, expected to accumulate readily in plant materials, although no direct measurements have been found.

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For more information on what happens to DNT in the environment, see Chapters 4 and 5.

1.3 HOW MIGHT I BE EXPOSED TO 2,4- AND 2,6-DINITROTOLUENE?

Members of the general population are likely to be exposed only if they are near a DNT contaminated waste site or manufacturing facilities that release DNT.

2,4- or 2,6-DNT may enter the environment from waste waters that industries discharge into rivers and streams or from the improper disposal of wastes. However, regular testing of water in the United States shows that 2,4- and 2,6-DNT were found in less than 2% of the water samples. Testing of hazardous waste sites shows that 2,4- and 2,6-DNT are present at less than 8.5% of these sites. Available information indicates that DNT does not appear to be widespread in the environment. More information on how people might be exposed to 2,4- or 2,6-DNT is given in Chapter 5.

1.4 HOW CAN 2,4- AND 2,6-DINITROTOLUENE ENTER AND LEAVE MY BODY?

When industrial workers are exposed to 2,4- or 2,6-dinitrotoluene, the major ways that these chemicals enter their bodies are by breathing or absorbing small amounts of the chemical through the skin. Some ingestion may also occur as the result of eating or smoking without prior handwashing.

After individuals breathe air, drink water, or eat food contaminated with 2,4- or 2,6-DNT, these chemicals are changed into different substances by the liver and in the intestines. After this, most of these chemicals leave the body within 24 hours in the urine, with a small amount in the feces. This information comes from animal experiments done in laboratories and from studies of industrial workers. More detailed information is given in Chapter 2.

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1.5 HOW CAN 2,4- AND 2,6-DINITROTOLUENE AFFECT MY HEALTH?

To protect the public from the harmful effects of toxic chemicals and to find ways to treat people who have been harmed, scientists use many tests.

One way to see if a chemical will hurt people is to learn how the chemical is absorbed, used, and released by the body; for some chemicals, animal testing may be necessary. Animal testing may also be used to identify health effects such as cancer or birth defects. Without laboratory animals, scientists would lose a basic method to get information needed to make wise decisions to protect public health. Scientists have the responsibility to treat research animals with care and compassion. Laws today protect the welfare of research animals, and scientists must comply with strict animal care guidelines.

Increases in death rate due to heart disease have been seen in workers exposed to 2,4-DNT or technical grade DNT (Tg-DNT), but these workers may also have been exposed to other chemicals. 2,4- and 2,6-DNT may also affect the nervous system and the blood of exposed workers. One study showed that male workers exposed to 2,4- and 2,6-DNT had reduced levels of sperm, but later studies did not confirm the finding.

Exposure to high levels of these compounds in animals regularly causes lowered numbers of sperm and reduced fertility. Studies of animals have also shown that nervous system disorders, liver damage, and kidney damage can occur, as well as a reduction in the numbers of red blood cells. Both 2,4- and 2,6-DNT can cause liver cancer in laboratory rats and may produce the same effect in humans. More information on the health effects of DNT is given in Chapter 2. The International Agency for Research on Cancer (IARC) has determined that 2,4- and 2,6-DNT are possibly carcinogenic to humans.

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1.6 HOW CAN 2,4- AND 2,6-DINITROTOLUENE AFFECT CHILDREN?

This section discusses potential health effects from exposures during the period from conception to maturity at 18 years of age in humans. Potential effects on children resulting from exposures of the parents are also considered.

2,4- and 2,6-DNT are not widespread throughout the environment. If you do not live near a plant or a waste site that contains DNT, it is unlikely that your children would be exposed to DNT. DNT is normally associated with industrial or military production plants or munitions storage sites. DNT is water soluble, so if contamination has occurred it will usually be carried in water. Children are at risk for exposure if DNT has leached into a community's drinking water supply from a nearby hazardous site since they drink more fluid in proportion to their body weight than adults. Children playing in DNT-contaminated surface water might be more exposed than adults, both because of this behavior and because of their larger skin area in proportion to their body weight.

Since DNT exposure is usually related to adult workers, health effects on children have not been studied. It is not known if DNT affects children differently than adults, or what long term effects might appear in adults exposed as children.

No studies have investigated effects of DNT on the developmental process in humans, and few studies have focused on animals. No studies have been done to see if DNT or its toxic breakdown products cross the placenta, or get into breast milk.

1.7 HOW CAN FAMILIES REDUCE THE RISK OF EXPOSURE TO 2,4- AND 2,6-DINITROTOLUENE?

If your doctor finds that you have been exposed to significant amounts of 2,4-DNT or 2,6-DNT, ask if children may also be exposed. When necessary your doctor may need to ask your State Department of Public Health to investigate.

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If you live near a site that could be contaminated with DNT you should discourage your children from putting foreign objects, groundwater, and dirt in their mouths since DNT is quite water soluble. Make sure they wash their hands frequently and before eating. Discourage your children both from putting their hands in their mouths and from other hand-to-mouth activities.

1.8 IS THERE A MEDICAL TEST TO DETERMINE WHETHER I HAVE BEEN EXPOSED TO 2,4- AND 2,6-DINITROTOLUENE?

Both 2,4- and 2,6-DNT and the chemicals they are changed into by the body can be measured in the blood and urine of exposed individuals (if urine is collected within 24 hours). The tests cannot show how much 2,4- or 2,6-DNT an individual has been exposed to. These tests are not usually available in doctors' offices but can be performed by special laboratories. More detailed information on medical tests is presented in Chapter 6.

1.9 WHAT RECOMMENDATIONS HAS THE FEDERAL GOVERNMENT MADE TO PROTECT HUMAN HEALTH?

The federal government develops regulations and recommendations to protect public health. Regulations can be enforced by law. Federal agencies that develop regulations for toxic substances include the Environmental Protection Agency (EPA), the Occupational Safety and Health Administration (OSHA), and the Food and Drug Administration (FDA). Recommendations provide valuable guidelines to protect public health but cannot be enforced by law. Federal organizations that develop recommendations for toxic substances include the Agency for Toxic Substances and Disease Registry (ATSDR) and the National Institute for Occupational Safety and Health (NIOSH).

Regulations and recommendations can be expressed in not-to-exceed levels in air, water, soil, or food that are usually based on levels that affect animals; then they are adjusted to help protect people. Sometimes these not-to-exceed levels differ among federal organizations because of different exposure times (an 8-hour workday or a 24-hour day), the use of different animal studies, or other factors.

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Recommendations and regulations are also periodically updated as more information becomes available. For the most current information, check with the federal agency or organization that provides it. Some regulations and recommendations for 2,4- and 2,6-DNT include the following:

EPA recommendations define the safe lifetime daily maximum oral dose for 2,4-DNT as 0.002 milligram per kilogram per day for 2,4-DNT, and 0.001 milligram per kilogram per day for 2,6-DNT. In addition to EPA recommendations, OSHA regulations state that an average 8-hour exposure to total DNT in workplace air should not be more than 1.5 milligrams per cubic meter of air (mg/m³). NIOSH has published a Recommended Exposure Limit (REL) guideline of 1.5 mg/m³. An REL is an average concentration for a 10-hour workday over a 40-hour workweek. Spills or releases to the environment of more than 1,000 pounds of DNT must be reported immediately to the federal government. More information on regulations and advisories for DNT is given in Chapter 7.

1.10 WHERE CAN I GET MORE INFORMATION?

If you have any more questions or concerns, please contact your community or state health or environmental quality department or

Agency for Toxic Substances and Disease Registry
Division of Toxicology
1600 Clifton Road NE, Mailstop E-29
Atlanta, GA 30333

* Information line and technical assistance

Phone: 1-800-447-1544
Fax: (404) 639-6359

ATSDR can also tell you the location of occupational and environmental health clinics. These clinics specialize in recognizing, evaluating, and treating illnesses resulting from exposure to hazardous substances.

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* To order toxicological profiles, contact

National Technical Information Service
5285 Port Royal Road
Springfield, VA 22161
Phone: (800) 553-6847 or (703) 487-4650

2. HEALTH EFFECTS

2.1 INTRODUCTION

The primary purpose of this chapter is to provide public health officials, physicians, toxicologists, and other interested individuals and groups with an overall perspective of the toxicology of 2,4- and 2,6-DNT. It contains descriptions and evaluations of toxicological studies and epidemiological investigations and provides conclusions, where possible, on the relevance of toxicity and toxicokinetic data to public health.

A glossary and list of acronyms, abbreviations, and symbols can be found at the end of this profile.

2.2 DISCUSSION OF HEALTH EFFECTS BY ROUTE OF EXPOSURE

To help public health professionals and others address the needs of persons living or working near hazardous waste sites, the information in this section is organized first by route of exposure-inhalation, oral, and dermal; and then by health effect-death, systemic, immunological, neurological, reproductive, developmental, genotoxic, and carcinogenic effects. These data are discussed in terms of three exposure periods-acute (14 days or less), intermediate (15-364 days), and chronic (365 days or more).

Levels of significant exposure for each route and duration are presented in tables and illustrated in figures. The points in the figures showing no-observed-adverse-effect levels (NOAELs) or lowest-observed-adverse effect levels (LOAELs) reflect the actual doses (levels of exposure) used in the studies. LOAELs have been classified into “less serious” or “serious” effects. “Serious” effects are those that evoke failure in a biological system and can lead to morbidity or mortality (e.g., acute respiratory distress or death). “Less serious” effects are those that are not expected to cause significant dysfunction or death, or those whose significance to the organism is not entirely clear. ATSDR acknowledges that a considerable amount of judgment may be required in establishing whether an end point should be classified as a NOAEL, “less serious” LOAEL, or “serious” LOAEL, and that in some cases, there will be insufficient data to decide whether the effect is indicative of significant dysfunction. However, the Agency has established guidelines and policies that are used to classify these end points. ATSDR believes that there is sufficient merit in this approach to warrant an attempt at distinguishing between “less serious” and “serious” effects. The distinction between “less serious” effects and “serious” effects is considered to be important because it helps the users of the profiles to identify levels of exposure at which major health effects start to appear. LOAELs or NOAELs should also help in

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determining whether or not the effects vary with dose and/or duration, and place into perspective the possible significance of these effects to human health.

The significance of the exposure levels shown in the Levels of Significant Exposure (LSE) tables and figures may differ depending on the user's perspective. Public health officials and others concerned with appropriate actions to take at hazardous waste sites may want information on levels of exposure associated with more subtle effects in humans or animals (LOAEL) or exposure levels below which no adverse effects (NOAELs) have been observed. Estimates of levels posing minimal risk to humans (Minimal Risk Levels or MRLs) may be of interest to health professionals and citizens alike.

Levels of exposure associated with carcinogenic effects (Cancer Effect Levels, CELs) of 2,4-DNT are indicated in Table 2-1. Because cancer effects could occur at lower exposure levels, Figure 2-1 also shows a range for the upper bound of estimated excess risks, ranging from a risk of 1 in 10,000 to 1 in 10,000,000 (10^{-4} to 10^{-7}), as developed by EPA.

Estimates of exposure levels posing minimal risk to humans (Minimal Risk Levels or MRLs) have been made for 2,4- and 2,6-DNT. An MRL is defined as an estimate of daily human exposure to a substance that is likely to be without an appreciable risk of adverse effects (noncarcinogenic) over a specified duration of exposure. MRLs are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration within a given route of exposure. MRLs are based on noncancerous health effects only and do not consider carcinogenic effects. MRLs can be derived for acute, intermediate, and chronic duration exposures for inhalation and oral routes. Appropriate methodology does not exist to develop MRLs for dermal exposure.

Although methods have been established to derive these levels (Barnes and Dourson 1988; EPA 1990), uncertainties are associated with these techniques. Furthermore, ATSDR acknowledges additional uncertainties inherent in the application of the procedures to derive less than lifetime MRLs. As an example, acute inhalation MRLs may not be protective for health effects that are delayed in development or are acquired following repeated acute insults, such as hypersensitivity reactions, asthma, or chronic bronchitis. As these kinds of health effects data become available and methods to assess levels of significant human exposure improve, these MRLs will be revised.

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A User's Guide has been provided at the end of this profile (see Appendix B). This guide should aid in the interpretation of the tables and figures for Levels of Significant Exposure and the MRLs.

Although the primary foci of this document are 2,4-DNT and 2,6-DNT, some information on technical grade DNT (Tg-DNT) is also provided. Tg-DNT contains approximately 76% 2,4-DNT, 19% 2,6-DNT, and other isomers. Unless specified otherwise, 2,4-DNT and 2,6-DNT are abbreviated generically as DNT in this document.

2.2.1 Inhalation Exposure

Most of the data on human health effects associated with exposure to 2,4-DNT or 2,6-DNT are derived from studies of workers in occupational settings. Exposure monitoring of workers in the past has generally been inadequate. Consequently, few dose-response data based on human exposure to 2,4- or 2,6-DNT are available.

Human exposure to chemicals in an occupational setting can occur via multiple routes: inhalation, dermal, and inadvertent ingestion (Hamill et al. 1982). Although the low vapor pressure of DNT makes inhalation of vapors unlikely, it can occur when contaminated particulate material is in the air. In addition, some dermal exposure is probable, and some ingestion may also occur as the result of eating or smoking without prior handwashing.

2.2.1.1 Death

In a retrospective cohort mortality study of 457 munitions workers who were exposed to either 2,4-DNT or Tg-DNT at 2 geographically different U.S. manufacturing plants, significant increases in death rates due to ischemic heart disease and residual diseases of the circulatory system were found (Standard Mortality Rates [SMR] of 126 and 143; 95% confidence intervals [CI] of 65-234 and 112-179, respectively) (Levine et al. 1986a). Residual diseases of the circulatory system include congestive heart failure, cardiac arrest, and arteriosclerosis. The workers had been exposed to unreported concentrations of either 2,4-DNT (98% pure) or Tg-DNT for periods ranging from 30 days to more than 5 years (Levine et al. 1986a). Cigarette smoking was not taken into account in this study, but the study authors suggested that it may not have been a risk factor because mortality from lung cancer was less than expected. Among workers at both plants, there appeared to be a latency period of more than 15 years for a significant increase in mortality due to ischemic

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heart disease. There also appeared to be a relationship between heart disease and the intensity of exposure to dinitrotoluenes. No statistical increase was found in death due to cancer, either from malignant neoplasms as a whole or from individual cancers, although the statistical power of the study was insufficient to detect anything but gross changes in the death rate due to cancer.

The Levine et al. (1986a) retrospective cohort mortality study was Limited by small cohort size, and thus, the study had diminished power to detect an effect. As a result, the finding of elevated mortality from heart disease among workers in two plants from different parts of the United States linked only by exposure to DNT is unusual. Workers in the United States generally have lower rates of heart disease than the general population because of the “healthy worker effect.” At both plants, mortality from ischemic heart disease during the first 15 years following cohort entry was less than expected, and mortality increased only in later years. Suggestive, but not significant, is evidence of a relationship between heart disease and duration and intensity of exposure, also reported by Levine et al. (1986a).

No studies were located regarding death in animals after inhalation exposure to 2,4- or 2,6-DNT.

2.2.1.2 Systemic Effects

No studies were located regarding respiratory, dermal, or ocular effects in humans or animals after inhalation exposure to 2,4- or 2,6-DNT.

Cardiovascular Effects. As described in Section 2.2.1.1, Levine et al. (1986a) reported a significant increase in heart disease mortality in workers involved in the manufacture and processing of 2,4-DNT and/or Tg-DNT.

No studies were located regarding cardiovascular effects in animals after inhalation exposure to 2,4- or 2,6-DNT.

Gastrointestinal Effects. Vomiting and nausea were mentioned as health complaints in a survey of 154 male workers involved in the production of smokeless gunpowders during World War II (McGee et al. 1942). The exposure concentrations of 2,4-DNT were not specified but may have been relatively high because of the lack of modern industrial hygiene practices. These symptoms were still mentioned in a followup survey of workers that was undertaken at the same facility after the installation of safeguards to reduce

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DNT exposure (McGee et al. 1947). Since exposure to other compounds cannot be ruled out, attribution of these symptoms to DNT cannot be verified.

No studies were located regarding gastrointestinal effects in animals after inhalation exposure to 2,4- or 2,6-DNT.

Hematological Effects. Several hematological effects, including anemia and cyanosis, were found in male workers employed by a munitions factory during World War II (McGee et al. 1942, 1947). In some cases there were increases in leukocyte count, which may be related to prolonged exposure to DNT. The study authors presumed that the exposure concentrations to 2,4-DNT were relatively high because of the relatively primitive industrial hygiene practices at that time. Although 36 of 154 workers were anemic in the earlier study and 73 of 714 workers were anemic in the follow-up study, no control groups were used as a basis for comparison. Because of possible exposure to other compounds, the lack of work histories, lack of exposure monitoring, lack of a control population, and small cohort size, the results obtained are equivocal and may be best used as qualitative descriptions of symptoms. Marked cyanosis and other incapacitating symptoms were reported after exposure to unspecified concentrations of Tg-DNT in a study of French workers in a DNT production plant during World War I (Perkins 1919). It is assumed that workers were exposed to high concentrations of Tg-DNT via both inhalation and dermal pathways, since the processes described involved direct handling of large amounts of Tg-DNT without protective equipment.

No studies were located regarding hematological effects in animals after inhalation exposure to 2,4- or 2,6-DNT.

Musculoskeletal Effects. Men who worked in a munitions plant during World War II mentioned muscular weakness as a complaint in a survey of these workers (McGee et al. 1942). However, the lack of 2,4-DNT exposure data and lack of an adequate control population prevent these data from being useful for anything other than qualitative description. Joint pain, especially in the knees, and other incapacitating symptoms were found in unspecified numbers of French workers in a plant that produced DNT during World War I (Perkins 1919). No exposure concentrations were reported, but it is assumed that they were high because of the direct handling of large amounts of Tg-DNT without protective equipment, which also suggests that the workers were exposed dermally. However, because exposure to other compounds cannot be ruled out and no control data are available, caution must be used when interpreting these results.

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No studies were located regarding musculoskeletal effects in animals after inhalation exposure to 2,4- or 2,6-DNT.

Hepatic Effects. Two of 154 workers at a munitions plant suffered from acute hepatitis with jaundice but recovered soon after they were no longer exposed to 2,4-DNT (McGee et al. 1942). Although the exposure concentrations were not known, they were assumed to be high based upon the industrial hygiene practices prevalent at that time. Furthermore, since an adequate control population was not used, this study provides only a qualitative description of symptoms, at best. A later study of 714 workers at this same plant found that 29 experienced liver tenderness (McGee et al. 1947). This incidence would indicate an increased effect on the liver from the 1942 study, despite improvements in industrial hygiene and engineering practices designed to decrease worker exposure in the interval between the two studies. Other factors, such as alcohol consumption, may account for these results which should be viewed with caution because of the lack of control data, lack of information on exposure concentrations, and possible multiple chemical exposure.

Medical surveys of 52 male workers exposed to Tg-DNT in a chemical plant that manufactured toluenediamine (TDA) revealed no differences in hepatic blood chemistry profiles (Ahrenholz and Meyer 1982). Air samples contained concentrations ranging from 0.026 to 0.890 mg/m³ Tg-DNT (mean 0.207 mg/m³ Tg-DNT).

No studies were located regarding hepatic effects in animals after inhalation exposure to 2,4- or 2,6-DNT.

Renal Effects. No effects were observed on either of the renal parameters (BUN, creatinine) monitored in blood chemistry in a medical survey of 52 male workers exposed to Tg-DNT in a chemical plant that manufactured TDA (Ahrenholz and Meyer 1982). Exposure concentrations in air samples taken for this study ranged from 0.026 to 0.890 mg/m³ Tg-DNT (mean 0.207 mg/m³ Tg-DNT). The study was limited by a small exposure population and lack of historical individual exposure monitoring.

No studies were located regarding renal effects in animals after inhalation exposure to 2,4- or 2,6-DNT.

2.2.1.3 Immunological and Lymphoreticular Effects

No studies were located regarding immunological or lymphoreticular effects in humans or animals after inhalation exposure to 2,4- or 2,6-DNT.

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2.2.1.4 Neurological Effects

Dizziness and headache were reported by Perkins (1919) in a study of French workers exposed to Tg-DNT at a production plant during World War I. Although no exposure concentrations were reported, it is assumed that the workers were exposed to high concentrations of Tg-DNT via both inhalation and dermal pathways, since the manufacturing processes required workers to handle large amounts of Tg-DNT without protective equipment. Exposure to chemicals other than DNT in this environment could not be ruled out. Health effects of munitions workers exposed to unspecified levels of what was presumed to be 2,4-DNT were studied by McGee et al. (1942,1947). Neurological signs reported by these workers included headache, dizziness, insomnia, unpleasant taste in the mouth, and pain, numbness, and tingling in the extremities. The 2,4-DNT exposure concentrations were not specified but were considered by these authors to be relatively high as a result of the lack of safety practices.

No studies were located regarding neurological effects in animals after inhalation exposure to 2,4- or 2,6-DNT.

2.2.1.5 Reproductive Effects

Studies of men occupationally exposed to Tg-DNT at DNT and TDA plants showed no significant differences in sperm counts or morphology, follicle stimulating hormone (FSH) levels, or in the incidence of miscarriage in their wives compared to controls (Ahrenholz and Meyer 1982; Hamill et al. 1982). In the Ahrenholz and Meyer (1982) study, DNT concentrations ranged from 0.026 to 0.890 mg/m³ (mean 0.207 mg/m³ Tg-DNT). Interpretation of these studies is somewhat confounded by the lack of distinction between DNT and TDA exposure and by the lack of information regarding exposure concentration in the Hamill et al. (1982) study. The limitations of these studies are similar (small exposure populations and the lack of individual exposure monitoring) and limit the ability of the studies to detect adverse effects.

No significant effects on the fertility of workers occupationally exposed to Tg-DNT have been found in several studies (Ahrenholz and Meyer 1982; Hamill et al. 1982; Levine et al. 1985a). However, Levine et al. (1985a) estimate that only a 50-70% reduction in fertility could have been detected in the worker population that they studied.

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One study by the CDC (1981) noted that sperm counts were decreased by more than 50% in workers in a Kentucky chemical plant exposed to DNT and TDA compared to workers unexposed to these chemicals. The study was limited because of multiple chemical exposures and the small numbers of workers examined. Thirty workers participated in the study: 9 currently exposed, 12 previously exposed, and 9 with no history of exposure to DNT/TDA.

No studies were located regarding reproductive effects in animals after inhalation exposure to 2,4-DNT or 2,6-DNT.

2.2.1.6 Developmental Effects

No studies were located regarding developmental effects in humans or animals after inhalation exposure to 2,4- or 2,6-DNT.

2.2.1.7 Genotoxic Effects

No studies were located regarding genotoxic effects in humans or animals after inhalation exposure to 2,4- or 2,6-DNT.

Genotoxicity studies are discussed in Section 2.5.

2.2.1.8 Cancer

The mortality of a cohort of 4,989 men who worked at least 5 months in a munitions facility was analyzed to determine whether DNT exposure was associated with an increased risk of cancer of the liver and biliary tract (Stayner et al. 1993). Workers were considered exposed if they had worked at least 1 day on a job with probable exposure to DNT. In this study, a significant increase in hepatobiliary cancer mortality (standard rate ratio [SRR] = 3.88, 95% CI 1.04, 14.41) was observed among DNT-exposed workers compared to unexposed control workers. However, no significant changes were noted when compared to the U.S. population, the SRR for hepatobiliary cancer being 2.67 (95% CI = 0.98, 5.83; $p = 0.052$). No quantitative data were available on the DNT exposure of these men. This study is limited by the small numbers of hepatobiliary cancer cases, small numbers of workers with long exposure to DNT, and possible exposure of the workers to other chemicals. However, no significant increases in mortality from malignant neoplasms as

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a group or from particular cancers (liver, lung, gallbladder, kidney, and connective tissues) were observed in workers occupationally exposed to 2,4-DNT and/or Tg-DNT (Levine et al. 1986b). Exposures were not quantified and again the cohort was small. The study authors estimated that an 8-fold increase in liver and gallbladder cancer in exposed workers would be necessary in order to be detected at the $p = 0.05$ level, thus the statistical analysis was not strong enough to detect small increases in cancer.

No studies were located regarding cancer in animals following inhalation exposure to 2,4- or 2,6-DNT.

2.2.2 Oral Exposure

No studies were located regarding health effects in humans following oral exposure to 2,4- or 2,6-DNT. However, it is assumed that oral ingestion could be a secondary route for occupationally exposed humans.

2.2.2.1 Death

No studies were located regarding death in humans after oral exposure to 2,4- or 2,6-DNT.

2,4-DNT is lethal to experimental animals after oral administration. Animals generally developed cyanosis and ataxia after dosing. In general, rats are more sensitive than mice to the lethal effects of 2,4-DNT. The LD₅₀s that have been determined for rats after gavage dosing with 2,4-DNT range from 270 to 650 mg/kg (Ellis et al. 1978; Lee et al. 1975; Vernot et al. 1977); in mice, LD₅₀s were reported to be between 1,340 and 1,954 mg/kg after 2,4-DNT administration (Ellis et al. 1978; Lee et al. 1975; Vernot et al. 1977). In a dominant lethal study by Lane et al. (1985), 8 of 15 male Sprague-Dawley rats died after receiving 5 daily doses of 240 mg/kg 2,4-DNT. No deaths were reported when male and female Sprague-Dawley rats were fed 78 or 82 mg/kg/day 2,4-DNT, respectively, in the diet for 14 days (McGown et al. 1983).

Death has been reported after intermediate- and chronic-duration exposure to 2,4-DNT in numerous studies. One of 8 male and 8 of 8 female CD rats died after 3-13 weeks of ingesting 2,4-DNT in the diet (Lee et al. 1978, 1985). Concentrations in the feed causing these deaths were equivalent to doses of 93 and 145 mg/kg/day in males and females, respectively. Death has also been reported in rodents fed concentrations equivalent to doses of 347-413 mg/kg 2,4-DNT in the diet for up to 6 months (Hong et al. 1985; Kozuka et al. 1979; Lee et al. 1978). No treatment-related deaths were reported in rats fed up to 16.5 mg/kg/day 2,4-DNT or mice fed up to 28.5 mg/kg/day 2,4-DNT for 4 weeks, or in rats fed up to

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22 mg/kg/day 2,4-DNT or mice fed up to 76 mg/kg/day 2,4-DNT for 78 weeks (NCI 1978). In a 3-generation reproductive study, there appeared to be an increased incidence of death among F₁ dams during parturition after receiving 45.3 mg/kg/day 2,4-DNT in the diet for 6 months (Ellis et al. 1979). These deaths were associated with prolonged parturition, hemorrhage, and placental retention. However, because these effects were also seen to a lesser extent in control animals, it may be that the effects of 2,4-DNT simply enhanced effects caused by the advancing age of the dams (Ellis et al. 1979).

In a 13-week study, some dogs fed 25 mg/kg/day became moribund after 22 or more days and had to be terminated, whereas no treatment-related deaths were reported in dogs fed 5 mg/kg/day (Ellis et al. 1985; Lee et al. 1978). In addition to severe weight loss, severe neurological effects and histopathological changes were found in these animals, including vacuolization and focal gliosis in the cerebellum and perivascular hemorrhages in the cerebellum and brain stem, as well as peripheral neuropathy, testicular degeneration, and biliary hyperplasia. In a 24-month study of dogs, the administration of 10 mg/kg/day 2,4-DNT by capsule caused death within 6 months, but no deaths were reported at 1.5 mg/kg/day; clinical signs prior to death were similar to those reported in the 13-week study (Ellis et al. 1979, 1985). Decreased longevity was reported in 1-2-year studies of CD rats at average daily intakes as low as 3.9 mg/kg/day (males) and 5.1 mg/kg/day (females), and of CD-l mice at 898 mg/kg/day (Ellis et al. 1979; Hong et al. 1985; Lee et al. 1978, 1985).

The experimental data are more limited for 2,6-DNT than for 2,4-DNT. After administration of 2,6-DNT, LD₅₀s have been reported to range from 180 to 795 mg/kg in rats and from 621 to 807 mg/kg in mice (Ellis et al. 1978; Lee et al. 1975; Vernot et al. 1977). The maximum tolerated dose (MTD) of 2,6-DNT corresponding to 100% survival of A/J mice after 6 doses over a 2-week period was 250 mg/kg (Schut et al. 1983).

Intermediate-duration studies have also shown an increase in mortality of mice and dogs after 2,6-DNT administration. After feeding 51 mg/kg/day 2,6-DNT to male Swiss albino mice in the diet for up to 13 weeks, 8 of 16 of these animals died; 6 of 16 females fed 55 mg/kg/day 2,6-DNT also died (Lee et al. 1976). No treatment-related deaths were reported when rats were fed up to 155 mg/kg/day 2,6-DNT for the same duration (Lee et al. 1976). Two of 8 dogs treated with 20 mg/kg 2,6-DNT by capsule died in a 13-week study (Lee et al. 1976). Thus, dogs seem to be the most sensitive of the three species to intermediate-duration oral 2,6-DNT exposure.

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Administration of up to 150 mg/kg/day Tg-DNT for 14 days was lethal to 6 of 13 pregnant Fischer-344 rats when administered by gavage during gestation (Jones-Price et al. 1982), yet this same concentration of Tg-DNT fed in the diet for 30 days did not kill any of the same strain of rats in another study (Hazleton Laboratories 1977). Decreased survival was found in CDF rats fed 35 mg/kg/day Tg-DNT for 52 weeks or 14 mg/kg/day Tg-DNT for 104 weeks (Hazleton Laboratories 1982).

For 2,4-DNT, all LOAEL values from each reliable study for death in each species and duration category are recorded in Table 2-1 and plotted in Figure 2-1. For 2,6-DNT, all LOAEL values from each reliable study for death in each species and duration category are recorded in Table 2-2 and plotted in Figure 2-2.

2.2.2.2 Systemic Effects

No studies were located regarding musculoskeletal effects in humans or animals after oral exposure to 2,4-DNT or 2,6-DNT. The systemic effects observed after oral exposure are discussed below.

For 2,4-DNT, the highest NOAEL values and all LOAEL values from each reliable study for systemic effects in each species and duration category are recorded in Table 2-1 and plotted in Figure 2-1. For 2,6-DNT, all LOAEL values from each reliable study for systemic effects in each species and duration category are recorded in Table 2-2 and plotted in Figure 2-2.

Respiratory Effects. No studies were located regarding respiratory effects in humans after oral exposure to 2,4-DNT or 2,6-DNT.

No histopathological effects on the lungs were found when Sprague-Dawley rats were fed 261 mg/kg/day (males) or 273 mg/kg/day (females) 2,4-DNT for 14 days (McGown et al. 1983).

No respiratory system effects were observed when CDF rats were fed 14 mg/kg/day Tg-DNT for 2 years (Hazleton Laboratories 1982). Histopathological examination of the lungs and respiratory tract tissues of rats exposed to 14 mg/kg/day Tg-DNT for 2 years or 35 mg/kg/day for 1 year did not reveal any abnormalities (Hazleton Laboratories 1982).

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference					
					Less serious (mg/kg/day)	Serious (mg/kg/day)						
ACUTE EXPOSURE												
Death												
1	Rat (Sprague- Dawley)	5 d 1x/d (GO)				240 M (8/15 died)	Lane et al. 1985					
2	Rat (CD)	once (GO)				568 M (LD ₅₀) 650 F (LD ₅₀)	Lee et al. 1975; Ellis et al. 1978					
3	Rat (Sprague- Dawley)	once (G)				270M (LD ₅₀)	Vernot et al. 1977					
4	Mouse (Swiss albino)	once (GO)				1,954 M (LD ₅₀) 1,340 F (LD ₅₀)	Lee et al. 1975					
5	Mouse (CF-1)	once (G)				1,630 M (LD ₅₀)	Vernot et al. 1977					
Systemic												
6	Rat (Sprague- Dawley)	5 d 1x/d (GO)	Hemato Bd Wt	180M	60M (slight cyanosis) 240M (weight loss)		Lane et al. 1985					

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
7	Rat (Sprague-Dawley)	14d ad lib	Resp	260.9 M 272.7 F			McGown et al. 1983
			Cardio	260.9 M 272.7 F			
			Gastro	260.9 M 272.7 F			
			Hepatic		78.3 M (increased alanine aminotransferase and cholesterol) 81.8 F (increased cholesterol)		
			Renal		78.3 M (hyaline droplet formation) 81.8 F (hyaline droplet formation)		
			Dermal	260.9 M 272.7 F			
			Ocular	260.9 M 272.7 F			
Immunological/Lymphoreticular							
8	Rat (Sprague-Dawley)	14d ad lib		260.9 M 272.7 F			McGown et al. 1983
Neurological							
9	Dog (Beagle)	12d 1x/d (C)		5 ^b		25 (incoordination, stiffness, abnormal gait)	Ellis et al. 1985; Lee et al. 1978

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
Reproductive							
10	Rat (Sprague- Dawley)	5 d 1x/d (GO)		60M		180M (decreased fertility)	Lane et al. 1985
11	Rat (Sprague- Dawley)	14d ad lib				78.3M (decreased thickness of spermatogenic sperm layers)	McGown et al. 1983
12	Mouse DBA/2J	2 d 1x/d (G)				250M (decreased fertility)	Soares and Lock 1980

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference					
					Less serious (mg/kg/day)	Serious (mg/kg/day)						
INTERMEDIATE EXPOSURE												
Death												
13	Rat (CD)	3 or 6 mo ad lib (F)			45.3 F	(increased incidence of death during parturition)	Ellis et al. 1979					
14	Rat (Wistar)	6 mo ad lib (F)			347 M	(71% died)	Kozuka et al. 1979					
15	Rat (CD)	4 or 13 wk ad lib (F)			93 M	(1/8 died)	Lee et al. 1985					
					145 F	(8/8 died)						
16	Mouse (CD-1)	4 or 13 wk ad lib (F)			413	(2/16M, 2/16F died))	Hong et al. 1985; Lee et al. 1978					
17	Dog (Beagle)	6 mo 1x/d (C)			10 M	(4/6 died)	Ellis et al. 1979, 1985					
18	Dog (Beagle)	4 or 13 wk 1x/d (C)			25	(5/8 died)	Ellis et al. 1985; Lee et al. 1978					

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
Systemic							
19	Rat (Sprague- Dawley)	3 wk ad lib (F)	Bd Wt	76.7M	156.4M (10% decrease body weight)		Bloch et al. 1988
20	Rat (CD)	3 or 6 mo ad lib	Bd Wt			34.5M (23-25% decrease in body weight)	Ellis et al. 1979
						45.3 F (10-23% decrease in body weight)	
21	Rat (Wistar)	6 mo ad lib (F)	Hemato		347M (increased methemoglobin)		Kozuka et al. 1979
			Hepatic		347M (increased relative liver weight; increased SGOT, LDH, alkaline phosphatase, acid phosphatase, triglycerides, glucose; formation of puruloid matter)		
			Renal	347M			
			Bd Wt			347M (41% decrease body weight)	

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
22	Rat (CD)	4 or 13 wk ad lib (F)	Hemato	34 M 38 F	93 M (reticulocytosis; hemosiderosis) 108 F (reticulocytosis; hemosiderosis)	266 M (anemia) 145 F (anemia)	Lee et al. 1978, 1985
			Hepatic	266 M 145 F			
			Renal	266 M 145 F			
			Bd Wt			34 M (75% decrease body weight gain with decreased food consumption) 38 F (94% decrease body weight gain with decreased food consumption)	
23	Rat (Fischer- 344)	6 or 26 wk ad lib (F)	Bd Wt		27M (11% decrease body weight)		Leonard et al. 1987
24	Mouse (CD-1)	4 or 13 wk ad lib (F)	Hemato	137 M 147 F	413M (mild anemia, 468 F reticulocytosis)		Hong et al. 1985; Lee et al. 1978
			Hepatic	47 M 147 F		137M (mild hepatocellular 468 F dysplasia)	
			Renal	413 M 468 F			
			Bd Wt			413M (body weight loss with 468 F decreased food consumption)	

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
25	Dog (Beagle)	4 or 13 wk 1x/d (C)	Hemato	5		25 (anemia, Heinz bodies)	Ellis et al. 1985; Lee et al. 1978
			Hepatic	25			
			Renal	25			
Immunological/Lymphoreticular							
26	Rat (Wistar)	6 mo ad lib (F)			347M (increased relative spleen weight)		Kozuka et al. 1979
27	Dog (Beagle)	4 or 13 wk 1x/d (C)		25			Ellis et al. 1985; Lee et al. 1978
Neurological							
28	Rat (Wistar)	6 mo ad lib (F)				347M (humpback incoordination)	Kozuka et al. 1979
29	Rat (CD)	4 or 13 wk ad lib (F)		34 M		93M (demyelination of cerebellum and brain stem)	Lee et al. 1978, 1985
				108 F		(widespread and stiff-legged 145 F gait)	
30	Mouse (CD-1)	4 or 13 wk ad lib (F)		413 M			Hong et al. 1985;
				468 F			Lee et al. 1978

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
31	Dog (Beagle)	4 or 13 wk 1x/d (C)		5		25 (incoordination, abnormal gait, paralysis)	Ellis et al. 1985; Lee et al. 1978
Reproductive							
32	Rat (Sprague-Dawley)	3 wk ad lib (F)		76.7M (multinucleated spermatids, mild irregularity of basal lamina, vacuolation and lipid accumulation in Sertoli cells)		153.4M (extensive degeneration of spermatids and spermatocytes; ultrastructural changes in Sertoli cells; 63% decrease sperm count)	Bloch et al. 1988
33	Rat (CD)	3 or 6 mo ad lib (F)			34.5M 45.3 F	(decreased fertility; difficult parturition)	Ellis et al. 1979
34	Rat (CD)	3 or 6 mo ad lib (F)		34.5M		45M (decreased implantation index; severe atrophy/degeneration of seminiferous tubules)	Ellis et al. 1979
35	Rat (Wistar)	6 mo ad lib (F)			347M	(testicular atrophy)	Kozuka et al. 1979
36	Rat (CD)	4 or 13 wk ad lib (F)		34 M		93M (severe decrease in spermatogenesis)	Lee et al. 1978, 1985

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
37	Rat (CD)	13 wk ad lib (F)		9.3 M		93M (decreased fertility)	Lee et al. 1978, 1985
38	Mouse (CD-1)	4 or 13 wk ad lib (F)		137 M 468 F		413M (mild degeneration of seminiferous tubules)	Hong et al. 1985; Lee et al. 1978
39	Mouse (albino-Swiss)	4 wk (C)		295M		1032M (decreased fertility index)	Lee et al. 1978
40	Dog (Beagle)	4 or 13 wk 1x/d (C)		5 M 25 F		25M (testicular degeneration, decreased spermatogenesis)	Ellis et al. 1985; Lee et al. 1978
Developmental							
41	Rat (CD)	3 or 6 mo ad lib (F)		5.1 F		45.3 F (difficult parturition; decreased pup viability)	Ellis et al. 1979

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference					
					Less serious (mg/kg/day)	Serious (mg/kg/day)						
CHRONIC EXPOSURE												
Death												
42	Rat (CD)	1-2 yr ad lib (F)				3.9 M (decreased survival) 5.1 F (decreased survival)	Lee et al. 1978, 1985; Ellis et al. 1979					
43	Mouse (CD-1)	24 mo ad lib (F)				898 (decreased survival)	Ellis et al. 1979; Hong et al. 1985					
Systemic												
44	Rat (CD)	1-2 yr ad lib (F)	Hemato	0.6 M 5.1 F	3.9M (decreased RBC count)	34.5 M (anemia) 45.3 F (anemia)	Ellis et al. 1979; Lee et al. 1978, 1985					
			Hepatic	5.1 F		0.6 M (preneoplastic foci of 45.3 F altered or hyperplastic hepatocytes)						
		Bd Wt	Renal	34.5 M 45.3 F		34.5 M (30% decrease body weight with decreased food consumption)	Leonard et al. 1987					
				3.9 M 5.1 F		45.3 F (27% decrease body weight with decreased food consumption)						
45	Rat (Fischer- 344)	52 wk ad lib (F)	Hepatic			27M (hepatocellular degeneration and vacuolation; basophilic and acidophilic foci of cellular alteration)	Leonard et al. 1987					
			Bd Wt			27M (25% body weight decrease)						

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
46	Rat (Fischer- 344)	78 wk ad lib (F)	Bd Wt	8 M		20M (25% decrease body weight) (decrease body weight)	NCI 1978
				8.8 F		22 F	
47	Mouse (CD-1)	24 mo ad lib (F)	Hemato	95		898 (anemia; reticulocytosis; Heinz bodies)	Ellis et al. 1979; Hong et al. 1985
			Hepatic	95 F		14M (hepatocellular dysplasia) 898 F (hepatocellular dysplasia)	
			Renal			14M (cystic dysplasia; toxic nephropathy)	
			Bd Wt	14 M	95M (16% decrease in body weight)		
				95 F		898 F (20% decrease in body weight)	
48	Mouse (C57BL/6N)	78 wk ad lib (F)	Bd Wt	14.4 M	72M (18% decrease in body weight gain)		NCI 1978
					15.2 F (11% decrease in body weight gain)	76 F (24% decrease in body weight gain)	
49	Dog (Beagle)	24 mo 1x/d (C)	Hemato	0.2 ^c	1.5 (methemoglobinemia, Heinz bodies)		Ellis et al. 1979, 1985
			Hepatic	0.2	1.5 (biliary hyperplasia)		
			Renal	10			
Neurological							
50	Rat (CD)	1-2 yr ad lib (F)			34.5 M (wide-spread and stiff-legged gait)		Lee et al. 1978, 1985; Ellis et al. 1979
					45.3 F (wide-spread and stiff-legged gait)		

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
51	Mouse (CD-1)	24 mo ad lib		95		898 (stiff-legged gait, hyperactivity)	Ellis et al. 1979; Hong et al. 1985
52	Dog (Beagle)	24 mo 1x/d (C)		0.2		1.5 (loss of hindquarter control, convulsions)	Ellis et al. 1979, 1985
Reproductive							
53	Rat (CD)	1-2 yr ad lib (F)			0.6 M	(atrophy of seminiferous tubules, aspermatogenesis)	Lee et al. 1978, 1985; Ellis et al. 1979
54	Mouse (CD-1)	24 mo ad lib			14M	(decreased spermatogenesis and degenerative change; testicular atrophy)	Ellis et al. 1979; Hong et al. 1985
				95 F	898 F	(ovarian atrophy; nonfunctioning follicles)	
55	Dog (Beagle)	24 mo 1x/d (C)		10M			Ellis et al. 1979, 1985
Cancer							
56	Rat (CD)	1-2 yr ad lib (F)			34.5 M	(CEL: hepatocellular carcinoma, mammary and skin tumors)	Ellis et al. 1979; Lee et al. 1978, 1985
					45.3 F	(CEL: hepatocellular carcinoma; mammary and skin tumors)	

Table 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
57	Rat (Fischer- 344)	78 wk ad lib (F)			7.5-8 M (CEL: skin and subcutaneous fibroma)	22 F (CEL: mammary fibroadenoma)	NCI 1978
58	Mouse (CD-1)	24 mo ad lib			95 M (CEL: renal solid carcinoma, cystic papillary carcinoma and adenoma, cystic adenoma)		Ellis et al. 1979; Hong et al. 1985

^aThe number corresponds to entries in Figure 2-1.

^bUsed to derive an acute-duration oral Minimal Risk Level (MRL) of 0.05 for 2,4-dinitrotoluene by dividing by an uncertainty factor of 100 (10 for animal-to-human extrapolation and 10 for human variability).

^cUsed to derive a chronic-duration oral MRL of 0.002 for 2,4-dinitrotoluene by dividing by an uncertainty factor of 100 (10 for animal-to-human extrapolation and 10 for human variability).

Bd Wt = body weight; (C) = capsule; Cardio = cardiovascular; d = day(s); (F) = feed; F = female; (G) = gavage; Gastro = gastrointestinal; (GO) = gavage, oil; Hemato = hematological; kg = kilogram; LD₅₀ = lethal dose producing 50% death; M = male; mg = milligram; mo = month; Resp = respiratory; wk = week; x = times; yr = year;

Figure 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral

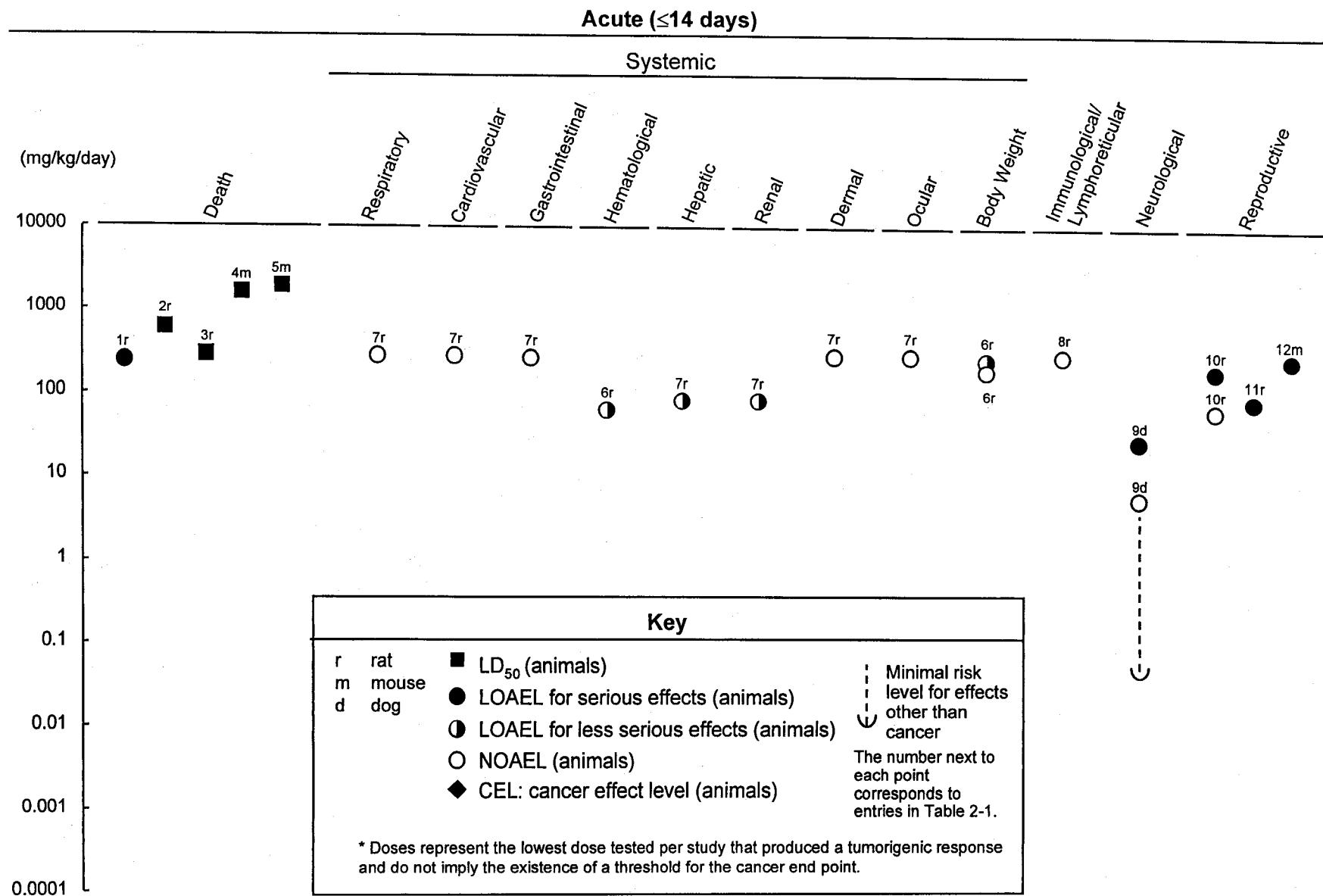


Figure 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)
Intermediate (15-364 days)

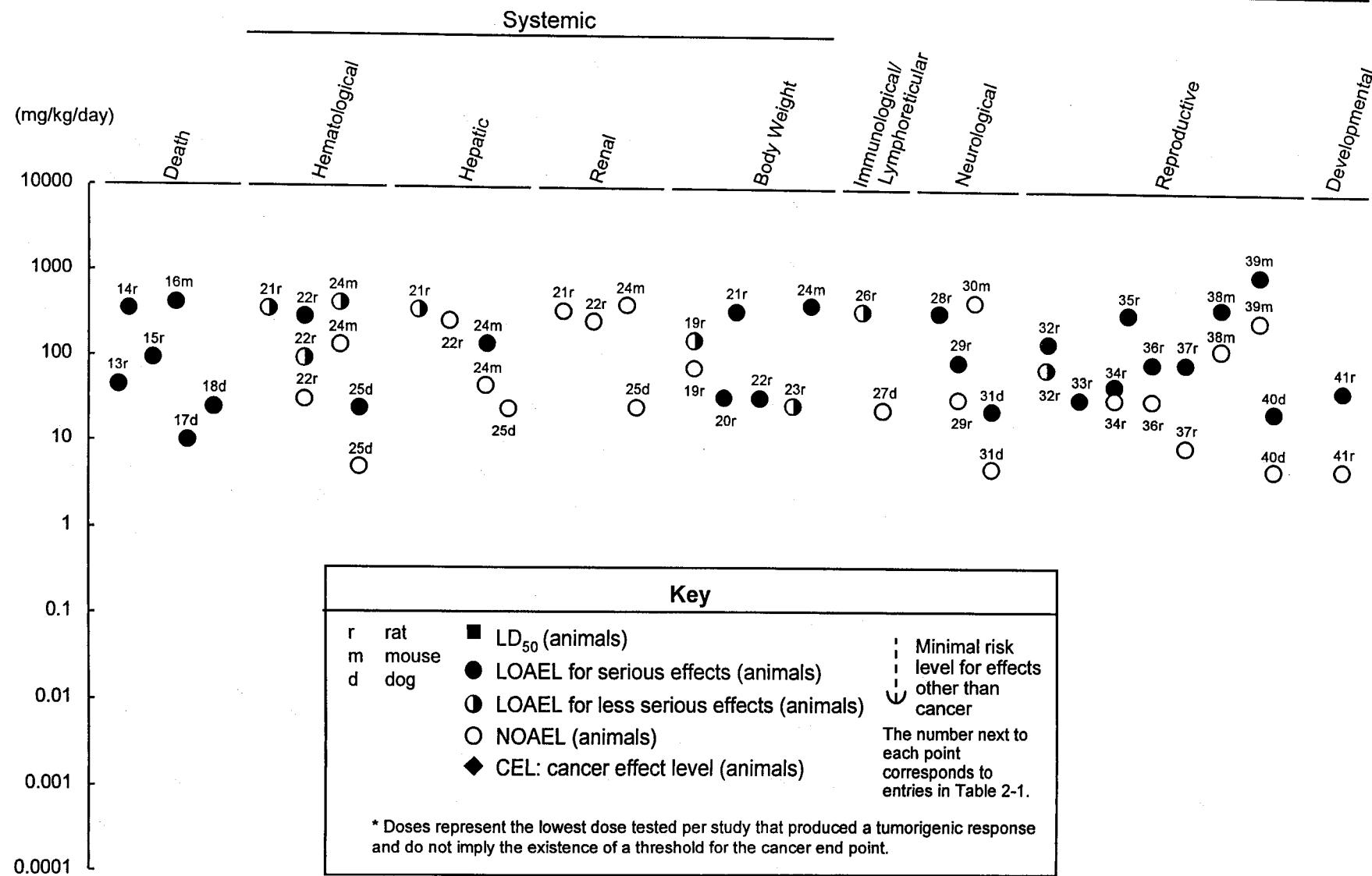


Figure 2-1. Levels of Significant Exposure to 2,4-Dinitrotoluene - Oral (continued)
Chronic (≥ 365 days)

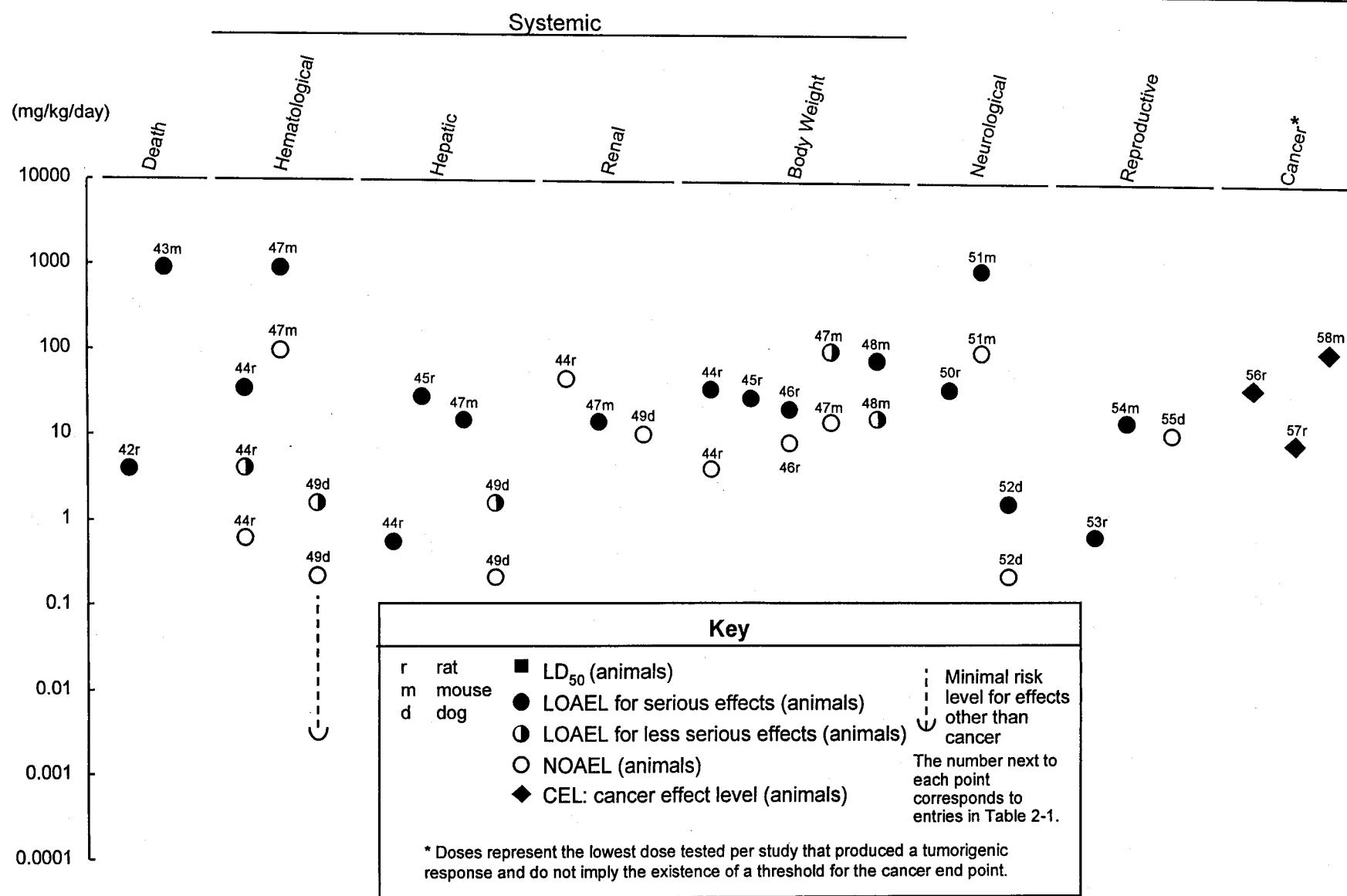


Table 2-2. Levels of Significant Exposure to 2,6-Dinitrotoluene - Oral

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference					
					Less serious (mg/kg/day)	Serious (mg/kg/day)						
ACUTE EXPOSURE												
Death												
1	Rat (CD)	once (GO)			535 M (LD ₅₀) 795 F (LD ₅₀)		Lee et al. 1975; Ellis et al. 1978					
2	Rat (Sprague- Dawley)	once (G)			180 M (LD ₅₀)		Vernot et al. 1977					
3	Mouse (CD)	once (GO)			621 M (LD ₅₀) 807 F (LD ₅₀)		Lee et al. 1975					
4	Mouse (CF-1)	once (G)			1,000 M (LD ₅₀)		Vernot et al. 1977					

Table 2-2. Levels of Significant Exposure to 2,6-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference					
					Less serious (mg/kg/day)	Serious (mg/kg/day)						
INTERMEDIATE EXPOSURE												
Death												
5	Mouse (Swiss- albino)	4 or 13 wk ad lib (F)				51 M (8/16 died) 55 F (6/16 died)	Lee et al. 1976					
6	Dog (Beagle)	4 or 13 wk ad lib (C)				20 F (2/8 died)	Lee et al. 1976					
Systemic												
7	Rat (CD)	4 or 13 wk ad lib (F)	Hemato	7 M 7 F	35 M (splenic hemosiderosis; 37 F extramedullary hematopoiesis)		Lee et al. 1976					
			Hepatic		35 M (bile duct hyperplasia; hemosiderosis) 37 F (bile duct hyperplasia; hemosiderosis)							
			Renal	145 M 155 F								
			Bd Wt	7	35 M (decreased body weight 37 F gain)	145 M (body weight loss) 155 F						
8	Rat (Fischer- 344)	6 or 26 wk ad lib (F)	Bd Wt	7 M		14 M (20% decrease body weight)	Leonard et al. 1987					

Table 2-2. Levels of Significant Exposure to 2,6-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
9	Mouse (Swiss- albino) (F)	4 or 13 wk ad lib	Hemato	11 M 11 F	51 M (extramedullary hematopoiesis) 55 F (extramedullary hematopoiesis)		Lee et al. 1976
			Hepatic	11	51 M (bile duct hyperplasia) 55 F (bile duct hyperplasia)		
			Renal	289 M 299 F			
			Bd Wt	11		51 M (weight loss) 55 F (weight loss)	
10	Dog (Beagle)	4 or 13 wk ad lib (C)	Hemato		4 ^b (mild extramedullary erythropoiesis and lymphoid depletion)		Lee et al. 1976
			Hepatic	4		20 (bile duct hyperplasia; degenerative and inflammatory liver changes)	
			Renal	4	20 (dilated tubules, degenerative foci)		
			Bd Wt	4		20 (body wt. loss with decreased food consumption)	
Immunological/Lymphoreticular							
11	Rat (CD)	4 or 13 wk ad lib (F)		145 M 155 F			Lee et al. 1976
12	Dog (Beagle)	4 or 13 wk ad lib (C)		20	100 (thymic involution)		Lee et al. 1976

Table 2-2. Levels of Significant Exposure to 2,6-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference
					Less serious (mg/kg/day)	Serious (mg/kg/day)	
Neurological							
13	Rat (CD)	4 or 13 wk ad lib (F)		145 M 155 F			Lee et al. 1976
14	Mouse (Swiss- albino)	4 or 13 wk ad lib (F)		289 M 299 F			Lee et al. 1976
15	Dog (Beagle)	4 or 13 wk ad lib (C)		4		20 (incoordination, lack of balance)	Lee et al. 1976
Reproductive							
16	Rat (CD)	4 or 13 wk ad lib (F)		7 M 155 F		35 M (decreased spermatogenesis; degeneration of testes)	Lee et al. 1976
17	Mouse (Swiss- albino)	4 or 13 wk ad lib (F)		11 M 299 F		51 M (decreased spermatogenesis)	Lee et al. 1976
18	Dog (Beagle)	4 or 13 wk ad lib (C)		4 M 100 F		20 M (testicular degeneration)	Lee et al. 1976

Table 2-2. Levels of Significant Exposure to 2,6-Dinitrotoluene - Oral (continued)

Key to ^a figure	Species (Strain)	Exposure/ duration/ frequency (Specific route)	System	NOAEL (mg/kg/day)	LOAEL		Reference					
					Less serious (mg/kg/day)	Serious (mg/kg/day)						
CHRONIC EXPOSURE												
Systemic												
19	Rat	52 wk (Fischer- 344)	Hepatic ad lib (F)			7 M (hepatocellular degeneration, vacuolation; acidophilic and basophilic foci of cellular alteration)	Leonard et al. 1987					
			Bd Wt		7M (18% decrease body weight)							
Cancer												
20	Rat	52 wk (Fischer- 344)	ad lib (F)			7 M (CEL: cholangiocarcinoma, hepatocellular carcinoma)	Leonard et al. 1987					

^aThe numbers correspond to entries in Figure 2-2.

^bUsed to derive an intermediate-duration oral Minimal Risk Level (MRL) of 0.004 for 2,6-dinitrotoluene by dividing by an uncertainty factor of 1000 (10 for use of a LOAEL, 10 for animal-to-human extrapolation, and 10 for human variability).

Bd Wt = body weight; (C) = capsule; F = female; (F) = feed; (G) = gavage; (GO) = gavage, oil; Hemato = hematological; kg = kilogram; LD₅₀ = lethal dose producing 50% death; M = male; mg = milligram; wk = week;

Figure 2-2. Levels of Significant Exposure to 2,6-Dinitrotoluene - Oral
Acute (≤ 14 days)

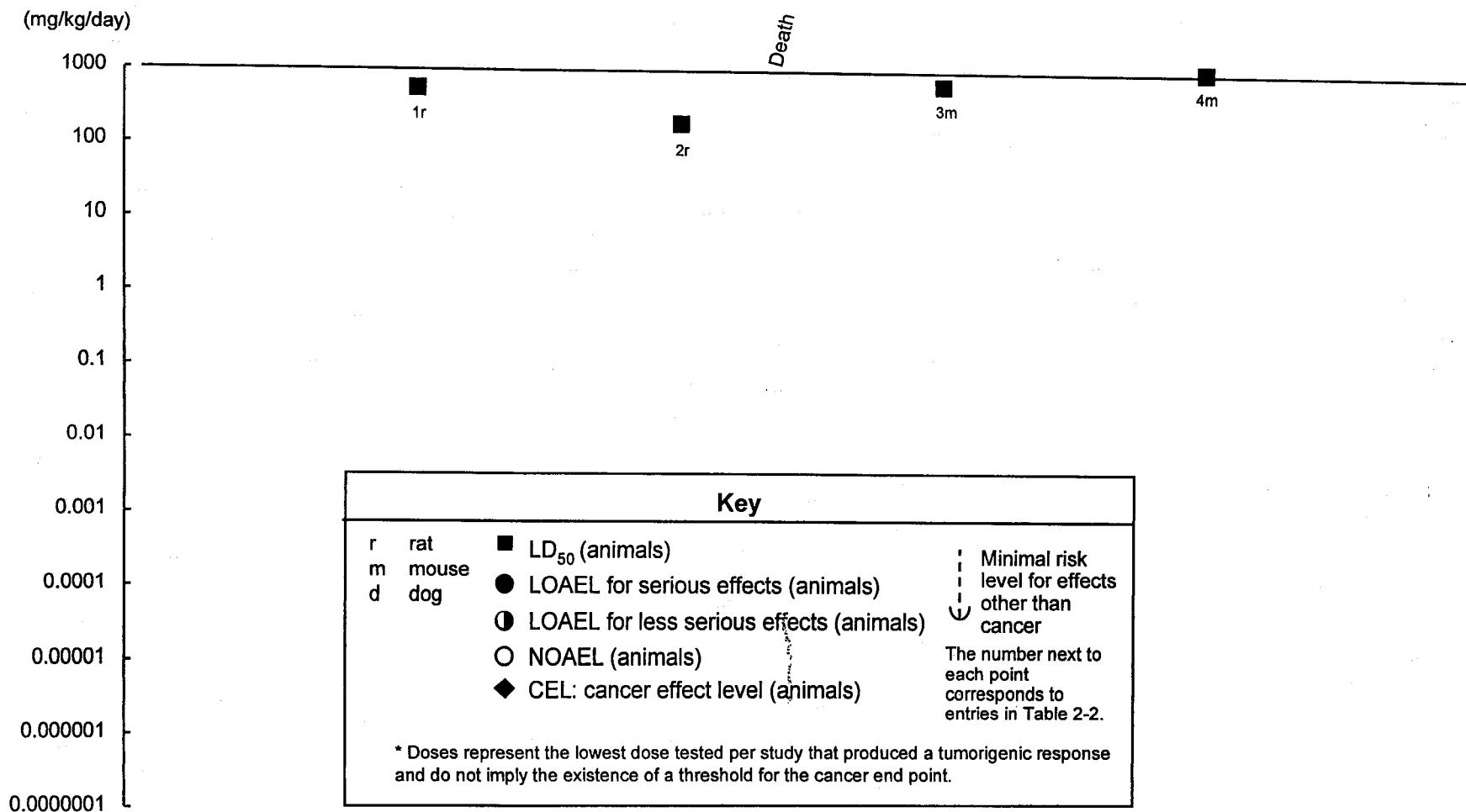


Figure 2-2. Levels of Significant Exposure to 2,6-Dinitrotoluene - Oral (continued)
Intermediate (15-364 days)

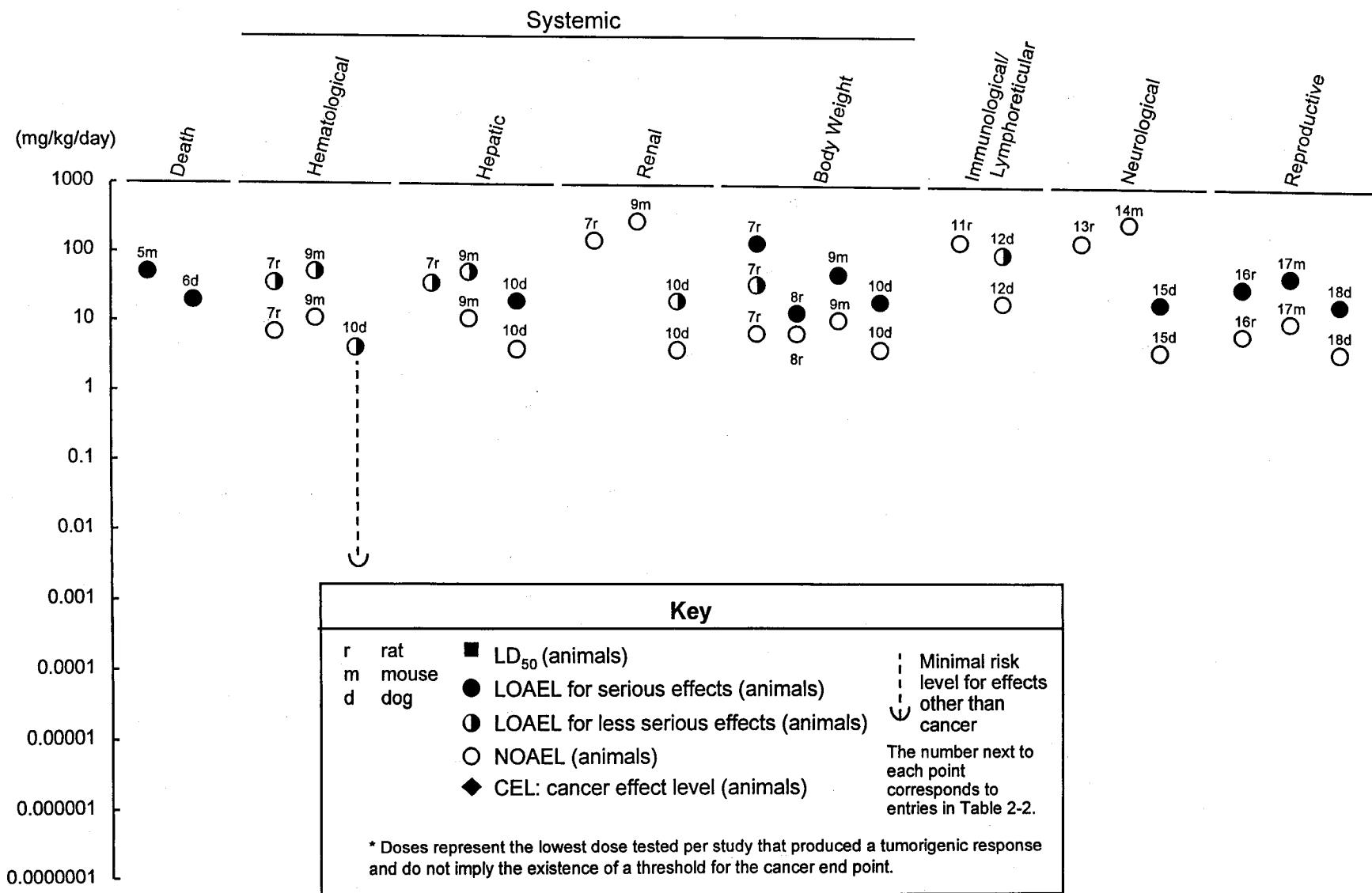
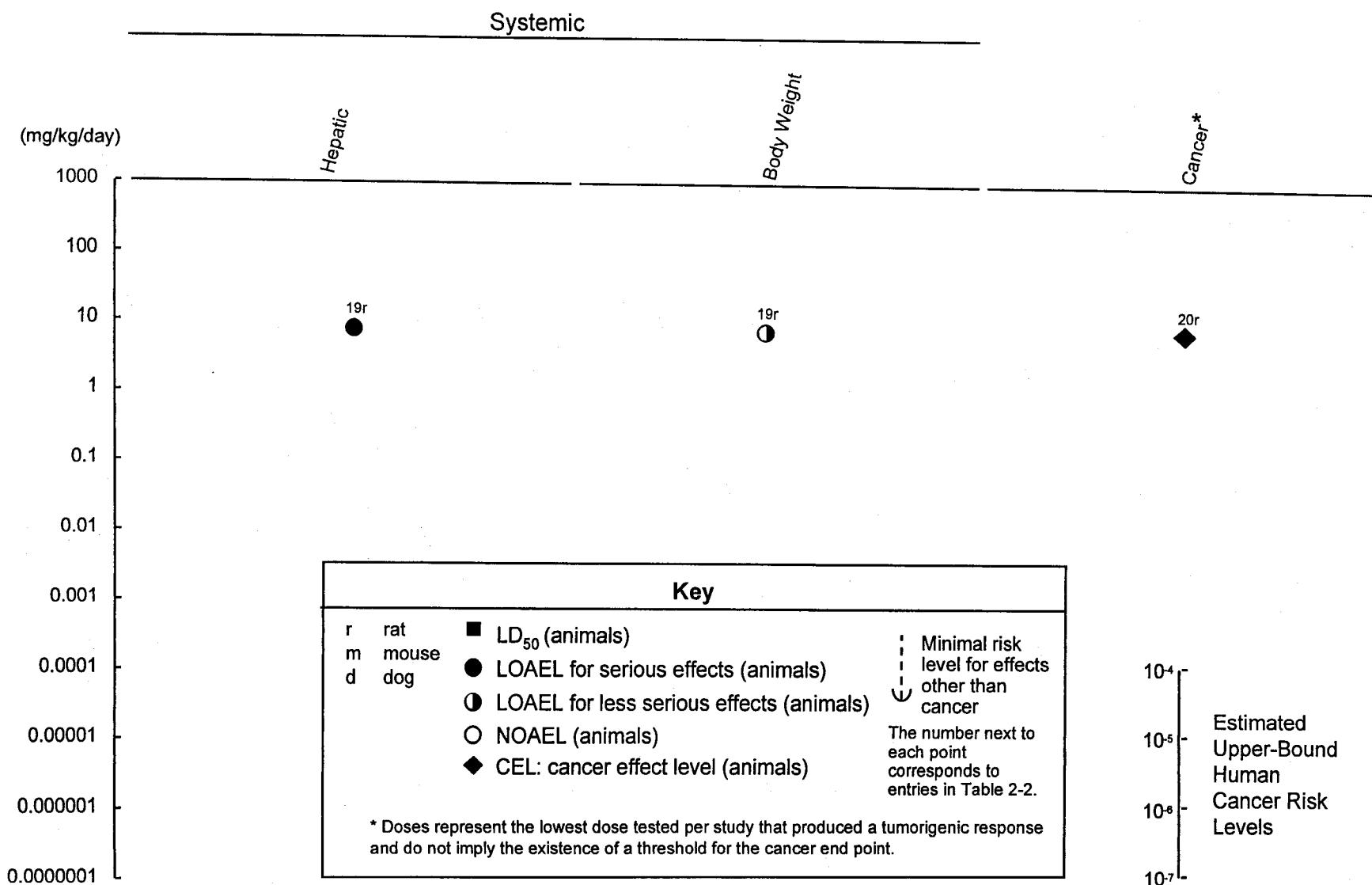


Figure 2-2. Levels of Significant Exposure to 2,6-Dinitrotoluene - Oral (continued)
Chronic (≥ 365 days)



2. HEALTH EFFECTS

Cardiovascular Effects. No studies were located regarding cardiovascular effects in humans after oral exposure to 2,4-DNT or 2,6-DNT.

No histopathological effects on the cardiovascular system were found after Sprague-Dawley rats received 261 mg/kg/day (males) or 273 mg/kg/day (females) 2,4-DNT in the diet for 14 days (McGown et al. 1983).

At the 26-week interim sacrifice in a 104-week study in which CDF rats were fed 0,3.5, 14, or 35 mg/kg/day Tg-DNT in the diet, an increased incidence and severity of myocarditis was noted in males at 35 mg/kg/day (Hazleton Laboratories 1982). It was believed that this spontaneous inflammatory condition was exacerbated by ingestion of Tg-DNT in the high-dose animals. Although this condition was also observed at the 55-week sacrifice, it was not observed at 52 or 104 weeks.

Gastrointestinal Effects. No studies were located regarding gastrointestinal effects in humans after oral exposure to 2,4-DNT or 2,6-DNT.

There were no histopathological effects on the gastrointestinal tract of Sprague-Dawley rats fed 261 mg/kg/day (males) or 273 mg/kg/day (females) for 14 days (McGown et al. 1983).

Treatment of rats with up to 35 mg/kg/day Tg-DNT for up to 1 year or 14 mg/kg/day for up to 2 years did not cause any histopathological changes in the gastrointestinal tract (Hazleton Laboratories 1982).

Hematological Effects. No studies were located regarding hematological effects in humans after oral exposure to 2,4-DNT or 2,6-DNT.

Hematological effects have been noted in virtually all animal studies of oral exposure to 2,4-DNT, 2,6-DNT, and Tg-DNT in which circulating blood has been examined. The most common findings are methemoglobinemia, anemia, reticulocytosis, and an increase in Heinz bodies. The hematological effects are caused by oxidation of the iron in hemoglobin, producing methemoglobin. Heinz bodies are granules in erythrocytes that are believed to result from denatured hemoglobin. Reticulocytosis, a finding in many animals in these studies, is caused by the increased production of immature erythrocytes (red blood cells [RBCs]) and is seen as a compensatory mechanism in anemia resulting from exposure to 2,4- and 2,6-DNT.

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This hematotoxic syndrome is a common effect of exposure to aromatic amines and most organic and inorganic nitrates, and it has been implicated for many oxidizing agents (Ellis et al. 1979; Smith 1996).

Slight cyanosis was observed in rats administered 60 mg/kg 2,4-DNT by gavage for 5 days (Lane et al. 1985). No changes in hematological parameters were found in Sprague-Dawley rats fed 261 mg/kg/day (males) or 273 mg/kg/day (females) 2,4-DNT in the diet for 14 days (McGown et al. 1983). Kozuka et al. (1979) found methemoglobin concentrations increased to 7 times those of controls in the blood of rats fed 347 mg/kg/day 2,4-DNT in the diet for 6 months. Anemia was observed in a 3-week feeding study in which male and female CD rats were fed 145 mg/kg/day in the diet; milder effects, such as reticulocytosis and hemosiderosis or abnormal pigment in the spleen, were found at 93 and 108 mg/kg/day in males and females, respectively (Lee et al. 1978, 1985). No hematological effects were observed in males and females administered 34 and 38 mg/kg/day, respectively. Mild anemia (as indicated by decreases in erythrocyte count, hematocrit, or hemoglobin concentration) and concurrent reticulocytosis were also observed in male and female CD-1 mice administered 413 mg/kg/day and 468 mg/kg/day, 2,4-DNT, respectively in the diet for 13 weeks (Hong et al. 1985; Lee et al. 1978). Anemia, accompanied by the presence of Heinz bodies, was observed in beagle dogs given 25 mg/kg/day 2,4-DNT in capsules (Ellis et al. 1985; Lee et al. 1978).

Subchronic administration of 2,6-DNT to dogs, rats, and mice resulted in hematological effects in dogs and rats at concentrations of 20 and 35 mg/kg/day, respectively (Lee et al. 1976). Dogs were found to be anemic and showed signs of compensatory reticulocytosis at this concentration, and rats showed hemosiderosis and extramedullary hematopoiesis. Statistically significant hematological effects were not observed in mice at levels up to 289 mg/kg/day 2,6-DNT but were observed in individual animals (Lee et al. 1976). Dogs administered 4 mg/kg/day 2,6-DNT by capsule for 13 weeks were found to have extramedullary erythropoiesis and lymphoid depletion of the spleen (Lee et al. 1976). An intermediate-duration oral MRL of 0.004 mg/kg/day for 2,6-DNT was derived from this LOAEL as described in the footnote in Table 2-2. The 2,6-DNT isomer was not tested for hematological endpoints in studies of chronic duration.

Chronic studies of animals administered 2,4-DNT fortify the weight-of-evidence supporting hematological effects. In 24-month studies, hematological effects were observed, but the animals often exhibited "compensated anemia," an adaptive response to 2,4-DNT exposure (Ellis et al. 1979). Methemoglobinemia and the presence of Heinz bodies were observed in dogs administered 1.5 mg/kg/day in capsules; no effect was observed at 0.2 mg/kg/day (Ellis et al. 1985; Lee et al. 1978). The NOAEL of 0.2 mg/kg/day in this dog study was used as the basis for a chronic oral MRL of 0.002 mg/kg/day for 2,4-DNT. In a 2-year study (with

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a-1-year interim sacrifice) in which CD rats were fed 0.6, 3.9, or 34.5 mg/kg/day (males) or 0.7, 5.1, or 45.3 mg/kg/day (females) 2,4-DNT, significant decreases in REK count were found in mid-dose males compared to controls, and anemia was found in high-dose animals after 1 year (Ellis et al. 1979; Lee et al. 1978, 1985). No changes in methemoglobin or Heinz bodies were found, however. CD-1 mice that were administered 14, 95, or 898 mg/kg/day 2,4-DNT in the diet for 24 months were found to be anemic at the high concentration, with compensatory increases in reticulocytes (Ellis et al. 1979; Hong et al. 1985).

Hematological changes consistent with those observed in anemia were found in pregnant Fischer-344 rats administered 100 mg/kg Tg-DNT by gavage during gestation days 7-20 (Jones-Price et al. 1982).

Administration of Tg-DNT to rats in the diet for 4 weeks (Hazleton Laboratories 1977) or 26 weeks (Hazleton Laboratories 1982) resulted in dose- and duration-related adverse effects on hematological parameters. In the 4-week study, at 37.5 mg/kg/day significant increases in reticulocytes and percentage of Heinz bodies were noted in both sexes and significant increases in methemoglobin levels were found in females; anemia was observed at 100 mg/kg/day in both sexes (Hazleton Laboratories 1977). Spleens of rats fed 150 mg/kg Tg-DNT for 30 days in the diet were altered in appearance; these alterations included discoloration, enlargement, and surface irregularity (Hazleton Laboratories 1977). An increased incidence of extramedullary hematopoiesis was noted in the splenic red pulp of male, but not female, rats fed 35 mg/kg/day Tg-DNT in the diet for 52 weeks (Hazleton Laboratories 1982). In rats sacrificed after 26 weeks in a 24-month study, no effects on hematological parameters were observed at 14 mg/kg/day Tg-DNT. However, at 35 mg/kg/day, there were increases in reticulocytes and methemoglobin and decreases in REXs along with hemosiderosis and extramedullary hematopoiesis in males, and increases in mean cell volume (MCV) in females (Hazleton Laboratories 1982). After 1 year, slight-to-moderate myeloid and erythroid hyperplasia was noted in the bone marrow of most male rats treated with 35 mg/kg/day Tg-DNT (Hazleton Laboratories 1982). In a 24-month study in which Tg-DNT was administered to rats in the diet, anemia was observed at 14 mg/kg/day in males but not in females; the NOAEL for this effect in males was 3.5 mg/kg/day Tg-DNT (Hazleton Laboratories 1982).

The consistent observation of adverse hematological effects following exposure of laboratory animals to DNT indicates that the blood is a primary target of DNT toxicity.

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Musculoskeletal Effects. No studies were located regarding musculoskeletal effects in humans or animals after oral exposure to 2,4-DNT, 2,6-DNT, or Tg-DNT.

Hepatic Effects. No studies were located regarding hepatic effects in humans after oral exposure to 2,4- or 2,6-DNT.

The hepatotoxic effects of DNT have been consistently observed in animals. The liver appears to be a target organ of DNT toxicity, particularly when administered to rats, but hepatotoxic effects have also been observed in mice and dogs. Hepatic effects of DNT include liver discoloration and inflammation, alteration of hepatocytes, proliferation of bile duct epithelium, and hyperplastic foci.

Increased blood cholesterol was found in male and female Sprague-Dawley rats fed 78 or 82 mg/kg/day 2,4-DNT, respectively, in the diet for 14 days, and increased alanine aminotransferase levels were found in males (McGown et al. 1983). Blood glucose levels trended upward in all male and female groups in this study, but were increased significantly only in females fed 273 mg/kg/day.

Oral administration of 2,4-DNT for 13 weeks to rats (266 or 145 mg/kg/day in males and females, respectively) and dogs (25 mg/kg/day) did not result in liver toxicity (Ellis et al. 1985; Lee et al. 1978). After 26 weeks of treatment, rats fed 27 mg/kg/day in the diet had significant increases in epoxide hydrolase (EH) activity, which is sometimes considered to be a phenotypic marker of neoplastic nodules; however, hepatocellular lesions did not develop in these animals when treatment was carried through 52 weeks (Leonard et al. 1987). Mild hepatocellular dysplasia was observed in mice fed 137 mg/kg/day (males) or 468 mg/kg/day (females) of 2,4-DNT for 13 weeks (Hong et al. 1985; Lee et al. 1978).

The most severe hepatotoxicity was found in rats. Concentrations of 2,4-DNT as low as 0.6 mg/kg/day caused foci of altered hepatocytes or hyperplastic nodules when fed to male rats for 1 year (Ellis et al. 1979; Lee et al. 1978, 1985). Hepatocellular degeneration and vacuolation accompanied by acidophilic foci and occasional basophilic foci of cellular alteration were found in Fischer-344 rats fed 27 mg/kg/day 2,4-DNT for 52 weeks (Leonard et al. 1987). The incidences of focal areas of alteration were less in the 2,4-DNT.-treated rats than they were in rats similarly treated with 2,6-DNT or Tg-DNT. Wistar rats fed dietary concentrations of 347-395 mg/kg/day 2,4-DNT for 6 months had increased relative liver weights, formation of puruloid matter, and increased levels of serum glutamic-oxaloacetic transaminase (SGOT), lactate dehydrogenase (LDH), alkaline and acid phosphatase, triglycerides, and blood glucose levels compared to controls (Kozuka

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et al. 1979). In this study, the levels of serum albumin and the albumin/globulin ratios were decreased. Hepatocellular dysplasia was found in male and female CD-1 mice fed 14 or 898 mg/kg/day 2,4-DNT, respectively, for 24 months (Ellis et al. 1979; Hong et al. 1985). Administration of 1.5 mg/kg/day 2,4-DNT for 24 months resulted in biliary hyperplasia in dogs; this effect was not seen in dogs administered 0.2 mg/kg/day (Ellis et al. 1979, 1985).

Six weeks of dietary consumption of 7 mg/kg/day 2,6-DNT caused a 380% increase in epoxide hydrolase (EH) levels in rats but did not increase the level of DT-diaphorase (DTD) (Leonard et al. 1987). In the same study, both of these enzymes were elevated after 6 weeks of treatment with 14 mg/kg/day of 2,6-DNT. Dosing of rats and dogs with 2,6-DNT for 13 weeks resulted in liver toxicity (Lee et al. 1976). Bile duct hyperplasia was observed in rats fed 35 mg/kg/day and mice fed 51 mg/kg/day 2,6-DNT for 13 weeks (Lee et al. 1976). Liver degeneration and bile duct hyperplasia were observed in dogs dosed with 20 mg/kg/day 2,6-DNT but were not seen in dogs dosed with 4 mg/kg/day (Lee et al. 1976). After 52 weeks of treatment with 7 mg/kg/day 2,6-DNT, hepatocellular degeneration and vacuolation accompanied by acidophilic and basophilic foci of cellular alteration were found in Fischer-344 rats (Leonard et al. 1987).

Irregular liver surfaces were found in male Fischer-344 rats fed 37.5 mg/kg/day Tg-DNT for 30 days (Hazleton Laboratories 1977). Hepatocytic necrosis, nonsupportive pericholangitis, and periportal megalocytosis were found in CDF rats fed 14 mg/kg/day for 26 weeks, and when treatment of these animals was extended to 2 years, slight-to-severe biliary cirrhosis was found in males (Hazleton Laboratories 1982). It has been suggested that this latter lesion may be a precursor to cholangiocarcinoma (Hazleton Laboratories 1982). Hepatocytic degeneration, and acidophilic and basophilic foci of cellular alteration were observed in Fischer-344 rats fed 35 mg/kg/day Tg-DNT in the diet for 52 weeks (Leonard et al. 1987). When administration of Tg-DNT was continued for 24 months, liver discoloration resulted at 3.5 mg/kg/day and liver nodules and malignancies at 14 mg/kg/day (Hazleton Laboratories 1982).

Renal Effects. No studies were located regarding renal effects in humans after oral exposure to 2,4- or 2,6-DNT.

Hyaline droplet accumulation in the epithelium of the proximal convoluted tubule was found in both sexes of Sprague-Dawley rats after they were administered 78, 104, 165, or 261 mg/kg/day 2,4-DNT (males) or 82, 109, 173, or 273 mg/kg/day 2,4-DNT (females) in the diet (McGown et al. 1983). Although this effect was observed at all concentrations, there was no dose response evident. Oral administration of 2,4-DNT to mice

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(413 mg/kg/day), rats (145 mg/kg/day), and dogs (25 mg/kg/day) for 13 weeks did not result in significant adverse effects in the kidney (Hong et al. 1985; Lee et al. 1978). Treatment of the same species for 24 months resulted in renal dysplasia in male mice at a dose of 14 mg/kg/day of 2,4-DNT, but no renal effects were observed in rats or dogs dosed with 34.5 mg/kg/day or 10 mg/kg/day, respectively (Ellis et al. 1979). Adverse effects in the kidneys of mice included cystic dysplasia in the tubular epithelium, atypical epithelium lining the cysts, and a variety of tumors (Hong et al. 1985). These effects were more pronounced in male mice than in female mice.

Dosing of dogs with 20 mg/kg/day 2,6-DNT for 13 weeks resulted in dilated tubules, foci of inflammation, and degeneration of the kidney (Lee et al. 1976). No treatment-related effects on the kidney were found when rats were fed 2,6-DNT for 13 weeks (Lee et al. 1976). The severe renal effects observed after 2,4-DNT administration in mice were not observed when mice were fed 289 mg/kg/day 2,6-DNT for 13 weeks (Lee et al. 1976). However, the renal toxicity of 2,4-DNT in mice was observed only after chronic administration. Chronic studies of 2,6-DNT have not been performed in mice.

After 26 or 52 weeks of dietary consumption of 35 mg/kg/day Tg-DNT, blood urea nitrogen (BUN) levels were significantly increased in CDF rats (Hazleton Laboratories 1982). Exacerbation of chronic interstitial nephritis that was also observed in controls was observed at 14 mg/kg/day Tg-DNT in a chronic study in rats (Hazleton Laboratories 1982).

The kidney does not appear to be a sensitive target of DNT toxicity for all species tested. Severe renal effects were observed only in CD-1 mice fed 2,4-DNT for 24 months, and less severe renal effects were observed in dogs administered 20 mg/kg/day 2,6-DNT for 13 weeks.

Endocrine Effects. No studies were located regarding endocrine effects in humans after oral exposure to 2,4- or 2,6-DNT.

Administration of 2,4-DNT in the diet for 14 days, at 78 mg/kg/day for males or 82 mg/kg/day for females did not cause any histopathological changes in adrenal, pituitary, or thyroid glands of Sprague-Dawley rats (McGown et al. 1983).

No histopathological effects on adrenal, pituitary, or thyroid glands were found in rats treated with 14 mg/kg/day Tg-DNT for up to 2 years or 35 mg/kg/day for 1 year (Hazleton Laboratories 1982). Increases

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in the incidence and severity of parathyroid hyperplasia (males) and increases in the incidence and severity of fatty metamorphosis and vascular ectasia (males and females) were found in rats fed 14 mg/kg/day Tg-DNT in the diet in a chronic study (Hazleton Laboratories 1982).

Dermal Effects. No studies were located regarding dermal effects in humans after oral exposure to 2,4- or 2,6-DNT.

Concentrations of up to 261 mg/kg/day 2,4-DNT for males or 273 mg/kg/day 2,4-DNT for females administered in the diet for 14 days to Sprague-Dawley rats caused no histopathological changes in their skin (McGown et al. 1983).

No effects were found on the skin of rats treated for up to 2 years with 14 mg/kg/day Tg-DNT or up to 1 year with 35 mg/kg/day Tg-DNT (Hazleton Laboratories 1982).

Ocular Effects. No studies were located regarding ocular effects in humans after oral exposure to 2,4- or 2,6-DNT.

The eyes of male and female Sprague-Dawley rats administered up to 261 mg/kg/day or 273 mg/kg/day 2,4-DNT, respectively, in the diet for 14 days did not exhibit any alterations upon histopathological examination (McGown et al. 1983).

No effects were found on the eyes of rats treated for up to 2 years with 14 mg/kg/day Tg-DNT in feed or up to 1 year with 35 mg/kg/day Tg-DNT in feed (Hazleton Laboratories 1982).

Body Weight Effects. No studies were located regarding body weight effects in humans after oral exposure to 2,4- or 2,6-DNT.

Adverse effects on body weight and body weight gain in rats, mice, and dogs were observed after oral administration of 2,4-DNT, 2,6-DNT, and Tg-DNT. In most of these studies, a concurrent decrease in food consumption was also observed. Because exposure resulted from intake of the test article in feed in most of these studies, it is possible that some of the body weight changes resulted from inpalatability.

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Adverse effects on body weight, including body weight loss, have been reported after almost all acute-, intermediate-, and chronic-duration oral administration of 2,4-DNT (Bloch et al. 1988; Ellis et al. 1979, 1985; Hazleton Laboratories 1982; Hong et al. 1985; Kozuka et al. 1979; Lane et al. 1985; Lee et al. 1978, 1985; Leonard et al. 1987; McGown et al. 1983; NCI 1978). In an acute study, rats dosed by gavage with 240 mg/kg 2,4-DNT for 5 days lost weight (Lane et al. 1985). In general, there were losses of 10-40% in body weight in acute-, intermediate-, and chronic-duration studies in rats. After 6 months, decreases in body weight gain were noted in rats fed 27 mg/kg/day 2,4-DNT (Leonard et al. 1987), and a 25% decrease in body weight was seen in rats fed 34.5 mg/kg/day (Ellis et al. 1979; Lee et al. 1978, 1985). This reduction in body weight gain tended to become more pronounced when 2,4-DNT was continued for periods of 1-2 years (Leonard et al. 1987). Body weight was decreased 25% in rats that received 20 mg/kg/day 2,4-DNT in the diet for 78 weeks; the NOAEL in this study was 8 mg/kg/day (NCI 1978). Mice showed similar decreases in body weight after intermediate- and chronic-duration exposure, but the concentrations of the test article needed to evoke this effect were considerably higher than in rats (Hong et al. 1985; Lee et al. 1978; NCI 1978). An 18-24% decrease in body weight was seen in rats receiving 72-76 mg/kg/day 2,4-DNT in the diet for 78 weeks (NCI 1978).

Administration of 2,6-DNT also caused decreased body weight gain or body weight loss in rats, mice, and dogs at concentrations ranging from 14 to 145 mg/kg/day in intermediate-duration studies (Lee et al. 1976). Treatment with 7 mg/kg/day 2,6-DNT decreased body weight in rats at 52 weeks by 18% (Leonard et al. 1987).

A 29% decrease in absolute maternal weight gain was observed in dams fed 14 mg/kg/day Tg-DNT for 14 days during gestation (Jones-Price et al. 1982). Decreased body weight or decreased body weight gain was reported in rats at levels as low as 14 mg/kg/day Tg-DNT in intermediate- or chronic-duration studies (Hazleton Laboratories 1982). Other intermediate- and chronic-duration studies also confirmed these body weight effects (Hazleton Laboratories 1977; Leonard et al. 1987; NCI 1978).

2.2.2.3 Immunological and Lymphoreticular Effects

No studies were located regarding immunological and lymphoreticular effects in humans after oral exposure to 2,4- or 2,6-DNT.

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Testing for immunological effects of DNT is limited. No changes in serum concentrations of IgE were observed in rats and dogs administered 2,4-DNT at levels up to 206 and 25 mg/kg/day, respectively, for 13 weeks (Ellis et al. 1985; Lee et al. 1978, 1985). In these studies, the rats received the test article in feed, while the dogs received it in capsules. No histopathological changes were found in the spleen or thymus of Sprague-Dawley male rats fed 78 mg/kg/day 2,4-DNT or female rats fed 82 mg/kg/day 2,4-DNT in the diet for 14 days (McGown et al. 1983).

Administration of 2,6-DNT to dogs (up to 100 mg/kg) and rats (up to 145 mg/kg/day) for 13 weeks resulted in no observable changes in IgE serum concentrations (Lee et al. 1976). IgE is the antibody associated with allergic or hypersensitive reactions, and so it may be expected that the human sensitizing potential of 2,4-DNT and 2,6-DNT would be low. Involution of the thymus was noted when dogs were administered 100 mg/kg 2,6-DNT, but was not noted when they were administered 20 mg/kg 2,6-DNT by capsule for 13 weeks (Lee et al. 1976).

For 2,4-DNT, the highest NOAEL values and all LOAEL values from each reliable study for immunological/lymphoreticular effects in each species and duration category are recorded in Table 2-1 and plotted in Figure 2- 1. For 2,6-DNT, the highest NOAEL values and all LOAEL values from each reliable study for immunological/lymphoreticular effects in each species and duration category are recorded in Table 2-2 and plotted in Figure 2-2.

2.2.2.4 Neurological Effects

No studies were located regarding neurological effects in humans after oral exposure to 2,4- or 2,6-DNT. Severe clinical signs of neurotoxicity, including incoordination and stiffness that led to an abnormal gait were observed in dogs given 25 mg/kg/day 2,4-DNT in capsules for 12 days (Ellis et al. 1985; Lee et al. 1978). These effects progressed with time and this dose was lethal after 22 days. The NOAEL for the neurotoxicity observed after 12 days was 5 mg/kg/day. An acute-duration oral MRL of 0.05 mg/kg/day was derived based on this NOAEL as described in the footnote in Table 2-1. No histopathological changes were found in the brain or spinal cord of male and female Sprague-Dawley rats fed 2,4-DNT for 14 days in the diet at doses of 78 and 82 mg/kg/day, respectively (McGown et al. 1983). Neurotoxicity has been reported in laboratory animals after intermediate- or chronic-duration exposure to 2,4-DNT with symptoms ranging from tremors, convulsions, and ataxia to paralysis. These effects were observed in 13-week studies of rats and dogs.

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Administration of 93 mg/kg/day 2,4-DNT in the diet for 13 weeks caused demyelination in the cerebellum and brain stem of 1 male rat, while at 266 mg/kg/day, some rats exhibited a widespread or stiff-legged gait that did not progress to the rigid paralysis observed in dogs (Lee et al. 1978, 1985). After 3 months of being fed 2,4-DNT in the diet, Wistar rats exhibited humpback and jerky incoordination (Kozuka et al. 1979). Dogs that were administered 25 mg/kg/day 2,4-DNT in capsules for 13 weeks began to show neurotoxic effects within 2 months; these effects included incoordination, abnormal gait, rigid paralysis of the hind legs, eventually progressing to paralysis up to the neck (Ellis et al. 1985; Lee et al. 1978). No neurological signs were observed in mice fed 413 mg/kg/day in males or 468 mg/kg/day in females 2,4-DNT in the diet for 13 weeks (Hong et al. 1985; Lee et al. 1978).

An abnormal gait was also observed in chronic studies of laboratory animals fed 2,4-DNT. The characteristic widespread and stiff-legged gait was observed after feeding 34.5 mg/kg/day or 45.3 mg/kg/day 2,4-DNT to male and female rats, respectively, for up to 2 years (Ellis et al. 1979; Lee et al. 1978, 1985). This stifflegged gait and hyperactive behavior were also noted in mice fed 898 mg/kg/day in the diet for 24 months but were not observed in mice at 95 mg/kg/day (Ellis et al. 1979, 1985). Dogs dosed at 1.5 mg/kg/day 2,4-DNT in a 2-year study showed loss of hindquarter control (Ellis et al. 1979, 1985). Central nervous system lesions were identified in high-dose (10 mg/kg/day) dogs in this study and included vacuolization, hypertrophy, endothelial mitosis, and focal gliosis in the cerebellum, as well as some perivascular hemorrhage in the cerebellum and brain stem (Ellis et al. 1979, 1985).

Dogs dosed with 20 or 100 mg/kg/day of 2,6-DNT for 13 weeks exhibited dose-dependent neurotoxic symptoms that included muscular incoordination, weakness, tremors, and paralysis (Lee et al. 1976). Rats and mice dosed at 145 and 289 mg/kg/day of 2,6-DNT, respectively, for 13 weeks did not display neurotoxic symptoms (Lee et al. 1976).

Administration of 150 mg/kg/day Tg-DNT to Fischer-344 dams during gestation days 7-20 caused hindlimb weakness in 7 of 13 animals (Jones-Price et al. 1982). No clinical signs of neurotoxicity or histopathological changes were found in rats fed up to 35 mg/kg/day Tg-DNT in the diet for 26 or 52 weeks (Hazleton Laboratories 1982).

Neurotoxicity appears to be a characteristic syndrome of DNT poisoning of animals. Neurotoxic symptoms, of decreased severity compared to dogs, were observed in mice and rats at doses higher than neurotoxic doses in dogs; however, the test article was administered in feed to rodents and in capsules to dogs.

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For 2,4-DNT, the highest NOAEL values and all LOAEL values from each reliable study for neurological effects in each species and duration category are recorded in Table 2-1 and plotted in Figure 2-1. For 2,6-DNT, the highest NOAEL values and from each reliable study for neurological effects in each species and duration category are recorded in Table 2-2 and plotted in Figure 2-2.

2.2.2.5 Reproductive Effects

No studies were located regarding reproductive effects in humans after oral exposure to 2,4- or 2,6-DNT. Studies in laboratory animals have shown that oral exposure to 2,4-DNT can result in adverse effects on reproduction. This is evidenced by decreased fertility as well as by lesions of the male and female reproductive tracts. The male reproductive system seems to be particularly sensitive; observed effects include decreased sperm production, testicular atrophy, changes in Sertoli cell morphology, and degenerated seminiferous tubules (Bloch et al. 1988; Ellis et al. 1979; Kozuka et al. 1979; Lane et al. 1985; Lee et al. 1976, 1978; McGown et al. 1983). In the female reproductive system, ovarian atrophy and dysfunction were observed (Ellis et al. 1979).

The effects on the male reproductive system have been reported in studies of brief durations. Decreased fertility was noted in male rats dosed with 180 mg/kg 2,4-DNT for 5 days; no dominant lethal effect was observed at this dose (Lane et al. 1985). Sprague-Dawley rats administered 104, 165, or 261 mg/kg/day 2,4-DNT in the diet for 14 days exhibited oligospermia with degenerative changes, such as syncytial cell formation and focal spermatocytic granuloma, in a dose-dependent manner (McGown et al. 1983). A concentration of 78 mg/kg/day 2,4-DNT caused a decrease in the thickness of spermatogenic cell layers. No histopathological changes were found in the reproductive organs of females in this study (McGown et al. 1983). Although no changes were found in sperm morphology of male mice that were administered 250 mg/kg/day 2,4-DNT for 2 days, significant decreases in fertile matings of these animals were observed during weeks 2, 3, and 6 post-treatment (Soares and Lock 1980). However, sperm morphology was examined at 8 weeks post-treatment, so it is possible that a toxic effect was selective for specific types of sperm cells.

In intermediate studies of 2,4-DNT, serious effects on the male reproductive system have been observed in numerous animal studies. In a series of 3 dominant lethal studies using male rats for 13 weeks, 45 mg/kg/day 2,4-DNT in the diet caused severe atrophy and degeneration of the seminiferous tubules, resulting in

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decreased fertility, although no dominant lethal effect was observed (Ellis et al. 1979). Another study using CD rats found that spermatogenesis was impaired after 4 weeks of feeding 93 mg/kg/day 2,4-DNT in the diet and had completely ceased after 13 weeks (Lee et al. 1978, 1985). This effect was not reversible after a 4-week post-treatment period. Higher concentrations of 2,4-DNT were needed to cause these effects in mice. Testicular atrophy and aspermatogenesis occurred in CD-1 mice fed 4 13 mg/kg/day 2,4-DNT for 13 weeks (Hong et al. 1985; Lee et al. 1978) and rats fed 347-395 mg/kg/day 2,4-DNT for 6 months in feed (Kozuka et al. 1979). Decreased fertility was observed after male mice were treated with 1,032 mg/kg/day, but not 295 mg/kg/day 2,4-DNT in the feed for 4 weeks in a dominant lethal study (Lee et al. 1978). The decreased fertility was not observed in mice fed 295 mg/kg/day (Lee et al. 1978). The testicular atrophy was considered to be due to a direct toxic effect on spermatogenic cells. Mild-to-severe testicular degeneration with decreased spermatogenesis has also been observed in dogs administered 25 mg/kg 2,4-DNT in capsules for 13 weeks (Ellis et al. 1985; Lee et al. 1978). No testicular effects were found at 5 mg/kg in the study.

Chronic-duration studies in laboratory animals have also demonstrated both male and female reproductive effects. Rats that received 0.6 mg/kg/day 2,4-DNT in the diet for up to 2 years had an increased incidence of seminiferous tubule atrophy (29%) compared to controls (16%) (Ellis et al. 1979; Lee et al. 1978, 1985). This was the lowest concentration used in this study in which dose-related changes were found. At the highest dose, 35 mg/kg/day, an 8 1% increase in seminiferous tubule atrophy and spermatogenesis was reported. Similar changes were found in male CD-1 mice fed 14 mg/kg/day 2,4-DNT for 24 months (Hong et al. 1985). Female mice fed 898 mg/kg/day 2,4-DNT in this study had ovarian atrophy with non-functioning follicles and, therefore, a lack of corpora lutea (Hong et al. 1985). The NOAEL for these effects was 95 mg/kg/day. No adverse reproductive effects were found in dogs fed 10 mg/kg/day 2,4-DNT for 24 months (Ellis et al. 1979,1985).

Histopathological examination of the testes after treatment with 2,4-DNT has revealed changes which suggest specific causes for the male infertility observed in animal studies. Dose-dependent changes in sperm cell morphology were found in Sprague-Dawley rats fed 76.7 or 153.4 mg/kg/day 2,4-DNT in the diet for 3 weeks (Bloch et al. 1988). At the low dose, vacuolation and lipid accumulation were noted in Sertoli cells; multinucleated spermatid and irregularities of the basal lamina were also found. These changes were limited and variable with most samples, demonstrating patchy damage. More extensive degenerative changes in both spermatocytes and spermatids were found at the high dose as well as ultrastructural changes in Sertoli cells; epididymal sperm counts were decreased 63%. The high-dose animals also had increased levels of serum luteinizing hormone (LH) and FSH but not testosterone (Bloch et al. 1988).

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A three-generation reproductive toxicity study was performed in rats fed 2,4-DNT for up to 6 months before mating of original prenatal animals (Ellis et al. 1979). Effects on neonatal viability were observed at the highest concentration of 2,4-DNT used, 40 mg/kg/day. Reductions in neonatal viability became more severe with successive litters within each generation, such that no second litters were produced by the second generation of high-dose animals, which were fed 34.5 mg/kg/day (male) or 45.3 mg/kg/day (female) 2,4-DNT. Decreases in the number of fetal implants were attributed to the adverse impact of 2,4-DNT on sperm production.

In studies of rats, mice, and dogs dosed with 2,6-DNT for 13 weeks (Lee et al. 1976), decreased spermatogenesis was observed in male mice administered 51 mg/kg/day, but normal spermatogenesis was observed in animals dosed with 11 mg/kg/day. Testicular atrophy was reported in rats administered 35 mg/kg/day 2,6-DNT, and no effects were observed in rats dosed with 7 mg/kg/day (Lee et al. 1976). Dogs dosed with 20 and 100 mg/kg/day had testicular degeneration, but no effects were observed in dogs dosed with 4 mg/kg/day (Lee et al. 1976).

Based upon the testicular effects observed after administration of 2,4-DNT or 2,6-DNT, it is not surprising that these effects are found after treatment with Tg-DNT. Testicular degeneration was found in male rats fed 35 mg/kg/day Tg-DNT for 26 weeks, but since the finding was unilateral, the relationship to treatment may be considered equivocal (Hazleton Laboratories 1982). When treatment with this concentration was carried through 52 weeks, however, bilateral mild-to-severe testicular degeneration and hypospermatogenesis were observed (Hazleton Laboratories 1982). No changes were found in the fertility or sperm morphology of male mice that received 250 mg/kg Tg-DNT by gavage for 2 days in a dominant lethal study (Soares and Lock 1980).

The highest NOAEL values and all reliable LOAEL values for reproductive effects in each species and duration category are recorded in Tables 2-1 and 2-2 and plotted in Figures 2-1 and 2-2.

2.2.2.6 Developmental Effects

No studies were located regarding developmental effects in humans after oral exposure to 2,4- or 2,6-DNT. However, developmental toxicity from DNT could potentially occur because exposure to any substance that depletes the amount of oxygen available to developing fetal tissues can have adverse consequences. The hematological (and potential oxygen depleting) effects of DNT are reported in Section 2.2.1.2.

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Ellis et al. (1979) conducted a 3-generation reproductive study in which 2,4-DNT was administered to male and female rats at doses up to 34.5 and 45.3 mg/kg/day, respectively. Normal birth weights, liveborn index, and weight at weaning were observed. Decreases in pup viability at 45.3 mg/kg/day in this study resulted from maternal neglect and a high incidence of maternal death during parturition; these decreases did not appear to result from pup defects since no anomalies were detected in offspring from any generation. These effects were not observed in animals fed 5.1 mg/kg/day 2,4-DNT (Ellis et al. 1979).

Tg-DNT was administered by gavage to pregnant rats for 14 days during gestation, and pups were evaluated for developmental toxicity either at gestation day 20 or postpartum day 60 (Jones-Price et al. 1982). Adverse effects on hematologic parameters and altered organ weights were observed in both dams and fetuses when dams were administered 100 or 150 mg/kg/day. However, the fetal toxicity was not dose related. A decrease in relative liver weight was observed, however, in the postpartum pups at the low dose of 14 mg/kg/day; this dose is considered to be a LOAEL. Dose-related effects on postnatal development were not observed in pups when dams were administered 35 or 75 mg/kg/day. Transient and statistically significant signs of neurotoxicity, which were not dose-related, included delayed eye opening and cliff avoidance when dams were treated with 35 or 75 mg/kg/day. No evidence of toxicity was found in pups at postpartum day 60 of the postnatal study.

The highest NOAEL values and all reliable LOAEL values for developmental effects in each species and duration category are recorded in Tables 2-1 and 2-3 and plotted in Figures 2-1 and 2-3.

2.2.2.7 Genotoxic Effects

Studies of the effects of various DNT isomers on sperm morphology (Soares and Lock 1980), spermatocyte DNA repair (Working and Butterworth 1984), and dominant lethal mutations (Ellis et al. 1979; Hodgson et al. 1976; Soares and Locke 1980) were generally negative for these specific endpoints.

Other genotoxicity studies are discussed in Section 2.5.

2.2.2.8 Cancer

No studies were located regarding cancer in humans after oral exposure to 2,4- or 2,6-DNT.

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The carcinogenic activity of DNT has been extensively studied in typical chronic bioassays and in some less than-lifetime studies. 2,4-DNT produced renal tumors in male mice and was hepatocarcinogenic in rats. 2,6-DNT and Tg-DNT are potent hepatocarcinogens in rats (Ellis et al. 1979; Lee et al. 1978, 1985).

2,4-DNT (98% 2,4-DNT, 2% 2,6-DNT) produced renal tumors (76%) in male CD-1 mice fed 95 mg/kg/day for 2 years (Ellis et al. 1979). A statistically significant increase in renal tumors in female mice was not observed. A National Cancer Institute (NCI) bioassay (NCI 1978) of 2,4-DNT (95% 2,4-DNT, the other components not specified) did not detect a carcinogenic effect in mice dosed with 72 mg/kg/day for 78 weeks. The NCI bioassay used the C57BL/6N strain of mouse, lower doses, and a shorter treatment schedule than did Ellis et al. (1979).

Hepatocellular carcinoma were significantly increased in male CD rats fed 34.5 mg/kg/day 2,4-DNT and in females fed 45.3 mg/kg/day 2,4-DNT for 2 years (Ellis et al. 1979). The tumor response in females was higher than in the males. Two other studies of rats in which malignancies were not observed used the Fischer-344 strain, lower doses, and shorter exposure durations than did Ellis et al. (1979): 10 mg/kg/day for 78 weeks (NCI 1978) and 27 mg/kg/day for 52 weeks (Leonard et al. 1987). NCI (1978) reported significant increases in subcutaneous tissue fibroma in male rats at 7.5-8 mg/kg/day and mammary gland fibroadenomas in female rats at 22 mg/kg/day. Ellis et al. (1979) found significant increases in subcutaneous tissue fibromas in male rats at 34.5 mg/kg/day and mammary gland fibroadenomas in female rats at 45.3 mg/kg/day; these were benign tumors.

2,4-DNT was not found to be carcinogenic in the Strain A/J mouse pulmonary tumor bioassay when 250 mg/kg was administered by gavage twice a week for 12 weeks (Stoner et al. 1984). 2,4-DNT was a hepatic tumor promoter, but not a tumor initiator, using *in vivo* hepatic initiation-promotion protocols (Leonard et al. 1986).

2,6-DNT administered for 1 year at 7 and 14 mg/kg/day produced hepatocellular carcinomas in 85% and 100%, respectively, of male Fischer-344 rats (Leonard et al. 1987). Pulmonary metastases of hepatocytic origin were also observed. Both tumor-initiating and tumor-promoting activities of 2,6-DNT in rat liver were reported (Leonard et al. 1983, 1986; Mirsalis and Butterworth 1982). 2,6-DNT was not found to be a lung carcinogen in the Strain A/J mouse pulmonary tumor bioassay when 250 mg/kg was administered by gavage twice a week for 12 weeks (Schut et al. 1983; Stoner et al. 1984).

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The effect of diet-induced changes in gut microflora on the hepatocarcinogenicity of 2,6-DNT was studied in male F344 rats (Goldsworthy et al. 1986). Groups of the rats were placed one of three diets containing 2,6-DNT at doses of 0, 0.6-0.7 or 3-3.5 mg/kg/day. Ten animals from each group were sacrificed at 3,6, and 12 months, and the livers were evaluated histopathologically. The diets used were NIH-07, an open formula cereal-based diet high in pectin content; AIN-76A, a purified pectin-free diet; or AR, which is AIN-76A supplemented with 5% pectin. The number and size of γ -glutamyl transpeptidase-staining foci in the liver increased in a dose- and time-dependent manner in animals given 2,6-DNT in the NM-07 diet. Hepatocellular carcinomas and neoplastic nodules were observed only in rats fed NM-07 containing 2,6-DNT. No tumor was observed in rats receiving the control diets or 2,6-DNT in the AIN-76 diet with or without pectin. This finding suggested that pectin did not influence the tumor outcome of the experiment. Unidentified contaminants in cereal-based diets may influence liver foci and tumor production in the rat liver during carcinogen treatment.

Tg-DNT provided positive hepatocarcinogenic results in two bioassays of less-than-lifetime duration. In a 52-week study of male rats dosed with 35 mg/kg/day of Tg-DNT, Leonard et al. (1987) observed a 47% increase in hepatocellular carcinoma; cholangiocarcinomas were also found in 10% of rats treated with 35 mg/kg/day Tg-DNT in the Leonard et al. (1987) study. Hazleton Laboratories (1982) reported that dietary administration of 35 mg/kg/day Tg-DNT to rats for 55 weeks resulted in an increased incidence (100% in males and 55% in females) of hepatocellular carcinoma; this lesion was found in some animals treated at this level for 26 weeks. The administration of 3.5 mg/kg/day Tg-DNT for 104 weeks caused hepatocellular carcinoma in 9 of 70 males compared to 1 of 61 controls. Mammary fibroadenoma and subcutaneous fibroma were also found in both sexes at 3.5 mg/kg/day after 104 weeks (Hazleton Laboratories 1982). Administration of 14 mg/kg/day Tg-DNT for 104 weeks caused cholangiocarcinomas and parathyroid adenomas in males and hepatocellular carcinomas and hepatocholangiocarcinomas in females (Hazleton Laboratories 1982).

Tg-DNT contains about 76% 2,4-DNT and 19% 2,6-DNT, as well as small amounts of other isomers. Rats that received 35 mg/kg/day in the Leonard et al. (1987) and Hazleton Laboratories (1982) studies were provided approximately 28 and 7 mg/kg/day of 2,4- and 2,6-DNT, respectively. This dose of 2,6-DNT in Tg-DNT bioassays is equivalent to the low dose of 2,6-DNT administered to rats by Leonard et al. (1987) that produced hepatocellular carcinomas.

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In hepatic tumor initiation-promotion protocols, Tg-DNT was reported to have tumor promoting and tumor initiating activity (Leonard et al. 1983, 1986; Mirsalis and Butterworth 1982). The results of the initiationpromotion protocols for 2,4-, 2,6-, and Tg-DNT indicate that 2,6-DNT is a complete hepatocarcinogen and is primarily responsible for the carcinogenic activity of Tg-DNT.

2.2.3 Dermal Exposure

There are data on occupational exposure of humans to 2,4-DNT and Tg-DNT (see Section 2.2.1) in which dermal exposure probably occurred, but the primary route of exposure in these studies is believed to be inhalation. The relative contribution of dermal exposure to total occupational exposure cannot be determined from these studies. Levine et al. (1985b) reported that small amounts of 2,4-DNT were detected on the hands, face, and forehead when a wipe-sample survey was conducted on workers in a DNT manufacturing plant. The highest quantity found on a worker's skin was 180 µG and may account for the quantity of excreted urinary metabolites that exceeded the amount of inhaled DNT in the operators and loaders.

2.2.3.1 Death

One study was located that examined death among humans exposed to DNT. A retrospective mortality study of munitions workers exposed to either 2,4-DNT or Tg-DNT revealed an increased death rate due to ischemic heart disease and residual diseases of the circulatory system in the exposed cohort (Levine et al. 1986a, 1986b). The residual diseases included cardiac arrest and arteriosclerosis. Exposure levels were not reported, and the study is further limited by the small cohort size and concurrent inhalation exposure of the workers.

No studies were located regarding death in animals after dermal exposure to 2,4- or 2,6-DNT.

2.2.3.2 Systemic Effects

No studies were located regarding respiratory, cardiovascular, renal, body weight, or endocrine effects in humans or animals after dermal exposure to 2,4-DNT or 2,6-DNT.

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Gastrointestinal Effects. Gastrointestinal complaints of munitions workers exposed to either 2,4-DNT or Tg-DNT included nausea and vomiting (McGee et al. 1942, 1947). These workers also presumably inhaled DNT in the occupational setting.

No studies were located regarding gastrointestinal effects in animals after dermal exposure to 2,4-DNT or 2,6-DNT.

Hematological Effects. Hematological effects, such as anemia and cyanosis, have been found in men employed at munitions factories (McGee et al. 1942, 1947; Perkins 1919). These workers were exposed to either 2,4-DNT or Tg-DNT. Because these studies lacked worker histories, exposure data, and reported on small cohorts, the results are equivocal and are best used to qualitatively describe symptoms. In addition, the workers probably received their primary exposure via the inhalation pathway.

No studies were located regarding hematological effects in animals after dermal exposure to 2,4-DNT or 2,6-DNT.

Musculoskeletal Effects. Muscle weakness and joint pain have been reported by munitions workers after occupational exposure to unspecified concentrations of 2,4-DNT or Tg-DNT (McGee et al. 1942; Perkins 1919).

In the Perkins (1919) study, joint pain and other incapacitating symptoms were noted following exposure to what were presumed to be very high concentrations of Tg-DNT since the processes described required direct handling without protective equipment. In both of these studies, however, no exposure data were available; exposure to other compounds may have occurred, and concomitant exposure via inhalation was also likely.

No studies were located regarding musculoskeletal effects in animals after dermal exposure to 2,4-DNT or 2,6-DNT.

Hepatic Effects. In a follow-up study of male munitions workers exposed to unspecified concentrations of 2,4-DNT, 29 of 714 workers displayed tenderness of the liver (McGee et al. 1947). No other clinical evaluation was performed that might provide further insight into the significance of this finding. These workers were also exposed to DNT via inhalation.

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No studies were located regarding hepatic effects in animals after dermal exposure to 2,4- or 2,6-DNT.

Dermal Effects. The only human studies that noted dermal effects upon topical exposure to 2,4-DNT were of workers employed by a munitions factory during World War II. The first study found 6 of 154 workers who complained of dermatitis, which the authors attributed to DNT exposure (McGee et al. 1942). A followup study, conducted after changes in the manufacturing process designed to reduce DNT exposure were implemented, reported that 32 of 714 workers complained of dermatitis (McGee et al. 1947). Exposure levels were not quantified in either of these studies.

Both 2,4- and 2,6-DNT were shown to be mild primary dermal irritants in rabbits (Ellis et al. 1978; Lee et al. 1975).

Ocular Effects. No studies were located regarding ocular effects in humans after dermal exposure to 2,4- or 2,6-DNT.

No ocular irritation was found in rabbits in a primary eye irritation test using unspecified concentrations of 2,4- or 2,6-DNT (Ellis et al. 1978; Lee et al. 1975). However mild eye irritations were reported in rabbits treated with 2,4- and Tg-DNT (Ford 1981; Henry 1982).

2.2.3.3 Immunological and Lymphoreticular Effects

No studies were located regarding immunological and lymphoreticular effects in humans after dermal exposure to 2,4-DNT or 2,6-DNT.

In dermal sensitization tests, 2 of 10 guinea pigs exhibited mild sensitization to 2,6-DNT, but no sensitization was evident when 2,4-DNT was tested (Ellis et al. 1978; Lee et al. 1975).

2.2.3.4 Neurological Effects

Various neurological symptoms, including headache, vertigo, and pain or numbness in the extremities, have been reported in surveys of munitions workers exposed to unspecified concentrations of 2,4-DNT (McGee et al. 1942, 1947). Although it is assumed that some dermal exposure to 2,4-DNT occurred in these workers, inhalation was the probable primary route of exposure.

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No studies were located regarding neurological effects in animals after dermal exposure to 2,4- or 2,6-DNT.

No studies were located regarding the following health effects in humans or animals after dermal exposure to 2,4- or 2,6-DNT:

2.2.3.5 Reproductive Effects

2.2.3.6 Developmental Effects

2.2.3.7 Genotoxic Effects

Genotoxicity studies are discussed in Section 2.5.

2.2.3.8 Cancer

No studies were located regarding cancer in humans or animals after dermal exposure to 2,4- or 2,6-DNT.

2.3 TOXICOKINETICS

2.3.1 Absorption

No information regarding absorption of DNT in children has been located.

2.3.1.1 Inhalation Exposure

There are no available studies in which the absorption rates of inhaled 2,4- or 2,6-DNT in humans or laboratory animals have been evaluated. However, based on analyses of the urinary metabolites of workers in DNT manufacturing plants(Levine et al. 1985b; Turner 1986; Woollen et al. 1985), it is apparent that dermal absorption of these two compounds does occur under occupational settings.

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2.3.1.2 Oral Exposure

Rickert et al. (1983) suggested that the rapid disappearance of radioactivity from the first quarter of the small intestine of rats following the oral administration of uniformly [^{14}C]-ring-labeled 2,4- or 2,6-DNT indicates rapid and fairly complete absorption.

Excretion data and observed systemic effects indicate that DNT is absorbed following oral administration to experimental animals. Several strains of rats, New Zealand rabbits, beagle dogs, and rhesus monkeys excreted 55-90% of the radioactivity from orally-administered radiolabeled DNT in the urine, primarily within the first 24 hours (Lee et al. 1978; Long and Rickert 1982; Rickert and Long 1981). In mice, most of the radioactivity from ^3H -labeled 2,6-DNT was excreted in the urine (about 50% in 8 hours) (Schut et al. 1983), whereas most of the radioactivity from ^{14}C -labeled 2,4-DNT administered to mice was excreted in the feces, and only about 10% in the urine (Lee et al. 1978). Increased fecal excretion could be due to reduced absorption or to greater excretion via the bile.

2.3.1.3 Dermal Exposure

There were no data available specifically on the absorption of 2,4- or 2,6-DNT via the dermal route of exposure. Two studies of occupational exposure to Tg-DNT have suggested that dermal absorption can be a significant route of entry for these isomers in humans since the levels of urinary metabolites of 2,4- and 2,6-DNT in loaders and operators at a DNT manufacturing plant exceeded those that would have resulted from the inhaled concentrations (Levine et al. 1985b; Woollen et al. 1985).

2.3.2 Distribution

No information regarding distribution of DNT in children has been located.

2.3.2.1 Inhalation Exposure

No studies were located regarding distribution in humans or animals following inhalation exposure to 2,4- or 2,6-DNT.

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2.3.2.2 Oral Exposure

The tissue distribution of 2,4-DNT and its metabolites was studied by Rickert and Long (1980). 2,4-DNT was administered orally to male and female rats at doses of 10, 35, or 100 mg of ¹⁴C-labeled 2,4-DNT per kilogram. When distribution is studied solely by detecting a radioisotope label, it is the labeled atom(s) which are being followed and this label may be part of either the parent DNT molecule or a metabolite. Peak concentrations of radioactivity in plasma, red blood cells, liver, and kidney were proportional to dose. Levels in liver and kidney were 5-10 times higher than those in plasma or red blood cells. Levels of radioactivity in other tissues were lower than those in plasma. The only clear differences between males and females were the higher retention of radioactivity in red blood cells of females and the concentration of radioactivity in livers of females, which was only half that found in males. In addition, concentrations of 2,4-DNT in male kidneys peaked at 4-8 hours and were 3-10 times higher than the concentrations in female kidneys which peaked 1 hour after the dose.

Rickert et al. (1983) observed that hepatic concentrations of radioactivity in male rats increased in 2 stages, with the first peak occurring 1-2 hours and a second peak occurring 8-12 hours after an oral dose of 10 or 35 mg/kg of radiolabeled 2,4- or 2,6-DNT. The second peak was followed by a gradual decline up to 16 days and was thought to be the result of enterohepatic cycling.

In mice administered ³H-labeled 2,6-DNT, the distribution of the label was similar in the blood, liver, kidneys, lungs, and small and large intestines at 8 hours after administration, with very low levels detected in the brain, lungs, heart, and spleen (Schut et al. 1983).

In a radioisotope labeling study in dogs and monkeys, total 2,4-DNT and its metabolites recovered in blood and other tissues were approximately 3.6% (dogs) and 2.2% (monkeys) of the administered dose (Lee et al. 1978). Relative to blood concentrations, the liver had the highest levels of 2,4-DNT or metabolites.

Detectable levels of 2,4-DNT or metabolites were also found in the kidney and in skeletal muscle (Lee et al. 1975, 1978).

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2.3.2.3 Dermal Exposure

No studies were located regarding distribution in humans or animals following dermal exposure to 2,4- or 2,6-DNT.

2.3.3 Metabolism

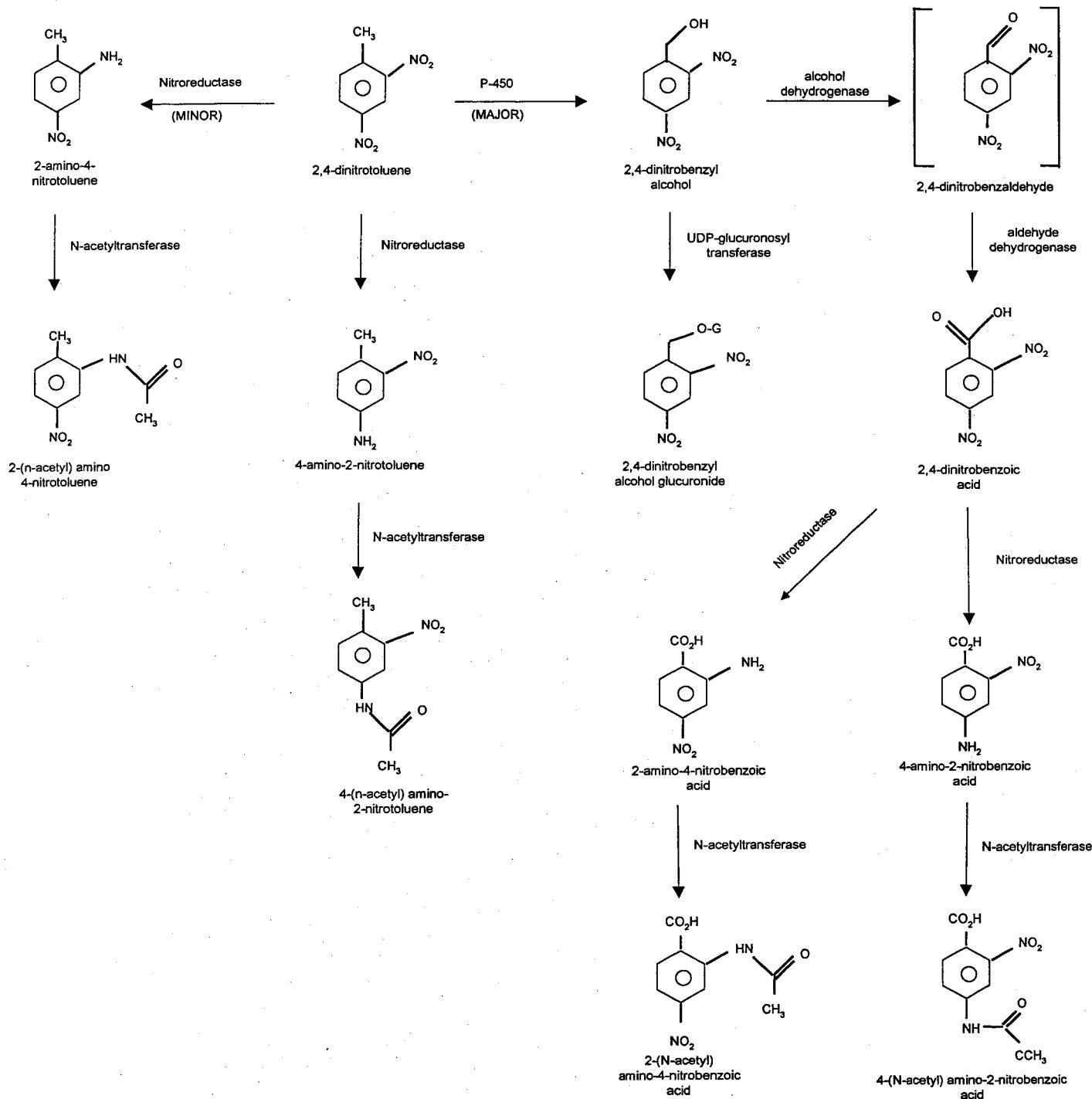
The metabolism of DNT in humans has been studied in workers exposed to Tg-DNT by the analysis of urinary metabolites. The routes of exposure in these studies were multiple. Since the amounts of metabolites excreted could not be accounted for by the inhalation exposure route alone, dermal contact and ingestion routes of exposure may also be of importance (Levine et al. 1985b; Woollen et al. 1985). Woollen et al. (1985) found that the major metabolite excreted in the urine of workers exposed to Tg-DNT was 2,4-dinitrobenzoic acid (conjugates were hydrolyzed before analysis). There were wide variations in the excretion of the metabolites in different workers. Concentrations of 2,4-dinitrobenzoic acid in end-of-shift urine samples from 20 male and 8 female workers, however, did not suggest a difference in the excretion of this metabolite between males and females. The study authors stated that lesser amounts of the following metabolites were also found in the urine: 2-amino-4-nitro-, 4-amino-2-nitro-, and 2-amino-6-nitrobenzoic acids, and 4-(Nacetyl)amino-2-nitrobenzoic acid. Trace levels of DNT were also detected. Dinitrobenzyl alcohols were not detected. Neither amounts nor relative percentages of metabolites were reported.

Studies of workers at a Tg-DNT manufacturing plant (Levine et al. 1985b; Turner et al. 1985) provide more detailed information regarding the metabolism of Tg-DNT in occupationally exposed men and women. The principal metabolites detected in the urine of 14 men were dinitrobenzoic acids (2,4- and 2,6-) and 2-amino-4-nitrobenzoic acid. In the urine of three women, these metabolites were detected together with dinitrobenzyl alcohol glucuronides (2,4- and 2,6-). Expressed as percent of total urinary metabolites, the dinitrobenzoic acids, 2-amino-4-nitrobenzoic acid, and the dinitrobenzyl glucuronides constituted 52.5, 37.2, and 9.5%, respectively, of the total urinary DNT metabolites in men and 28.8, 37.6 and 33.3%, respectively, of the total urinary DNT metabolite in women. 2,4- and 2,6-DNT metabolites were present in roughly the same proportions as in the Tg-DNT. Both men and women excreted relatively small amounts (less than 1% of urinary metabolites) of 2-(N-acetyl)amino-4nitrobenzoic acid (Levine et al. 1985b).

Studies in rats have identified a complex pathway for the metabolism of 2,4-DNT (Figures 2-3 and 2-4) and 2,6-DNT (Figure 2-5). Metabolism occurs in the liver and also in the intestine by microflora (Long and

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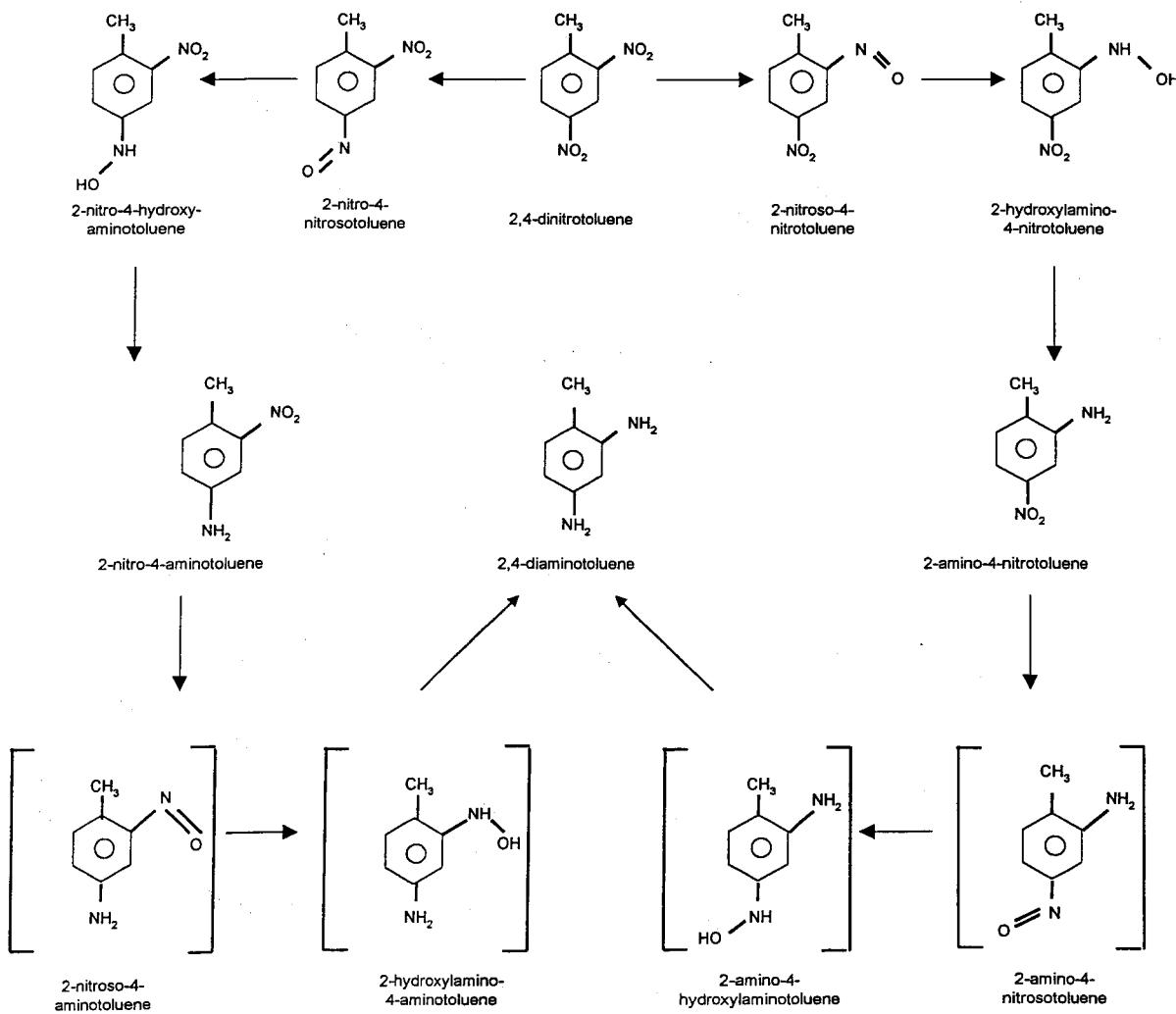
Figure 2-3. Proposed Metabolic Pathways for the Hepatic Metabolism of 2,4-DNT*



*Sources: Bond and Rickert 1981; Bond et al. 1981; Smith et al. 1995

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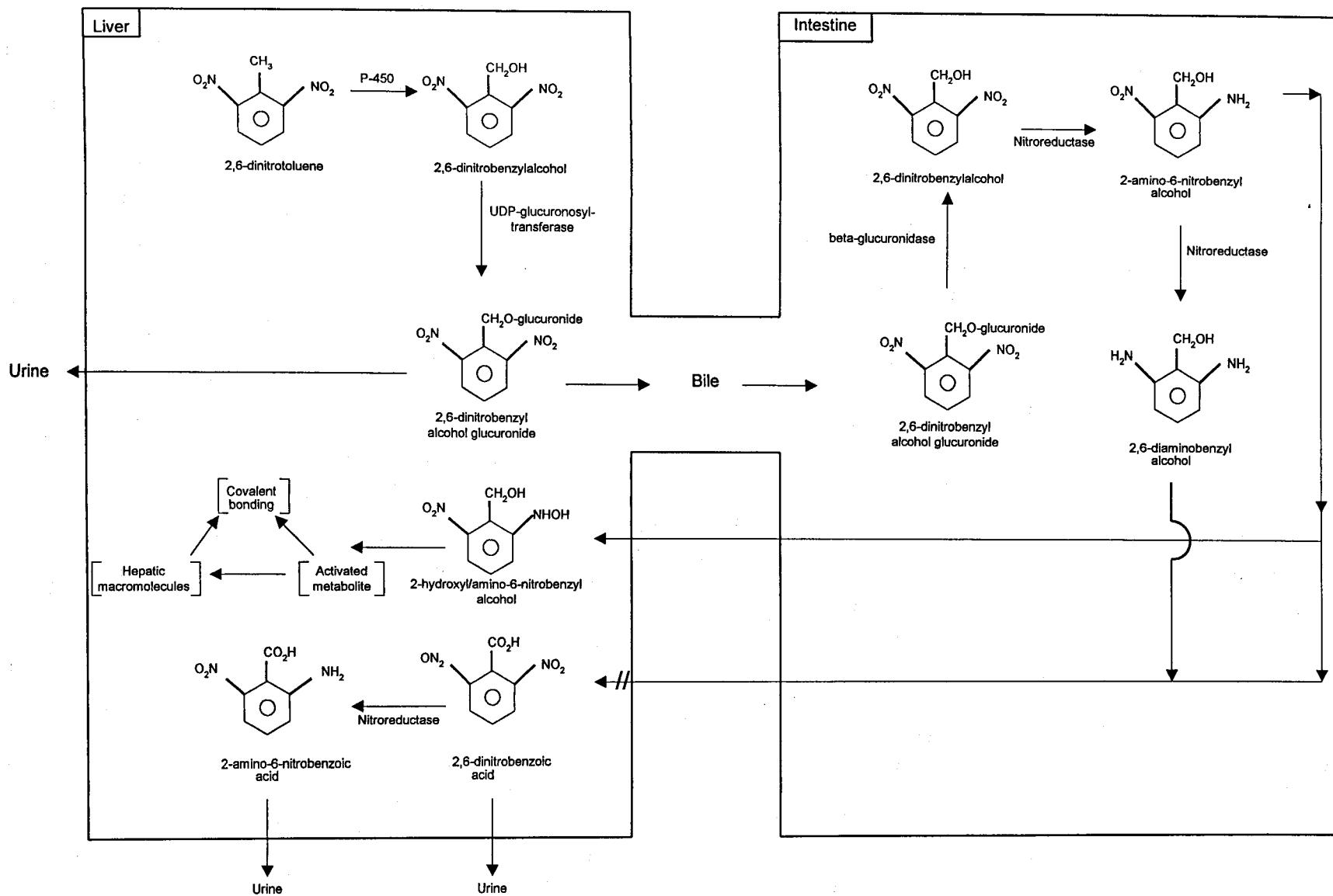
**Figure 2-4. Proposed Pathways
for the Anaerobic Metabolism of 2,4-DNT in Rat Intestinal
Microflora***



→ = Nitroreductase reaction

*Sources: Guest et al. 1982; Mori et al. 1985

Figure 2-5. Proposed Pathways for Metabolism of 2,6-DNT*



*Sources: Chapman et al. 1993; La and Froines 1993; Rickert et al. 1984; Smith et al. 1995

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Rickert 1982; Rickert et al. 1981). Both oxidized and reduced metabolites are excreted in the urine after oral administration of the compounds. The main urinary metabolites of 2,4- and 2,6-DNT are the corresponding dinitrobenzyl alcohol glucuronide, dinitrobenzoic acid, and aminonitrobenzoic acid (Long and Rickert 1982). An additional urinary metabolite of 2,4-DNT is 4-(N-acetyl)amino-2nitrobenzoic acid (Rickert et al. 1981).

Oxidative metabolism by cytochrome P450 predominates in the liver of experimental animals, leading to the formation of dinitrobenzyl alcohol which is either converted to glucuronide conjugate or further oxidized to dinitrobenzoic acid. Dinitrobenzyl alcohol glucuronide is partially excreted into the bile, followed by metabolism by gut microflora and enterohepatic cycling (Long and Rickert 1982; Medinsky and Dent 1983; Mori et al. 1997; Rickert and Long 1981). Thus, DNT appears to be first metabolized by the liver with the metabolites being excreted into the bile; the biliary metabolites are hydrolyzed and further metabolized in the intestine; after reabsorption and circulation back to the liver, the metabolites are activated and bound to macromolecules (Chadwick et al. 1993; Long and Rickert 1982).

2,4- and 2,6-dinitrobenzyl glucuronide have been detected directly in the bile following administration of 2,4- and 2,6-DNT to the male Wistar rat (Mori et al. 1997), accounting for about 35 and 51% of the dose respectively. Four other metabolites, 2-amino-4-nitrotoluene, 4-amino-2-nitrotoluene, 2,4-diaminotoluene, and 4-acetylamino-2-nitrobenzoic acid accounted for 0.02-0.12% of the dose; in addition to 2,4-dinitrobenzyl alcohol, 2,4-dinitrobenzaldehyde and 2,4-dinitrobenzoic acid (0.09-0.14%) were detected in the bile of rats given 2,4-DNT. 2,6-dinitrobenzyl alcohol, 2-amino-6-nitrotoluene, and 2,6-dinitrobenzaldehyde were detected in the bile of rats given 2,6-DNT.

Studies of the metabolism of 2,4-DNT by intestinal microflora in rats and mice (Guest et al. 1982; Mori et al. 1985) and studies in germ-free rats (Rickert et al. 1981) have shown that intestinal microflora are responsible for reductive metabolism of DNT. Intestinal microorganisms hydrolyze and reduce 2,4- and 2,6-dinitrobenzyl alcohol glucuronide to the corresponding aminonitrotoluenes, probably through nitroso derivatives and hydroxylamino derivatives (Mori et al. 1997). The deconjugated metabolites are reabsorbed and transported back to the liver by enterohepatic circulation (Medinsky and Dent 1983). In the liver, the newly formed amine group is N-hydroxylated by cytochrome P450 and conjugated with sulfate (Kedderis et al. 1984). The sulfate conjugate is unstable and can be decomposed to form a carbonium or nitrenium ion that can be bound to hepatic macromolecules; this ostensibly leads to mutations and the formation of liver tumors. Thus, sulfation may be involved in the initiation stage of hepatocarcinogenesis by 2,6-DNT. Metabolism by

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intestinal microflora appears to be essential for the production of metabolites that bind covalently to liver macromolecules.

The intestinal biotransformation of 2,6-DNT was investigated *in vitro* using suspended microflora preparation from the intestinal contents of male Wistar rats (Sayama et al. 1993). It was determined that the metabolites formed with the incubation of 2,6-DNT were 2-nitroso-, 2-hydroxyl amino-, and 2-amino-6-nitrotoluene and 2,6-diaminotoluene. Since no metabolites were detected when 2,6-diaminotoluene was incubated and the recovery of 2,6-diaminotoluene was about 95%, it appears that 2,6-diaminotoluene is the terminal intestinal metabolite of 2,6-DNT (Sayama et al. 1993). When 2,4-DNT was examined in this system, two nitroazoxy compounds (2,2'-dimethyl-5-5'-dinitroazoxybenzene and 4,4'-dimethyl-3,3'-dinitroazoxybenzene) were detected in addition to other known metabolites, such as nitrosonitrotoluenes, hydroxyl aminonitrotoluenes, aminonitrotoluenes, and diaminotoluene (Sayama et al. 1993). The nitroazoxy compounds were believed to be non-enzymatic products (Sayama et al. 1993).

The metabolites formed by the anaerobic incubation of potassium 2,4-dinitrobenzyl glucuronide or potassium 2,6-dinitrobenzyl glucuronide with rat intestinal microflora have been examined (Mori et al. 1997). Metabolites transformed from 2,4-dinitrobenzyl glucuronide were 2,4-dinitrobenzyl alcohol, 4-amino-2-nitrobenzyl alcohol, and 2-amino-4-nitrobenzyl alcohol, which peaked at 30,75, and 120 minutes of the incubation. 2,6-Dinitrobenzyl alcohol and 2-amino-6-nitrobenzyl alcohol were detected from potassium 2,6-dinitrobenzyl glucuronide incubation. Thus, intestinal metabolism includes the deconjugation of the glucuronide and the reduction of the nitro compound.

In rats, sex differences in the metabolism of 2,4-DNT have been observed. A larger percentage of the administered dose is excreted in the bile of male rats than is excreted in the bile of females. In females, a greater percentage of the dose is excreted in urine as the dinitrobenzyl alcohol glucuronide (Medinsky and Dent 1983; Rickert and Long 1981). The quantitative differences in urinary versus biliary excretion of the glucuronide conjugates by females may account for the sex differences in the susceptibility of the rat to the hepatocarcinogenic effects of 2,4-DNT (Ellis et al. 1979). Greater urinary excretion may decrease the amount of the glucuronide available to the intestinal microflora for metabolism to a carcinogenic metabolite.

Metabolism studies in rats, rabbits, dogs, and monkeys with 2,4-DNT revealed the major urinary metabolites as glucuronide conjugates of 2,4-dinitrobenzyl alcohol (20-33% of the dose) and 2,4-aminonitro-benzyl alcohols (8-19% of the dose). Lesser amounts of aminonitrotoluene, 2,4-diaminotoluene, 2,4-aminobenzyl

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alcohol, and 2,4-dinitrobenzoic acid were also identified in all four species. Mice were also evaluated in this same group of studies. In mice, approximately 3% of the administered dose was excreted in the urine as the glucuronide conjugate of the 2,4-dinitrobenzyl alcohol and approximately 3% as the glucuronide conjugates of 2,4-aminonitrobenzyl alcohol (Lee et al. 1978).

Another study using rats dosed with either 2,4- or 2,6-DNT also demonstrated that the primary urinary conjugate was the respective dinitrobenzyl glucuronide (11-17% of administered dose) (Mori et al. 1996). Other metabolites in rats administered 2,4-DNT included 2-amino-4-nitrobenzoic acid (0.71%), 4-amino-2-nitrobenzoic acid (0.52%), 4-acetylamino-2-nitrobenzoic acid (3.9%), 4-amino-2-nitrotoluene (0.04%), 2,4-dinitrobenzyl alcohol (0.25%), 2,4-dinitrobenzoic acid (6.9%), and 4-acetylamino-2-aminobenzoic acid (3.4%). After administration of 2,6-DNT, other metabolites in urine included 2,6-dinitrobenzoic acid (0.17%), 2-amino-6-nitrotoluene (0.44%), and 2,6-dinitrobenzyl alcohol (0.53%) (Mori et al. 1996).

The urinary metabolites of DNT and probably the glucuronides resulting from occupational exposure of humans are qualitatively the same as those resulting from oral administration to rats, but the proportions of nitro-reduced metabolites were lower relative to oxidized metabolites in the urine from humans (Turner et al. 1985). These differences may be due more to the particular routes of exposure (inhalation and dermal for humans; oral for rats) than differences in species. As seen in experimental animals, female subjects excreted a higher proportion of urinary metabolites as dinitrobenzyl alcohol glucuronides than did males.

Metabolism of DNT has not been studied in children. However, fetuses and neonates have been shown to be limited in their ability to biotransform xenobiotics. Although the cytochrome P-450 isoforms responsible for DNT metabolism have not been identified, cytochromes CYP2E1, CYP2B 1/2, and CYP2C1 1/6 are known to contribute to the side-chain oxidation of toluene by the rat liver, and multiple cytochrome P-450 isoforms may contribute to the side-chain oxidation of DNT (Chapman et al. 1993). In humans, CYP2E1 protein is absent from fetal and neonatal livers, but steadily increases during the first year of life (Vieira et al. 1996). Other isoforms' expression in fetuses and neonates is also qualitatively and quantitatively different from the expression observed in adults (Komori et al. 1990; Leeder and Kearns 1997). In rats, while sulfotransferase (the enzyme which catalyzes sulfation) activity is almost at adult levels at birth, UDP-glucuronosyltransferase (the enzyme which produces glucuronide conjugates) activity towards different xenobiotics varies with maturation (Young and Lietman 1978). Similarly, in humans, sulfation capabilities develop faster than glucuronidation capabilities (Leeder and Keams 1997). While the activity of some isoforms of sulfotransferase may exceed those seen in adults during infancy and early childhood, the activity of UDP-glucuronosyltransferase

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depends on the specific isoforms of the enzyme, and adult levels are generally attained by 6-18 months (Leeder and Kearns 1997). Since DNT undergoes bioactivation in the liver and by the intestinal microflora, the toxicity of DNT may be different in children. Newborns have a transient deficiency in methemoglobin reductase (Gruener 1976) and have a high concentration of fetal hemoglobin in their erythrocytes. Consequently, they are highly sensitive to methemoglobin-generating chemicals and to methemoglobinemia generated by DNT.

2.3.4 Elimination and Excretion

Information regarding excretion of DNT in children was not located.

2.3.4.1 Inhalation Exposure

In occupational settings, in addition to inhalation, some oral and dermal exposure can occur. The elimination of DNT in the urine of workers exposed to Tg-DNT has been studied by several investigators (Levine et al. 1985b; Turner et al. 1985; Woollen et al. 1985).

Woollen et al. (1985) observed that the highest rates of excretion of 2,4-dinitrobenzoic acid occurred near the end of the work shift. The half-life for urinary excretion of 2,4-dinitrobenzoic acid was calculated to be 2-5 hours. This estimate appears to be the initial phase of a biphasic elimination profile since even 3 days after the last exposure, detectable levels of 2,4-dinitrobenzoic acid were present in urine.

Turner et al. (1985) determined the metabolic profiles in workers exposed to DNT. The half-life for excretion of DNT metabolites in urine ranged from 0.8 to 4.5 hours. The half-lives for 2,4-dinitrobenzoic acid and 2,4-dinitrobenzyl alcohol glucuronide tended to be shorter than those for the metabolites that resulted from both oxidative and reductive metabolism.

2.3.4.2 Oral Exposure

No studies were located regarding excretion in humans following oral exposure to 2,4- or 2,6-DNT.

Schut et al. (1983) reported that in mice, urine was the main route of elimination of ^3H -labeled 2,6-DNT, with about 50% excreted after 8 hours. Lee et al. (1978) observed that most of the radioactivity from ^{14}C -labeled

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2,4-DNT administered to mice was excreted in the feces and only about 10% in the urine. Differences between these two studies could be due, in part, to the use of different species of mice.

Male and female rats excreted 55-90% of the radioactivity from ^{14}C -2,4-DNT or ^{14}C -2,6-DNT in the urine, and 15-30% in the feces, within 72 hours after dosing (Long and Rickert 1982; Rickert and Long 1981). With 2,4-DNT, the females excreted a greater percentage of the dose in the urine as 2,4-dinitrobenzylalcohol glucuronide than did the males (except at the highest dose), but with 2,6-DNT, no sex-related difference in urinary excretion was seen.

In experiments with bile duct-cannulated rats, male rats excreted 25% of the radioactivity from ^{14}C -2,4-DNT into the bile over a 36-hour period, whereas female rats excreted 18% (Medinsky and Dent 1983). Biliary excretion of radioactivity was linearly related to dose in males; females were evaluated only at one dose. Biliary excretion of radioactivity was virtually complete within 24 hours for males and 12 hours for females. Mean half-times of biliary excretion ranged from 3.3 to 5.3 hours. Urinary excretion was also significant, with greater amounts of radioactivity excreted in the urine of rats from which bile was not collected (60-90% of the dose) than in the urine of rats from which bile was collected (20-60% of the dose). This finding indicates that biliary metabolites were absorbed from the intestines (enterohepatic cycling). Whether or not bile was collected, female rats excreted more radioactivity in urine than did male rats. Greater than 90% of the urinary excretion of labeled metabolites appeared in urine collected during the first 24 hours. At the end of 36 hours, only 0.02-0.05% of the radioactivity was detectable in the livers; 20-60% of this was covalently bound.

2.3.4.3 Dermal Exposure

There are no kinetic data in humans in which the route of exposure was specifically dermal. Occupational exposure studies available for Tg-DNT involved multiple routes of exposure (Levine et al. 1985b; Turner et al. 1985; Woollen et al. 1985). The major routes of exposure in these studies were considered to be inhalation and dermal. The results were discussed previously in Section 2.3.4.1.

No studies were located regarding excretion in animals following dermal exposure to 2,4- or 2,6-DNT.

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2.3.5 Physiologically Based Pharmacokinetic (PBPK)/Pharmacodynamic (PD) Models

Physiologically based pharmacokinetic (PBPK) models use mathematical descriptions of the uptake and disposition of chemical substances to quantitatively describe the relationships among critical biological processes (Krishnan et al. 1994). PBPK models are also called biologically based tissue dosimetry models. PBPK models are increasingly used in risk assessments, primarily to predict the concentration of potentially toxic moieties of a chemical that will be delivered to any given target tissue following various combinations of route, dose level, and test species (Clewell and Andersen 1985). Physiologically based pharmacodynamic (PBPD) models use mathematical descriptions of the dose-response function to quantitatively describe the relationship between target tissue dose and toxic end points.

PBPK/PD models refine our understanding of complex quantitative dose behaviors by helping to delineate and characterize the relationships between: (1) the external exposure concentration and target tissue dose of the toxic moiety, and (2) the target tissue dose and observed responses (Andersen et al. 1987; Andersen and Ktishnan 1994). These models are biologically and mechanistically based and can be used to extrapolate the pharmacokinetic behavior of chemical substances from high to low dose, from route to route, between species, and between subpopulations within a species. The biological basis of PBPK models results in more meaningful extrapolations than those generated with the more conventional use of uncertainty factors.

The PBPK model for a chemical substance is developed in four interconnected steps: (1) model representation, (2) model parametrization, (3) model simulation, and (4) model validation (Krishnan and Andersen 1994). In the early 1990s, validated PBPK models were developed for a number of toxicologically important chemical substances, both volatile and nonvolatile (Krishnan and Andersen 1994; Leung 1993). PBPK models for a particular substance require estimates of the chemical substance-specific physicochemical parameters, and species-specific physiological and biological parameters. The numerical estimates of these model parameters are incorporated within a set of differential and algebraic equations that describe the pharmacokinetic processes. Solving these differential and algebraic equations provides the predictions of tissue dose. Computers then provide process simulations based on these solutions.

The structure and mathematical expressions used in PBPK models significantly simplify the true complexities of biological systems. If the uptake and disposition of the chemical substance(s) is adequately described, however, this simplification is desirable because data are often unavailable for many biological processes. A

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simplified scheme reduces the magnitude of cumulative uncertainty. The adequacy of the model is, therefore, of great importance, and model validation is essential to the use of PBPK models in risk assessment.

PBPK models improve the pharmacokinetic extrapolations used in risk assessments that identify the maximal (i.e., the safe) levels for human exposure to chemical substances (Andersen and Krishnan 1994). PBPK models provide a scientifically-sound means to predict the target tissue dose of chemicals in humans who are exposed to environmental levels (for example, levels that might occur at hazardous waste sites) based on the results of studies where doses were higher or were administered in different species. Figure 2-6 shows a conceptualized representation of a PBPK model.

If PBPK models for 2,4- and 2,6-dinitrotoluene exist, the overall results and individual models are discussed in this section in terms of their use in risk assessment, tissue dosimetry, and dose, route, and species extrapolations.

A PBPK model has not been developed for 2,4- or 2,6-DNT.

2.4 MECHANISMS OF ACTION

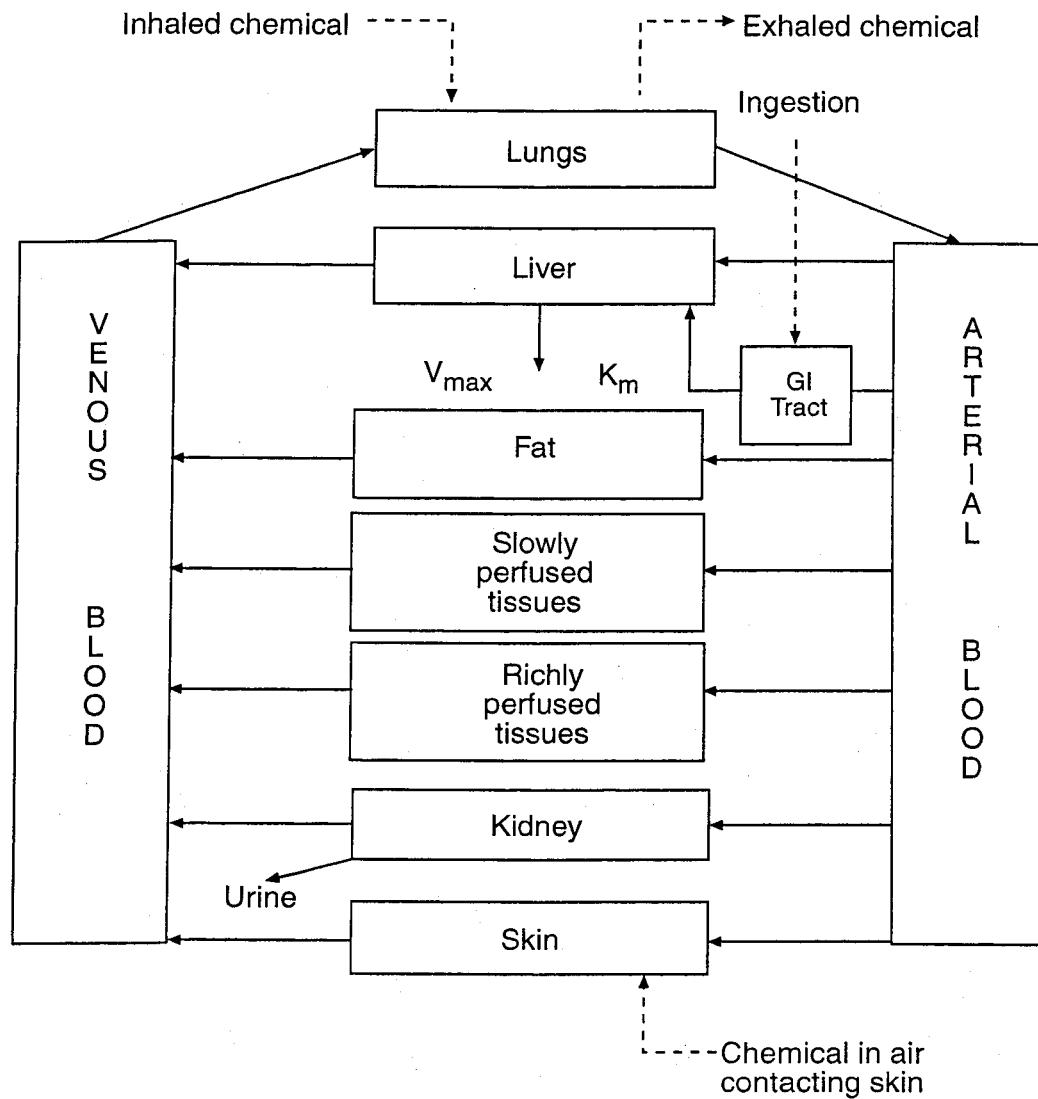
The mechanisms of action of DNT is not known to be different in children.

2.4.1 Pharmacokinetic Mechanisms

No information was located regarding the mechanism of absorption of 2,4- or 2,6-DNT. It is known that absorption occurs after inhalation exposure based on the metabolites found in the urine of workers at DNT manufacturing plants (Levine et al. 1985b; Turner 1986; Woollen et al. 1985). In studies of rats, rabbits, dogs, and monkeys, most orally administered 2,4- or 2,6-DNT has been shown to be absorbed (Lee et al. 1978; Long and Rickert 1982; Rickert and Long 1981). There appears to be minimal accumulation of these compounds after a single exposure. After repeated oral exposure in rats, 2,4-DNT and its metabolites were preferentially distributed to the liver, kidney, brain, lung, and skeletal muscle. The primary metabolite of 2,4-DNT excreted by humans exposed via inhalation and dermal routes of exposure in occupational studies or animals exposed via the oral route is 2,4-dinitrobenzyl alcohol and/or its glucuronide (EPA 1992). In addition to this, humans also excrete 2-amino-4-nitrobenzyl alcohol in the urine. 2,4-Dinitrobenzoic acid is another major metabolite (EPA 1992). Both 2-nitroso-4-nitrotoluene and 2-amino-4-nitrotoluene,

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Figure 2-6. Conceptual Representation of a Physiologically-Based Pharmacokinetic (PBPK) Model for a Hypothetical Chemical Substance



Note: This is a conceptual representation of a physiologically-based pharmacokinetic (PBPK) model for a hypothetical chemical substance. The chemical substance is shown to be absorbed via the skin, by inhalation, or by ingestion, metabolized in the liver, and excreted in the urine or by exhalation.

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metabolites of 2,4-DNT in humans, have been shown to be mutagenic in vitro (EPA 1992). It has been suggested that these intermediates may bind covalently to hepatic macromolecules, such as DNA and RNA (EPA 1992).

2.4.2 Mechanisms of Toxicity

Effects of Metabolism on Toxicity. The primary mechanism of toxicity for DNT involves bioactivation to form reactive intermediates (Kedderis et al. 1984; Sayama et al. 1989). Detailed information on the biotransformation of DNT is presented in Section 2.3.3. Briefly, metabolism of DNT begins in the liver, where it is oxidized by cytochrome P450 and conjugated with glucuronic acid to form the major metabolite dinitrobenzyl alcohol glucuronide and is excreted in bile or urine (Long and Rickert 1982; Medinsky and Dent 1983). The glucuronide excreted in bile undergoes biotransformation by intestinal microflora, where the conjugate is hydrolyzed and subsequently reduced by nitroreductase to the corresponding aminonitrobenzyl alcohol (Chadwick et al. 1993; Guest et al. 1982; Mori et al. 1985), probably through nitroso derivatives and hydroxylamino derivatives. The deconjugated metabolites are reabsorbed and transported back to the liver by enterohepatic circulation (Medinsky and Dent 1983). In the liver, the newly formed amine group is N-hydroxylated by cytochrome P450 and conjugated with sulfate (Kedderis et al. 1984). The sulfate conjugate is unstable and can be decomposed to form a carbonium or nitrenium ion that can be bound to hepatic macromolecules; this ostensibly leads to mutations and the formation of liver tumors. Thus, sulfation may be involved in the initiation stage of hepatocarcinogenesis by 2,6-DNT.

Target Organ Toxicity. The mechanism of toxicity of the hematological effects of DNT is described by Ellis et al. (1979). The effect of DNT on the blood is also produced by aromatic amines and most organic and inorganic nitrates. These compounds or their metabolites oxidize the ferrous ion in hemoglobin and produce methemoglobin. Hydroxylamine is probably the oxidizing species, because it is an intermediate in the reduction of nitro to amines. Within limits, the body can correct methemoglobinemia, but the corrective measures can be overwhelmed, producing numerous secondary effects including anoxia. The presence of methemoglobin leads to the formation of aggregates of hemoglobin degradation products called Heinz bodies. The presence of Heinz bodies is a sensitive indicator of blood toxicity as it indicates that some hemoglobin has been destroyed. High levels of methemoglobin are removed by catabolism, leading to the development of anemia. The body compensates for the destruction of red blood cells by increasing erythrocyte production, resulting in large numbers of immature erythrocytes, called reticulocytes, in the blood. If the toxic dose is not too severe, these compensatory mechanisms suffice. Thus, "compensated anemia," normal erythrocyte levels

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with reticulocytosis, may exist in exposed individuals. When the production of red blood cells can no longer keep pace with the hemolysis, frank anemia may be present (Ellis et al. 1979).

Carcinogenesis. In hepatic tumor initiation-promotion experiments, Tg-DNT was found to have tumor promoting and tumor-initiating activity (Leonard et al. 1983, 1986; Mirsalis and Butterworth 1982). 2,6 DNT was indicated to be a complete hepatocarcinogen and is primarily responsible for the carcinogenic activity of Tg-DNT. Hepatic DNA adducts have been detected by ³²P-postlabeling technique in 2,6-DNT-treated B6C3F₁ mice and Fischer 344 rats (George et al. 1996). 2,6-Dinitrobenzaldehyde, one of the metabolites of 2,6-DNT, was found to be a direct-acting mutagen in the *Salmonella typhimurium* strain TA98 and TA100 systems, not requiring metabolic activation by the S9 mix. 4-Amino-2-nitrobenzyl alcohol, 2-amino 4-nitrobenzyl alcohol, and 2-amino-6-nitrobenzyl alcohol are also mutagenic metabolites of 2,4- and 2,6-DNT, with their mutagenicity requiring metabolic activation (Mori et al. 1983; Sayama et al. 1989). Kedderis et al. (1984) proposed a bioactivation mechanism relating to the genotoxicity of 2,6-DNT in male Fischer 344 rats. They showed that the active metabolite of 2,6-DNT in the male Fischer 344 rat is the hydroxylamino sulfate of aminonitrobenzyl alcohol formed by the intestinal metabolism of benzyl glucuronide of 2,6-dinitrobenzyl alcohol excreted in bile. The sulfate conjugate is unstable, and the formation of electrophilic carbonium or nitrenium ions from these conjugates leads to subsequent binding to DNA.

2.4.3 Animal-to-Human Extrapolations

Correlation of toxic effects between humans and animals for 2,4- and 2,6-DNT with regard to hematologic and neurological effects has been noted (Ellis et al. 1979, 1985; Hong et al. 1985; Lane et al. 1985; Lee et al. 1978, 1985; McGee et al. 1942, 1947). Other effects for 2,4- and 2,6-DNT, such as reproductive, hepatic, renal, and cancer have been noted in animals (Ellis et al. 1979, 1985; Hong et al. 1985; Lee et al. 1976, 1978, 1985; Leonard et al. 1983, 1986; McGown et al. 1983; Stoner et al. 1984), but insufficient data are available to state definitively whether they are effects in humans. Two mutagenic metabolites of 2,4-DNT have been found in humans, mice, and rats (EPA 1992). Although rats appear to be more sensitive to the effects of 2,4- and 2,6-DNT than are mice (Ellis et al. 1978; Hong et al. 1985; Lane et al. 1985; Lee et al. 1975, 1978; Vemot et al. 1977), dogs appear to be the most sensitive of the three species (Ellis et al. 1979, 1985; Lee et al. 1976, 1978). However, limited intermediate-duration data using 2,6-DNT have shown mice to be more sensitive than rats (Lee et al. 1976). It should be noted that dogs were fed DNT by capsule in experimental studies, whereas the rodents received the test chemical in feed (Ellis et al. 1979, 1985; Hong et al. 1985; Lee et al. 1975, 1976, 1978, 1985).

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Extrapolating animal toxicity data to predict human risk from exposure to 2,4- and 2,6-DNT appears to be reasonable because of qualitative similarities in metabolism and known toxic effects.

2.5 RELEVANCE TO PUBLIC HEALTH

Issues relevant to children are explicitly discussed in 2.6 Children's Susceptibility and 5.6 Exposures of Children.

Overview.

The major route of exposure to DNT for humans living near hazardous waste sites is via ingestion of contaminated water. Because of its low octanol-water partition coefficient, DNT is not expected to accumulate in homegrown fruits and vegetables. Dermal exposure to DNT could also occur when washing or bathing with contaminated water. The low vapor pressure of DNT makes inhalation unlikely, although it could possibly be present in particulate matter in ambient air from leaks in storage containers or from contaminated soil.

Data for humans exposed to DNT are derived from occupational studies in which exposure concentrations via the inhalation route were measured. Some dermal exposure may be expected to occur in the workplace although this should be minimized with modern industrial hygiene practices. No studies were located regarding health effects in animals following inhalation exposure to DNT. No studies were located regarding health effects in humans following oral exposure to DNT. Available animal data on the toxic effects of DNT were derived from oral exposure studies. Discussions of health effects observed in animals and humans are thus complicated by the need to compare toxicity across different exposure routes. Thus, the effects observed from inhalation exposure studies in humans may not accurately reflect the effects that might be predicted after oral exposure to DNT near hazardous waste sites. However, there does appear to be some suggestive evidence of common target organs following exposure of animals and humans to DNT by different routes.

Possible effects of exposure of humans to primarily Tg-DNT include heart disease, hematological effects, and neurological effects.

In animal species, 2,4-DNT caused adverse effects in a variety of organs and tissues including the blood, nervous system, liver, kidney, and gonads. Subcutaneous and mammary gland carcinomas were also

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observed. 2,6-DNT affected the blood in all animal species tested. Effects on the liver or bile duct were generally observed, and males of all species exhibited adverse effects on the reproductive system. Two toxic effects, neurotoxicity and renal toxicity, were observed in dogs orally administered 2,6-DNT that were not observed in other animal species.

Minimal Risk Levels for 2,4- and 2,6-Dinitrotoluene

Inhalation MRLs.

Data were insufficient for the derivation of acute-, intermediate-, or chronic-duration inhalation MRLs for 2,4-DNT or 2,6-DNT. No suitable NOAEL or LOAEL was found for any duration category.

Oral MRLs.

Studies in humans did not provide sufficient data regarding exposure levels and their correlation with observed effects. Therefore, animal studies were used for the derivation of the oral MBLs for both 2,4- and 2,6-DNT.

2,4-DNT

- An acute-duration oral MRL of 0.05 mg/kg/day was derived for 2,4-DNT from a NOAEL value of 5 mg/kg/day for neurotoxicity observed in dogs (Ellis et al. 1985; Lee et al. 1978). Male and female beagle dogs were dosed by capsule with 0, 1.5, or 25 mg/kg/day 2,4-DNT in intermediate- and chronic-duration studies. However, after 12 days of treatment, the first clinical signs of neurotoxicity were evident. Minimal signs of neurotoxicity were incoordination and stiffness, giving the animals an abnormal gait. No clinical signs of neurotoxicity were observed at 5 mg/kg/day. The NOAEL of 5 mg/kg/day was divided by an uncertainty factor (UF) of 100 (10 for animal-to-human extrapolation, and 10 for human variability).
- A chronic-duration oral MRL of 0.002 mg/kg/day was derived for 2,4-DNT from a NOAEL value of 0.2 mg/kg in dogs (Ellis et al. 1979, 1985). Beagle dogs were administered 0.02, 1.5, or 10 mg/kg 2,4-DNT in capsules for up to 24 months. Methemoglobinemia and Heinz bodies were observed in dogs fed 1.5 mg/kg. Biliary hyperplasia and neurotoxicity (paralysis and cerebellar lesions) were also noted at this dose. No testicular degeneration was observed up to 10 mg/kg 2,4-DNT. The NOAEL of 0.2 mg/kg for hematological and neurological effects and biliary hyperplasia was divided by an uncertainty factor of 100 (10 for animal-to-human extrapolation and 10 for human variability).

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2,6-DNT

Data were insufficient for the derivation of acute-duration oral MRLs for 2,6-DNT. No suitable NOAEL or LOAEL was found.

- An intermediate-duration oral MRL of 0.004 mg/kg/day for 2,6-DNT was derived from a LOAEL value of 4 mg/kg, at which extramedullary erythropoiesis in the spleen and lymphoid depletion were observed in dogs (Lee et al. 1976). Beagle dogs were administered 0,4,20, or 100 mg/kg 2,6-DNT in capsules for up to 13 weeks. Treatment-related mortality occurred at 20 and 100 mg/kg 2,6-DNT. No neurological effects were found in the 4-mg/kg group, but at 20 mg/kg, listlessness, incoordination, and lack of balance were found; effects became more severe at 100 mg/kg and progressed to paralysis, occasional tremors, and inability to eat. Body weight loss correlated with food consumption at 20 and 100 mg/kg. Anemia and compensatory reticulocytosis were also found at 20 and 100 mg/kg. Other treatment-related effects observed at mid and/or high dose were thymic involution, bile duct hyperplasia, testicular degeneration, hepatic inflammation, and dilated renal tubules. None of these effects were observed in animals treated with 4 mg/kg. However, after 13 weeks, mild extramedullary erythropoiesis in the spleen and lymphoid depletion were observed at 4 mg/kg. The LOAEL of 4 mg/kg for hematological effects was divided by an uncertainty factor of 1,000 (10 for use of a LOAEL, 10 for animal-to-human extrapolation, and 10 for human variability).

Data were insufficient for the derivation of chronic-duration oral MBLs for 2,6-DNT. No suitable NOAEL or LOAEL was found.

Death.

The limited data available on inhalation exposure to 2,4-DNT or Tg-DNT suggest that there may be an increase in death due to ischemic heart disease and diseases of the circulatory system (Levine et al. 1986a). There was no increase found in death due to cancer in this study. Although no data are available regarding death in animals after inhalation exposure to 2,4- or 2,6-DNT, and only one study reported an increased incidence of death after occupational exposure to 2,4- or Tg-DNT, it appears unlikely that death would occur in people living near hazardous waste sites and exposed to low levels of 2,4- or 2,6-DNT. No data are available on death in humans from oral exposure to 2,4- or 2,6-DNT, but from animal data it can be reasonably expected that consumption of these compounds may be fatal. Insufficient data are available to predict whether dermal exposure to 2,4- or 2,6-DNT would cause death in humans.

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Systemic Effects.

Respiratory Effects. Because no studies are available in humans or animals examining the respiratory endpoints after inhalation or dermal exposure to 2,4- or 2,6-DNT, it is not possible to predict whether exposure to these compounds via these routes would cause respiratory effects in humans living near hazardous waste sites. Human data are not available on respiratory effects after oral exposure to 2,4- or 2,6-DNT, but no effects on the respiratory system were found when rats were fed Tg-DNT for up to 2 years (Hazleton Laboratories 1982).

Cardiovascular Effects. Excessive rates of mortality from ischemic heart disease and residual diseases of the circulatory system were observed in workers exposed to Tg-DNT and 2,4-DNT at two separate facilities (Levine et al. 1986a). However, this finding was unusual. The heart disease mortality risk of workers is commonly lower than that of the general population because of the “healthy worker” effect. It is unlikely that a chance aggregation of risk factors independent of DNT exposure could explain the observed excess of heart disease mortality.

Although treatment-related cardiovascular lesions have not been reported in rat or mouse bioassays, these species are generally resistant to naturally-occurring or experimentally-induced atherosclerosis (Jokinen et al. 1985). Interim sacrifices in a 2-year bioassay in rats did show an exacerbation of spontaneous myocarditis, although it was not seen at study termination (Hazleton Laboratories 1982). Thus, insufficient data are available to determine whether cardiovascular effects might result from living near hazardous waste sites.

Gastrointestinal Effects. Some gastrointestinal symptoms, such as nausea and vomiting, have been reported after workers were exposed to 2,4-DNT, presumably via inhalation and dermal exposure (McGee et al. 1942, 1947). No animal studies are available regarding gastrointestinal effects after inhalation exposure to 2,4- and 2,6-DNT. The only data available after oral exposure are from a study in which no histopathological effects on the gastrointestinal tract were observed in rats fed Tg-DNT for up to 2 years (Hazleton Laboratories 1982). Thus, insufficient data are available to predict whether low-level exposure to 2,4- or 2,6-DNT might result in gastrointestinal effects in persons living near hazardous waste sites.

Hematological Effects. Early observations of workers exposed to 2,4-DNT (levels of exposure not described) report symptoms of cyanosis (McGee et al. 1942, 1947; Perkins 1919). McGee et al. (1942, 1947) also reported anemia. Occupational conditions at the time of these reports (pre-1950) undoubtedly

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resulted in higher levels of human exposure than would occur in modern manufacturing facilities. Engineering, ventilation, and industrial hygiene improvements have reduced the levels of modern occupational exposure to DNT. Unfortunately, recent studies of exposed workers (Ahrenholz 1980; Ahrenholz and Meyer 1982; Hamill et al. 1982; Levine et al. 1985a) did not monitor hematological profiles.

In animal studies of intermediate duration, methemoglobinemia and its sequelae (Heinz bodies, anemia, reticulocytosis), hemosiderosis, extramedullary hematopoiesis, and slight cyanosis (Lane et al. 1985) were observed when 2,4-, 2,6-, and Tg-DNT were administered (Hazleton Laboratories 1977, 1982; Lee et al. 1976, 1978). Extramedullary erythropoiesis in the spleen and lymphoid depletion have been observed after intermediate-duration treatment of dogs with 2,6-DNT (Lee et al. 1976). In animal studies of chronic duration, anemia was observed (Ellis et al. 1979; Hazleton Laboratories 1982), but the animals often appeared to adapt, as indicated by a decrease in the hematological effects in the second year of exposure. Based on these findings in humans and animals, it appears that hematological effects may occur in persons exposed to 2,4-DNT at low levels near hazardous waste sites.

Musculoskeletal Effects. Workers at munitions plants that produced 2,4- or Tg-DNT have complained of muscle weakness, joint pain, and other incapacitating symptoms (McGee et al. 1942; Perkins 1919). Because these workers were most likely exposed to high levels of DNT without protective equipment, these effects may not necessarily be observed in persons exposed to low levels. No human or animal data are available on musculoskeletal effects of 2,4-, 2,6-, or Tg-DNT after oral exposure.

Hepatic Effects. Medical surveys of workers exposed to Tg-DNT performed during the 1970s and 1980s revealed no significant differences in hepatic blood chemistry profiles (Ahrenholz 1980; Ahrenholz and Meyer 1982). In earlier surveys of workers exposed presumably to 2,4-DNT, 2 of 154 workers (McGee et al. 1942) and 29 of 714 workers (McGee et al. 1947) indicated symptoms of liver tenderness. These reports are probably not significant. The incidence of liver tenderness reported by McGee et al. (1947) was increased over earlier reports by McGee et al. (1942), despite improvements in engineering, ventilation, and industrial hygiene in the intervening period that were likely to decrease the magnitude of worker exposures to DNT. In addition, alcohol consumption, which may also have been a cause of liver tenderness, was not taken into account.

A consistent finding in several studies of rats, mice, and dogs fed high doses of 2,4-, 2,6-, and Tg-DNT has been the occurrence of adverse effects on the liver or biliary tract (Ellis et al. 1979; Hazleton Laboratories

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1982; Hong et al. 1985; Lee et al. 1976, 1978, 1985). Lesions in rats were dose-dependent and may have progressed from foci of altered hepatocytes to proliferative nodules to hepatocellular carcinoma (Ellis et al. 1979; Hazleton Laboratories 1982; Leonard et al. 1987).

The significance of hepatotoxicity observed in animal studies to potential adverse liver effects in humans exposed to DNT isomers is uncertain. The negative findings in humans cannot be considered conclusive. Studies of DNT exposure are limited by the small groups of workers studied and by the lack of individual exposure monitoring. The consistent observation of hepatotoxicity in animals indicates that a potential exists for hepatotoxicity in humans.

Endocrine Effects. Although no histopathological effects on adrenal, pituitary, or thyroid glands have been observed in several bioassays, after oral exposure to 2,4- or Tg-DNT (Hazleton Laboratories 1985; McGown et al. 1983), an increase in the incidence and severity of parathyroid hyperplasia, and fatty metamorphoses and vascular ectasia were found in rats fed Tg-DNT in a chronic study (Hazleton Laboratories 1982). No data are available on endocrine effects in humans after exposure to 2,4-, 2,6-, or Tg-DNT. Therefore, the likelihood that exposure to 2,4- or 2,6-DNT will cause deleterious effects on the endocrine system in people cannot be determined.

Renal Effects. Medical surveys of workers exposed to Tg-DNT revealed no significant differences in renal blood chemistry profiles (Ahrenholz 1980; Ahrenholz and Meyer 1982). The limitations of these studies were discussed previously. Adverse effects in the kidney have been observed in laboratory animals exposed to 2,4- and 2,6-DNT, but the observations have not been as consistent as reports of hematological, reproductive, and hepatic effects in exposed animals.

Renal dysplasia was observed in male, but not female, mice exposed chronically (up to 2 years) to 2,4-DNT (Ellis et al. 1979). Cystic degeneration, atypical epithelium lining the cysts, and a variety of tumors were reported by Ellis et al. (1979). Subchronic administration of 2,6-DNT to dogs, but not mice or rats, resulted in less severe renal toxicity that included kidney degeneration (Lee et al. 1976).

The potential for renal toxicity in humans exposed to DNT is uncertain; 2,4-DNT has elicited severe renal effects only in the male mouse. Renal effects were observed in dogs administered 2,6-DNT but not when 2,4-DNT was administered.

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Dermal Effects. Dermatitis was attributed to dermal exposure to 2,4-DNT in a small percentage of munitions workers exposed topically to DNT (McGee et al. 1942, 1947). Both 2,4- and 2,6-DNT are mildly irritating to rabbit skin (Ellis et al. 1978; Lee et al. 1975). Mild sensitization has been observed in guinea pigs after 2,6-DNT exposure but was not observed after exposure to 2,4-DNT (Ellis et al. 1978; Lee et al. 1975). Therefore, although limited data are available, it appears that direct contact with low levels of 2,4- or 2,6-DNT may cause dermatitis and/or dermal sensitization in humans; however, insufficient data are available to state this with certainty.

Ocular Effects. Only animal data are available on ocular effects after exposure to 2,4- or 2,6-DNT. Histopathological examination of the eyes of rats fed Tg-DNT for up to 2 years showed no abnormalities (Hazleton Laboratories 1982). Neither 2,4- nor 2,6-DNT was found to cause eye irritation in rabbits (Ellis et al. 1978; Lee et al. 1975). There was no mention of ocular effects in studies of munitions workers exposed to 2,4- or Tg-DNT (McGee 1942, 1947; Perkins 1919). Therefore, it appears unlikely that low-level exposure to 2,4- or 2,6-DNT would cause ocular effects in people living near hazardous waste sites.

Body Weight Effects. Effects on body weight have not been reported in munitions workers (McGee et al. 1942, 1947; Perkins 1919), but numerous studies have shown body weight loss or decreased body weight gain in rats, mice, and dogs after oral treatment with 2,4-, 2,6-, or Tg-DNT (Bloch et al. 1988; Ellis et al. 1979, 1985; Hazleton Laboratories 1982; Hong et al. 1985; Kozuka et al. 1979; Lane et al. 1985; Lee et al. 1978, 1985; Leonard et al. 1987; NCI 1978). However, in most of these studies there was a concurrent decrease in food consumption. Therefore, it is not known whether exposure to low levels of 2,4- or 2,6-DNT found near hazardous waste sites would cause deleterious effects on body weight in humans.

Immunological and Lymphoreticular Effects. Although no data are available regarding immunological or lymphoreticular effects in humans, some data on these end points are available in animals. No effects on serum concentrations of IgE, the antibody associated with allergic or hypersensitive reactions, were reported in rats or dogs exposed to 2,4- or 2,6-DNT (Ellis et al. 1985; Lee et al. 1976, 1978, 1985). Mild sensitization was observed in guinea pigs after dermal exposure to 2,6-DNT, but not after exposure to 2,4-DNT (Ellis et al. 1978; Lee et al. 1975). Therefore, it may be expected that human sensitizing potential to 2,4-DNT would be low but may occur after exposure to 2,6-DNT. Lymphoreticular effects have been reported in rats, mice, and dogs administered 2,6-DNT (Lee et al. 1976) and rats administered Tg-DNT (Hazleton Laboratories 1977, 1982). Splenic effects included alterations in appearance such as discoloration, enlargement, and surface irregularities; thymic involution was also seen in dogs (Hazleton Laboratories 1977,

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1982; Lee et al. 1976). Based on these data in several species, it is possible that immunological or lymphoreticular effects may occur in people living near hazardous waste sites exposed to 2,4- or 2,6-DNT, but insufficient data are available to state this with certainty.

Neurological Effects. Early reports of workers occupationally exposed presumably to 2,4-DNT refer to symptoms indicative of neurotoxicity in humans. Perkins (1919) reported headaches, sleepiness, pain in the joints, and dizziness in exposed workers. McGee et al. (1942) reported that occupationally exposed workers complained of nausea, insomnia, headaches, dizziness, and tingling pains in the extremities. More recent occupational studies (Ahrenholz 1980; Ahrenholz and Meyer 1982; Hamill et al. 1982; Levine et al. 1985a) failed to examine workers for symptoms of neurotoxicity.

In animals, the nervous system has been observed to be a major target of 2,4- and 2,6-DNT toxicity (Ellis et al. 1979, 1985; Kozuka et al. 1979; Lee et al. 1978, 1985). Clinical signs have been most pronounced in dogs, and less severe symptoms have been observed in rats and mice (Ellis et al. 1979; Lee et al. 1976, 1978). Initial symptoms in dogs included incoordination and stiffness, especially of the hind legs, resulting in an abnormal gait. More seriously affected dogs had paralysis of the hind legs which progressed to the forelimbs and eventually the neck. CNS lesions, including cerebellar vacuolization, hypertrophy and focal gliosis, and some cerebellar and brain stem perivascular hemorrhage were seen in dogs fed 2,4-DNT (Ellis et al. 1979, 1985). In the mouse, there were signs of depression and hyperexcitability. Neurotoxic symptoms in rats administered 2,4-DNT were less common, but some rats administered 2,6-DNT had neuromuscular symptoms of incoordination and abnormal gait.

There are no data on the biochemical events involved in the toxicity of the nervous system. Based on the available human and animal data, it appears that neurological effects may occur in persons exposed to low levels of 2,4- or 2,6-DNT.

Reproductive Effects. A consistent finding in several studies in rats, mice, and dogs dosed with 2,4- or 2,6-DNT has been the impairment of the male reproductive system. Testicular atrophy, degeneration of the seminal vesicles, and decreased sperm production have been observed in the test animals (Bloch et al. 1988; Ellis et al. 1979; Lee et al. 1976, 1978).

Sertoli cells play a major role in maintaining spermatogenesis in the testis. Bloch et al. (1988) reported that fine structural alterations of Sertoli cells accompanied the diminished sperm count in rats acutely exposed to

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2,4-DNT. Other deleterious effects included frayed basal lamina and distortion of peritubular tissue. Sertoli cell cultures prepared from testes of Wistar rats and treated with up to 100 pM 2,4- or 2,6-DNT remained intact, although at 50 FM of 2,4- or 2,6-DNT, some damage was observed as many of the germ cells were absent and some Sertoli cells contained cytoplasmic vacuoles (Reader and Foster 1990). Germ cell detachment from Sertoli germ cell cocultures was significantly increased ($p<0.05$) compared to controls at 10 FM 2,4- or 2,6-DNT (Reader and Foster 1990). Further evidence of disruption of Sertoli cell function was observed as increased production of lactate and pyruvate, although increases in pyruvate production were minimal with 2,6-DNT (Reader and Foster 1990). Thus, it appears that structural changes in the Sertoli cells may be precipitating events responsible for spermatogenic disruption noted in 2,4-DNT exposed rats. Bloch et al. (1988) also reported increased serum levels of FSH. Increased FSH levels have been associated with Sertoli cell malfunction.

In rodent studies measuring the fertility of test animals dosed with 2,4-DNT, there have been marked and significant decreases in the number of fetal implants, which have been attributed to the adverse impacts of 2,4-DNT on sperm production (Ellis et al. 1979; Lane et al. 1985; Lee et al. 1978).

Based on the adverse effects on sperm production and fertility seen in oral studies in animals, assessments of the reproductive effects of 2,4- and 2,6-DNT on occupationally exposed workers have been conducted. As described previously, inhalation is assumed to be the main route of exposure, with probable concurrent dermal and oral exposure. Three of these studies, Hamill et al. (1982), Ahrenholz and Meyer (1982), and Levine et al. (1985a), reported no detectable differences in sperm levels or fertility rates as a result of occupational exposure.

In an earlier, study Ahrenholz (1980) reported a significant reduction in the sperm counts of exposed male workers as well as an increase of marginal statistical significance in the number of spontaneous abortions in the wives of exposed workers.

The small exposure populations and lack of historical individual exposure monitoring limit the effectiveness of the occupational studies to detect adverse effects on reproduction. Although the findings reported by Ahrenholz (1980) are suggestive of a reproductive problem in exposed workers (especially with regard to the findings of reproductive toxicity in animal studies), the human results cannot be considered conclusive, however, neither can they be dismissed as insignificant. Therefore, insufficient data are available to

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determine whether people exposed to low levels of 2,4- or 2,6-DNT near hazardous waste sites may have reproductive effects.

Developmental Effects. No defects were found in rat pups after dams were fed 2,4-DNT in the diet during gestation (Ellis 1979). Decreased pup viability in this study was believed to result from a high incidence of maternal death during parturition and maternal neglect.

Tg-DNT was orally administered to pregnant rats during gestation and adverse effects upon blood elements and organ weights were observed in the dams and the fetuses (Jones-Price et al. 1982). The findings of fetal hematotoxicity imply that oxygen supply to developing tissues may be impaired by exposure of mothers to DNT. Exposure to any substance that depletes the amount of oxygen available to developing fetal tissues can have adverse consequences. However, insufficient data are available to predict whether low-level exposure to 2,4- or 2,6-DNT may cause developmental effects in humans.

Genotoxic Effects. Some results of *in vitro* genotoxicity assays are presented in Tables 2-3 (2,4-DNT), 2-4 (2,6-DNT), and 2-5 (Tg-DNT). Results of *in vivo* genotoxicity assays are presented in Table 2-6. DNT causes gene mutations in the reverse mutation assay using *S. typhimurium*. However, the test system has given variable results because of the need for metabolic activation and the sensitivity of the tester strains.

Unscheduled DNA synthesis (UDS) and S-phase synthesis (SPS) were induced *in vitro* in the hepatocytes of Fischer-344 rats treated with 2,4-DNT *in vivo* (Mirsalis et al. 1989). The genotoxicity of Tg-DNT is believed to be due to the potent genotoxicity of the 2,6-DNT component, as was evidenced in an *in vivo-in vitro* hepatocyte UDS system (Mirsalis and Butterworth 1982). The mutagenicity of several of the metabolites of 2,6-DNT have been tested in *S. typhimurium*. Although neither 2,6-DNT nor its metabolites 2-amino-6-nitrotoluene, 2,6-dinitrobenzylalcohol, 2-acetylaminio-6-nitrobenzoic acid, and 2-amino-6-nitrobenzoic acid were mutagenic in this assay with or without S9 activation, other metabolites of 2,6-DNT were found to possess mutagenic activity (Sayama et al. 1989b). The putative metabolite 2,6-dinitrobenzaldehyde was a direct acting mutagen, that is, it did not require activation (Sayama et al. 1989b). Urine from Fischer-344 rats administered 75 mg/kg 2,6-DNT by gavage tested positive for mutagenicity using *S. typhimurium* TA 98 without S9 activation (Chadwick et al. 1993).

2,4-DNT induced lethal mutations but not reciprocal translocations in mutagenicity testing using *Drosophila melanogaster* (Woodruff et al. 1985).

TABLE 2-3. Genotoxicity of 2,4-Dinitrotoluene *In Vitro*

Species (test system)	End point	Results		Reference
		With activation	Without activation	
<i>Salmonella typhimurium</i>	Reverse mutation	-	+	Couch et al. 1981
TM 677	Forward mutation	+	+	Couch et al. 1981
<i>S. typhimurium</i>	Reverse mutation	NT	-	Chiu et al. 1978
<i>S. typhimurium</i> (TA 98)	Reverse mutation	-	-	Dellarco and Prival 1989
<i>S. typhimurium</i> (TA 100)	Reverse mutation	-	-	Dellarco and Prival 1989
<i>S. typhimurium</i>	Reverse mutation	+	+	Tokiwa et al. 1981
<i>S. typhimurium</i> with flavin mononucleotide (TA 98)	Reverse mutation	+	-	Dellarco and Prival 1989
<i>S. typhimurium</i> with flavin mononucleotide (TA 100)	Reverse mutation	-	-	Dellarco and Prival 1989
<i>S. typhimurium</i>	Reverse mutation	NT	+	Mori et al. 1982
<i>S. typhimurium</i>	Reverse mutation	+	+	Spanggord et al. 1982b
<i>S. typhimurium</i> (TA 100)	Base-pair substitution	+	+	Ellis et al. 1978
<i>S. typhimurium</i> (TA 1535)	Base-pair substitution	-	-	Ellis et al. 1978
<i>S. typhimurium</i> (TA 98, 1537)	Reverse mutation	-	-	Ellis et al. 1978
<i>S. typhimurium</i> (TA 1538)	Reverse mutation	+	-	Ellis et al. 1978

TABLE 2-3. Genotoxicity of 2,4-Dinitrotoluene *In Vitro* (continued)

Species (test system)	End point	Results		Reference
		With activation	Without activation	
<i>S. typhimurium</i>	Reverse mutation	+	NT	Pearson et al. 1979
<i>S. typhimurium</i> (TA 98)	Reverse mutation	NT	+	Einiö et al. 1991
(TA 98 NR)		NT	+	
(TA 98/1,8-DNP ₆)		NT	+	
(YG 1021)		NT	+	
(YG 1024)		NT	+	
<i>E. coli</i>	Reverse mutation	-	-	Dunkel et al. 1985
Chinese hamster ovary cells (CHO)	Sister chromatid exchange	+	-	Loveday et al. 1989
	Chromosomal aberrations	-	-	Loveday et al. 1989
CHO/HEPRT	Forward mutation	-	-	Abernethy and Couch 1982
P388 mouse lymphoma TK	Forward mutation	-	+	Styles and Cross 1983
Syrian hamster embryo cells	Morphological transformation	NT	-	Holen et al. 1990

+ = positive result; - = negative result; NT = not tested

TABLE 2-4. Genotoxicity of 2,6-Dinitrotoluene *In Vitro*

Species (test system)	End point	Results		Reference
		With activation	Without activation	
<i>Salmonella typhimurium</i>	Reverse mutation	+	+	Couch et al. 1981
TM 677	Forward mutation	+	+	Couch et al. 1981
<i>S. typhimurium</i>				
TA 100, TA 1535	Base-pair substitution	-	-	Ellis et al. 1978
TA 98, 1537	Reverse mutation	-	-	
TA 1538	Reverse mutation	-	+	
<i>S. typhimurium</i>	Reverse mutation	NT	+	Simmon et al. 1977
<i>S. typhimurium</i>				
TA 98, TA 100	Reverse mutation	-	-	Sayama et al. 1989
<i>S. typhimurium</i>	Reverse mutation	-	+	Tokiwa et al. 1981
<i>S. typhimurium</i>	Reverse mutation	+	+	Spanggord et al. 1982b
CHO/HEPRT	Forward mutation	-	-	Abernethy and Couch 1982
P388 mouse lymphoma TK	Forward mutation	-	-	Styles and Cross 1983
Syrian hamster embryo cells	Morphological transformation	NT	-	Holen et al. 1990

TABLE 2-4. Genotoxicity of Trichloroethylene *In Vitro* (continued)

Species (test system)	End point	Results		Reference
		With activation	Without activation	
<i>S. typhimurium</i> (TA 98)	Reverse mutation	NT	+	Einiö et al. 1991
		NT	-	
		NT	+	
		NT	+	
		NT	+	
<i>S. typhimurium</i>	Reverse mutation	+	NT	Pearson et al.

+ = positive result; - = negative result; NT = not tested

TABLE 2-5. Genotoxicity of Technical-Grade DNT

Species (test system)	End point	Results		Reference
		With activation	Without activation	
<i>Salmonella typhimurium</i>	Reverse mutation	+	+	Couch et al. 1981
<i>S. typhimurium</i>	Reverse mutation ^a	NT	+	Chadwick et al. 1990
TM 677	Forward mutation	+	+	Couch et al. 1981
CHO/HEPRT	Forward mutation	-	-	Abernethy and Couch 1982
P388 mouse lymphoma TK	Forward mutation	-	-	Styles and Cross 1983

^aUrine from CD-1 mice or Fischer-344 rats that had been treated *in vivo* with DNT was used.

+ = positive result; - = negative result; NT = not tested

TABLE 2-6. Genotoxicity of 2,4-Dinitrotoluene *In Vivo*

Species (test system)	End point	Results	Reference
Rat hepatocyte	UDS	+	Mirsalis et al. 1989
Rat hepatocyte	SPS	+	Mirsalis et al. 1989
Human lymphocyte	Chromosomal aberration	+	Huang et al. 1995
Mouse bone marrow	Micronucleus	-	Ashby et al. 1985
Rat hepatocyte	UDS	+	Ashby et al. 1985
Rat hepatocyte	UDS	+	Mirsalis and Butterworth 1982
Rat hepatocyte	DNA binding	+	La and Froines 1993
Rat hepatocyte	DNA binding	+	Chadwick et al. 1993
Human lymphocyte	Chromosomal aberration	+	Huang et al. 1995
Rat hepatocyte	UDS	+	Ashby et al. 1985

+ = positive result; - = negative result; DNA = deoxyribonucleic acid; SPS = S-phase synthesis; UDS = unscheduled DNA synthesis

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The formation of DNA adducts is generally thought to indicate carcinogenic risk (La and Froines 1993). Both 2,4- and 2,6-DNT have induced DNA adducts in rat liver. Following treatment with 2,4-DNT, three DNA adducts were found in the liver of Fischer-344 rats (La and Froines 1992). Four DNA adducts, which were not identified, were found in the liver of rats treated with 75 mg/kg 2,6-DNT by gavage (Chadwick et al. 1993). The formation of 4 DNA adducts was also observed after intraperitoneal administration of 219 mg/kg 2,6-DNT to Fischer-344 rats in a study by La and Froines (1992, 1993). One adduct accounts for the majority of the radioactivity measured; about 85% of the total was 1 adduct in the study using 2,4-DNT, while in the study with 2,6-DNT, about 60% of the total adducts measured were from a single adduct with the other adducts constituting 10-15% of the total (La and Froines 1992, 1993). No quantitative or qualitative differences in adduct formation were found when treatment occurred by gavage or intraperitoneal injection (La and Froines 1992). The proximate DNA binding species has been postulated to be 2-hydroxylamino-6-nitrobenzyl alcohol (La and Froines 1993; Rickert et al. 1984). The DNA adducts formed after exposure to 2,4- or 2,6-DNT were persistent over time; the persistence of these adducts was slightly more than 40% in the 2 weeks after exposure (La and Froines 1992).

Cancer. 2,4-DNT administered in the diet of mice caused cancer of the kidneys (Ellis et al. 1979); 2,4-, 2,6-, and Tg-DNT caused hepatocellular carcinoma in rats (Ellis et al. 1979; Hazleton Laboratories 1982; Leonard et al. 1987). 2,4-DNT was shown to be a tumor promoter, but not a tumor initiator, using *in vivo* hepatic initiation-promotion protocols (Leonard et al. 1983, 1986; Mirsalis and Butter-worth 1982). 2,6-DNT and Tg-DNT were shown to be complete carcinogens using the same protocols (Leonard et al. 1983, 1986). The results of Leonard et al. (1983, 1986, 1987) and Mirsalis and Butterworth (1982) suggest or show that 2,6-DNT is the isomer most responsible for the carcinogenic response observed in animal studies with Tg-DNT. No increase in morphological transformation was observed in Syrian hamster embryo cells (SHE) compared to controls when SHE cells were exposed to 2,4-, 2,6-, or Tg-DNT, or to the DNT metabolites 2,4-diaminotoluene, 2-amino-4-nitrotoluene, 2-amino-6-nitrotoluene, or 2,4-dinitrobenzoic acid (Holen et al. 1990). In addition, none of these isomers had initiator-like or promotor-like effects in this assay when tested with *O*-tetradecanoyl phorbol 13-acetate or benzo[a]pyrene, respectively.

In order to evaluate the possibility that DNT might also cause cancer in humans, Levine et al. (1986b) performed a retrospective cohort mortality study at two army ammunition plants that used Tg-DNT and/or 2,4-DNT. No significant increases in mortality from malignant neoplasms as a whole or from particular cancers (liver, lung, gallbladder, kidney, and connective tissues) were observed. The study was limited by small cohort size, and could have detected only an 8-fold or greater increase in liver or gallbladder cancer. As

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a result, the negative findings of Levine et al. (1986b) do not refute the animal cancer findings with regard to the potential for cancer in humans. The cancer classification for 2,4- and 2,6-DNT mixture is B2: probable human carcinogen (EPA 1992). An upperbound q_1^* for oral exposure to 2,4- and 2,6-DNT mixture was estimated by EPA (1986d, 1998) to be $0.68 \text{ (mg/kg/day)}^{-1}$ based on the combined tumor incidence of liver and mammary gland tumors in rats. Doses associated with excess cancer risks of 10^{-4} , 10^{-5} , 10^{-6} , and 10^{-7} by the oral routes are plotted in Figure 2-1.

2.6 CHILDREN'S SUSCEPTIBILITY

This section discusses potential health effects from exposures during the period from conception to maturity at 18 years of age in humans, when all biological systems will have fully developed. Potential effects on offspring resulting from exposures of parental germ cells are considered, as well as any indirect effects on the fetus and neonate due to maternal exposure during gestation and lactation. Relevant animal and in vitro models are also discussed.

Children are not small adults. They differ from adults in their exposures and may differ in their susceptibility to hazardous chemicals. Children's unique physiology and behavior can influence the extent of their exposure. Exposures of children are discussed in section 5.6 Exposures of Children.

Children sometimes differ from adults in their susceptibility to hazardous chemicals, but whether there is a difference depends on the chemical (Guzelian et al 1992; NRC 1993). Children may be more or less susceptible than adults to health effects and the relationship may change with developmental age (Guzelian et al 1992; NRC 1993). Vulnerability often depends on developmental stage. There are critical periods of structural and functional development during both pre-natal and post-natal life and a particular structure or function will be most sensitive to disruption during its critical period(s). Damage may not be evident until a later stage of development. There are often differences in pharmacokinetics and metabolism between children and adults. For example, absorption may be different in neonates because of the immaturity of their gastrointestinal tract and their larger skin surface area in proportion to body weight (Morselh et al 1980; NRC 1993); the gastrointestinal absorption of lead is greatest in infants and young children (Ziegler et al 1978). Distribution of xenobiotics may be different; for example, infants have a larger proportion of their bodies as extracellular water and their brains and livers are proportionately larger (Widdowson and Dickerson 1964; Foman et al 1982; Owen and Brozek 1966; Altman and Dittmer 1974; Foman 1966). The infant also has an immature blood-brain barrier (Adinolfi 1985; Johanson 1980) and probably an immature blood-testis

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barrier (Setchell, BP and Waites GMH 1975). Many xenobiotic metabolizing enzymes have distinctive developmental patterns and at various stages of growth and development, levels of particular enzymes may be higher or lower than those of adults and sometimes unique enzymes may exist at particular developmental stages (Leeder and Keams 1997; Komori 1990; Vieira et al 1996; NRC 1993). Whether differences in xenobiotic metabolism make the child more or less susceptible also depend on whether the relevant enzymes are involved in activation of the parent compound to its toxic form or in detoxification. There may also be differences in excretion, particularly in the newborn who has a low glomerular filtration rate and has not developed efficient tubular secretion and resorption capacities (Altman and Dittmer 1974; West et al 1948; NRC 1993). Children and adults may differ in their capacity to repair damage from chemical insults. Children also have a longer lifetime in which to express damage from chemicals; this potential is particularly relevant to cancer.

Certain characteristics of the developing human may increase exposure or susceptibility while others may decrease susceptibility to the same chemical. For example, the fact that infants breathe more air per kilogram of body weight than adults may be somewhat counterbalanced by their alveoli being less developed, so there is a disproportionately smaller surface area for absorption (NRC 1993).

No specific health effects resulting from DNT exposure have been observed in children. Generally, health effects observed in adults should also be of potential concern in children.

No direct information is available regarding the effects of DNT on the developmental process in humans and there are few developmental studies on animals. When Tg-DNT was administered by gavage to pregnant rats for 14 days during gestation, and pups were evaluated for developmental toxicity either at gestation day 20 or postpartum day 60 (Jones-Price et al. 1982), adverse effects on hematologic parameters and altered organ weights were observed in both dams and fetuses when dams were administered 100 or 150 mg/kg/day. However, the fetal toxicity was not dose-related. A decrease in relative liver weight was observed in the postpartum pups at the low dose of 14 mg/kg/day. Dose-related effects on postnatal development were not observed in pups when dams were administered 35 or 75 mg/kg/day.

No consistent changes were observed in the number of preimplantation losses, implantation sites, or living or non-living fetuses in male Sprague-Dawley rats gavaged with 2,4 DNT at 0, 60, 180, or 240 mg/kg/day for 5 days (Lane et al. 1985). Exposure of male animals to DNT does not cause dominant lethal mutation or increases in the proportion of nonviable conception (Ellis et al. 1979).

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DNT has been found to be genotoxic using *in vivo* test systems (Ashby et al. 1985; Huang et al. 1995; Mirsalis et al. 1989). Although 2,6-DNT itself showed no mutagenicity towards *Salmonella typhimurium* strains TA98 and TA100 with or without activation by S9 mix, 2,6-dinitrobenzaldehyde, a metabolite of 2,6-DNT, was found to be a direct-acting mutagen, not requiring metabolic activation (Sayama et al. 1989). The reason that DNT is not shown to bind to DNA and cause mutations in most of the short-term *in vitro* assays for genotoxicity is that the formation of DNA-reactive DNT metabolites involved several different biotransforming enzymes in the intestinal microflora and in the liver. However, DNT did not cause dominant lethal mutation or increases in the proportion of nonviable conceptions following exposure of male animals (Ellis et al. 1979), so it is not clear if the genotoxic form of DNT might potentially reach the germ cells following oral, inhalation, or dermal exposure.

It is unlikely that DNT and its metabolites will accumulate in maternal tissues because of its low octanol/water partition coefficient. No studies are available that demonstrate DNT or its metabolites cross the placenta or get into breast milk. Thus, it is unlikely that the developing fetus or nursing infant would be exposed to DNT as a consequence of maternal exposure prior to gestation. However, developmental toxicity from DNT could potentially occur because of its ability to deplete the amount of oxygen available to the developing fetus. Pregnant women and their fetuses may be susceptible to the oxygen depletion implied by the hemotoxicity of DNT based on a study of rats (Jones-Price et al. 1982). Newborns have a transient deficiency in methemoglobin reductase which reduces methemoglobin back to hemoglobin (Gruener 1976). They also have a high concentration of fetal hemoglobin in their erythrocytes (Smith 1996). Thus, newborns are unusually sensitive to methemoglobin-generating chemicals such as DNT. The metabolism of DNT has not been studied in children or appropriate animal models. However, while some of the enzymes involved in DNT metabolism reach or exceed adult levels during infancy and early childhood, other enzymes such as UDP-glucuronosyltransferase may attain adult levels by 6-18 months of age (Leeder and Keams 1997). Thus, the toxicity of DNT may be different in children.

2.7 BIOMARKERS OF EXPOSURE AND EFFECT

Biomarkers are broadly defined as indicators signaling events in biologic systems or samples. They have been classified as markers of exposure, markers of effect, and markers of susceptibility (NAS/NFX 1989).

Due to a nascent understanding of the use and interpretation of biomarkers, implementation of biomarkers as tools of exposure in the general population is very limited. A biomarker of exposure is a xenobiotic

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substance or its metabolite(s), or the product of an interaction between a xenobiotic agent and some target molecule(s) or cell(s) that is measured within a compartment of an organism (NAS/NRC 1989). The preferred biomarkers of exposure are generally the substance itself or substance-specific metabolites in readily obtainable body fluid(s) or excreta. However, several factors can confound the use and interpretation of biomarkers of exposure. The body burden of a substance may be the result of exposures from more than one source. The substance being measured may be a metabolite of another xenobiotic substance (e.g., high urinary levels of phenol can result from exposure to several different aromatic compounds). Depending on the properties of the substance (e.g., biologic half-life) and environmental conditions (e.g., duration and route of exposure), the substance and all of its metabolites may have left the body by the time samples can be taken. It may be difficult to identify individuals exposed to hazardous substances that are commonly found in body tissues and fluids (e.g., essential mineral nutrients such as copper, zinc, and selenium). Biomarkers of exposure to 2,4- and 2,6-dinitrotoluene are discussed in Section 2.7.1.

Biomarkers of effect are defined as any measurable biochemical, physiologic, or other alteration within an organism that, depending on magnitude, can be recognized as an established or potential health impairment or disease (NAS/NRC 1989). This definition encompasses biochemical or cellular signals of tissue dysfunction (e.g., increased liver enzyme activity or pathologic changes in female genital epithelial cells), as well as physiologic signs of dysfunction such as increased blood pressure or decreased lung capacity. Note that these markers are not often substance specific. They also may not be directly adverse, but can indicate potential health impairment (e.g., DNA adducts). Biomarkers of effects caused by 2,4- and 2,6-dinitrotoluene are discussed in Section 2.7.2.

A biomarker of susceptibility is an indicator of an inherent or acquired limitation of an organism's ability to respond to the challenge of exposure to a specific xenobiotic substance. It can be an intrinsic genetic or other characteristic or a preexisting disease that results in an increase in absorbed dose, a decrease in the biologically effective dose, or a target tissue response. If biomarkers of susceptibility exist, they are discussed in Section 2.9, Populations That Are Unusually Susceptible.

There are no biomarkers of exposure effects that have been validated in children or in adults exposed as children.

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2.7.1 Biomarkers Used to Identify or Quantify Exposure to 2,4- and 2,6-Dinitrotoluene

Spectrophotometric analysis of complexes of primary arylamines in urine, resulting from the reduction of DNT and its metabolites, has been used to biomonitor DNT workers (Smith et al. 1995). The method is specific for primary arylamines and eliminates interferences from other classes of amines that might be present in the urine. Metabolic intermediates and conjugates of DNT can also be detected if they are present as primary arylamines. Although this method cannot identify or quantify individual metabolites, it is simple and accurate and yields results in about 1 hour.

Workers exposed to DNT in a manufacturing plant excreted 2,4-DNT, 2,6-DNT, and their metabolites in the urine (Levine et al. 1985b). The concentrations of DNT in air ranged from 0.1 to 5.9 mg/m³. Concentrations of DNT and metabolites ranged from 1.68 to 16.74 mg/day (or 1.74-17.31 mg/L, based on an average daily urine volume of 967 mL), with widespread daily variations. Estimates of inhaled DNT ranged from 0.5 to 4.9 mg/day, less than the total excreted. These results indicate that dermal and oral exposure contributed to the body burden of DNT.

Woollen et al. (1985) determined that the urinary concentration of a Tg-DNT metabolite, 2,4-dinitrobenzoic acid, was less than 1 mg/L at the beginning of the work week and ranged from 3.4 to 41 mg/L at the end of the shift. Atmospheric DNT levels of undetectable to 0.03 mg/m³ were monitored with personal air samples. Static samples near dusty process areas monitored were 0.02-2.68 mg/m³. The study authors estimate that inhalation exposures ranged from 1 to 14 mg/day. As in Levine et al. (1985b), inhalation exposure does not account for the entire amount of DNT metabolites excreted in urine. Dermal and ingestion exposures are therefore, likely to have occurred.

2.7.2 Biomarkers Used to Characterize Effects Caused by 2,4- and 2,6-Dinitrotoluene

For more information on biomarkers for renal and hepatic effects of chemicals see ATSDRDC Subcommittee Report on Biological Indicators of Organ Damage (1990) and for information on biomarkers for neurological effects see OTA (1990).

After exposure to DNT, methemoglobin levels in the blood may be elevated (Elleuhorn 1997). The methemoglobinemia present may be quite profound and its onset is often delayed by up to 4 hours (Ellenhorn

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1997). Another hematological change that might be present in individuals who have undergone repeated or prolonged exposure to DNT is that which is consistent with Heinz bodies and hemolytic anemia.

DNA adducts have been found in the livers of rats treated orally with either 2,4- or 2,6-DNT (La and Froines 1992, 1993). The formation of DNA adducts is believed to be indicative of carcinogenic risk.

Decreased spermatogenesis has been reported in treated rats, mice, and dogs (Block et al. 1988; Ellis et al. 1979). However, decrease in sperm counts in workers exposed to DNT has been reported in only one study (CDC 1981).

2.8 INTERACTIONS WITH OTHER CHEMICALS

Reduced tolerance to alcohol was observed in some workers presumably exposed to 2,4-DNT and other workers reported that alcohol ingestion intensified the symptoms of 2,4-DNT exposure (McGee et al. 1942). Perkins (1919) reported that "alcoholic subjects have very little resistance to DNT."

Exposure of male rats to 2,6-DNT for 5 days reduced the rate of metabolism of phenobarbital; exposure to 2,6-DNT for 4 weeks increased phenobarbital metabolism (Short and Lee 1980). Exposure of rats to 2,4-DNT did not affect the rate of phenobarbital metabolism.

The effects of the herbicide 2,4,5-trichlorophenoxyacetic acid (2,4,5-T) on 2,6-DNT genotoxicity were examined in male weanling Fischer-344 rats (George et al. 1992). The rats were treated orally with 54.4 mg/kg 2,4,5-T for 4 weeks, then with 75 mg/kg 2,6-DNT, 1,2, or 4 weeks after the first dose of 2,4,5-T; urine was then collected for 24 hours. In animals treated for 1 week with 2,4,5-T, there was a decrease in transformation of 2,6-DNT to mutagenic metabolites in the urine, but there were no changes in intestinal enzyme activities (George et al. 1992). Longer treatments with 2,4,5-T did not alter urine genotoxicity compared to controls, and there was a transient increase in cecal azo reductase and nitroreductase after 2 weeks with a decrease in intestinal β -glucuronidase activity, but all levels were normal after 4 weeks.

Interaction of DNT with other chemicals has not been observed in children.

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2.9 POPULATIONS THAT ARE UNUSUALLY SUSCEPTIBLE

A susceptible population will exhibit a different or enhanced response to 2,4- and 2,6-dinitrotoluene than will most persons exposed to the same level of 2,4- and 2,6-dinitrotoluene in the environment. Reasons may include genetic makeup, age, health and nutritional status, and exposure to other toxic substances (e.g., cigarette smoke). These parameters may result in reduced detoxification or excretion of 2,4- and 2,6-dinitrotoluene, or compromised function of target organs affected by 2,4- and 2,6-dinitrotoluene. Populations that are at greater risk due to their unusually high exposure to 2,4- and 2,6-dinitrotoluene are discussed in Section 5.7, Populations with Potentially High Exposure.

Humans sensitive to DNT may include individuals with cardiovascular problems. Hematological effects associated with exposure to 2,4-DNT may place persons with anemia, including sickle cell anemia or other diseases of the blood, at an increased risk.

Persons with chronic neurological disorders may also have an increased sensitivity to DNT exposure. Although there are insufficient data available to draw firm conclusions, it appears that pregnant women and their fetuses may be susceptible to the oxygen depletion implied by the hematotoxicity of DNT based on a study on rats (Jones-Price et al. 1982). Although it has been reported that alcoholics may have a decreased resistance to the effects of Tg-DNT (Perkins 19 19), the extent of this compromise has not been determined. The susceptibility of children to the health effects of DNT may be different from that of adults, as discussed in Section 2.6.

2.10 METHODS FOR REDUCING TOXIC EFFECTS

This section will describe clinical practice and research concerning methods for reducing toxic effects of exposure to 2,4- and 2,6-dinitrotoluene. However, because some of the treatments discussed may be experimental and unproven, this section should not be used as a guide for treatment of exposures to 2,4- and 2,6-dinitrotoluene. When specific exposures have occurred, poison control centers and medical toxicologists should be consulted for medical advice. The following texts provide specific information about treatment following exposures to 2,4- and 2,6-dinitrotoluene: Bronstein and Currance 1994; Ellenhom 1997.

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There are no known pediatric-specific methods for reducing peak absorption following exposure, reducing burden, or interfering with the mechanism of action for toxic effects.

2.10.1 Reducing Peak Absorption Following Exposure

Limited information from humans indicates that DNT is absorbed after inhalation exposure, while animal data suggest that DNT is rapidly and completely absorbed after oral exposure. Efforts to reduce absorption following acute exposure to DNT should focus on removing the individual from the site of exposure and decontaminating exposed areas of the body. Contaminated clothing and jewelry should be removed and skin should be washed with soap and water (Bronstein and Currance 1994). It is suggested that eyes exposed to DNT be copiously irrigated with water and normal saline (Bronstein and Currance 1994). If ingestion of DNT occurs, it is suggested that the mouth be rinsed and water can be administered for dilution if the patient can swallow, has a good gag reflex, and is not drooling (Bronstein and Currance 1994). In addition, the use of activated charcoal has been suggested (Bronstein and Currance 1994). Induction of emesis is contraindicated (Bronstein and Currance 1994). In patients who present within 2 to 4 hours of DNT ingestion, gastric lavage may be helpful in decreasing peak absorption following exposure (Ellenhorn 1997). There may also be some benefit in administering activated charcoal and cathartics after lavage (Ellenhorn 1997).

2.10.2 Reducing Body Burden

There are no data to support the use of hemodialysis, forced diuresis, hyperbaric oxygen, or hemoperfusion for treatment of methemoglobinemia alone, but these treatments may provide adjunctive care after DNT ingestion when supportive care is inadequate (Ellenhorn 1997).

2.10.3 Interfering with the Mechanism of Action for Toxic Effects

Exposure to DNT can cause profound methemoglobinemia with its sequelae (cyanosis and Heinz body formation), anoxia, and death (Ellenhorn 1997). The antidote used for serious methemoglobinemia is methylene blue (tetramethylthionine chloride), but treatment with methylene blue is not indicated for all patients (Ellenhorn 1997). Treatment with methylene blue is believed to be effective because it acts as a cofactor to increase the erythrocyte reduction of methemoglobin in the presence of NADPH (Ellenhorn 1997). Methylene blue is oxidized and the resulting molecule becomes an electron donor for the nonenzymatic

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reduction of methemoglobin to oxyhemoglobin (Ellenhorn 1997). Exchange transfusion and/or packed RBC transfusion may be useful for patients who do not respond to methylene blue or for patients with G6PD- or NADPH-methemoglobin reductase deficiencies (Ellenhorn 1997).

2.11 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of 2,4- and 2,6-DNT is available. Where adequate information is not available, ATSDR, in conjunction with the National Toxicology Program (NTP), is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of 2,4- and 2,6-dinitrotoluene.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

2.11.1 Existing Information on Health Effects of 2,4- and 2,6-Dinitrotoluene

The existing data on health effects of inhalation, oral, and dermal exposure of humans and animals to 2,4-, 2,6-, and Tg-DNT are summarized in Figures 2-7, 2-8, and 2-9, respectively. The purpose of these figures is to illustrate the existing information concerning the health effects of 2,4-, 2,6, and Tg-DNT. Each dot in the figure indicates that one or more studies provide information associated with that particular effect. The dot does not necessarily imply anything about the quality of the study or studies, nor should missing information in this figure be interpreted as a "data need." A data need, as defined in ATSDR's *Decision Guide for Identifying Substance-Specific Data Needs Related to Toxicological Profiles* (ATSDR 1989), is substance-specific information necessary to conduct comprehensive public health assessments. Generally, ATSDR defines a data gap more broadly as any substance-specific information missing from the scientific literature.

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2.11.2 Identification of Data Needs

As shown in Figures 2-7 (2,4-DNT), 2-8 (2,6-DNT), and 2-9 (Tg-DNT), there are limited data on health effects in humans, primarily for Tg-DNT, following inhalation exposure.

The available reports generally lack quantitative information on exposure levels. Human data are particularly sparse. Most toxicity studies have focused on the main systemic effects of obvious clinical significance, as described in the previous sections.

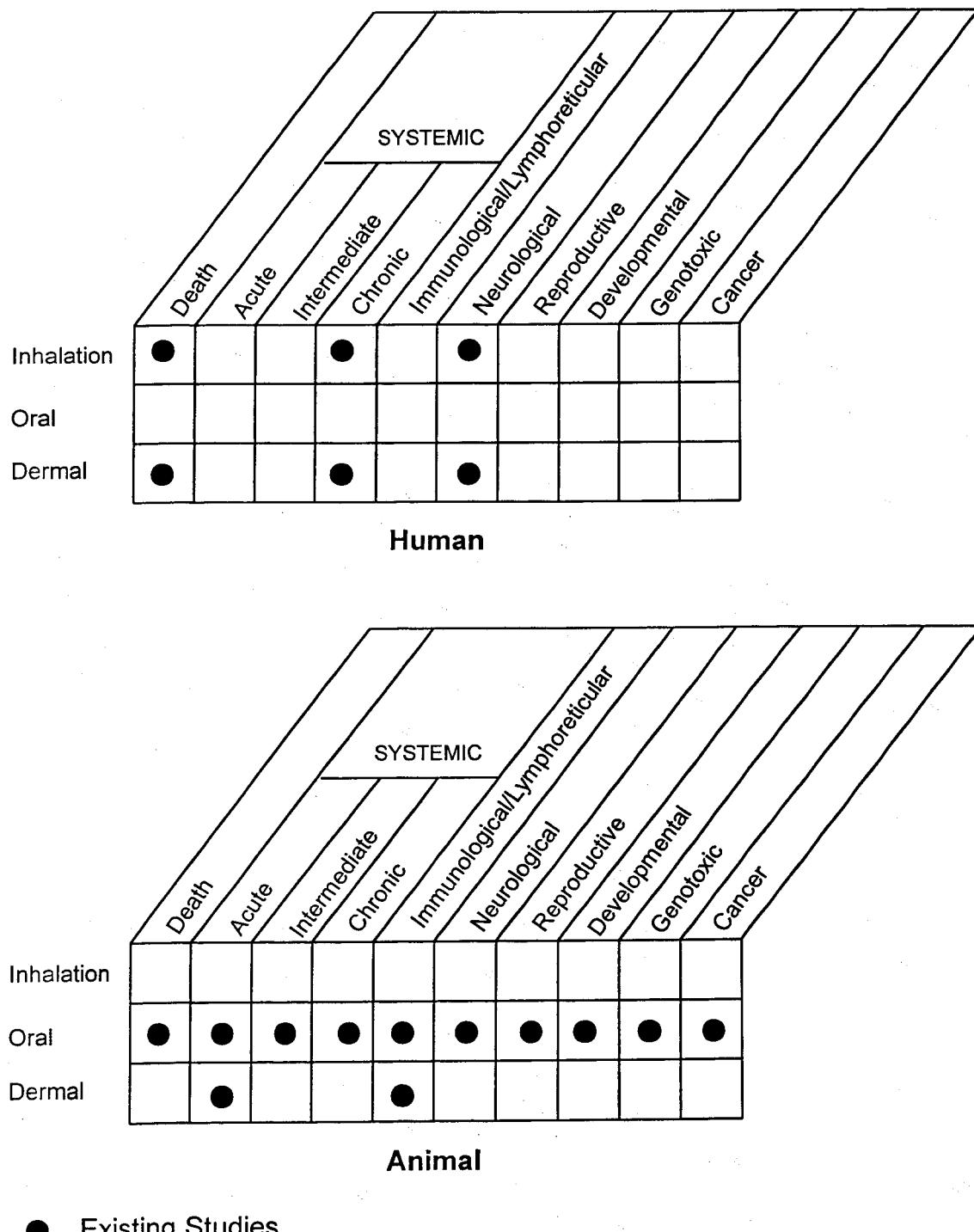
The toxicity of these chemicals has been extensively investigated in animals after oral exposure, but not after inhalation exposure, and only in a very limited way after dermal exposure. The potential carcinogenicity of these chemicals has been investigated following oral exposure in typical chronic bioassays as well as in less than-life-time studies.

Acute-Duration Exposure. Although there are no human data available from acute-duration oral exposure to 2,4- or 2,6-DNT, the data currently available from animal studies using single-dose exposure to 2,4- and 2,6-DNT are appropriate for evaluation of oral toxicity (Ellis et al. 1978; Lee et al. 1975; Vernot et al. 1977). The LD₅₀s for 2,4-DNT determined after gavage dosing ranged from 270 to 650 mg/kg in rats and from 1,340 to 1,954 mg/kg in mice (Ellis et al. 1978; Lee et al. 1975; Vernot et al. 1977). After oral administration of 2,6-DNT, LD₅₀s ranged from 180 to 795 mg/kg in rats and from 621 to 807 mg/kg in mice (Ellis et al. 1978; Lee et al. 1975; Vemot et al. 1977). Ataxia was observed in these animals before death. Slight cyanosis was observed in rats administered 60 mg/kg 2,4-DNT by gavage for 5 days (Lane et al. 1985), but no changes in hematological parameters were found in rats fed up to 273 mg/kg/day in the diet for 14 days (McGown et al. 1983). Hepatic effects, including increased blood cholesterol and alanine aminotransferase levels, and renal effects, such as hyaline droplet accumulation, were observed in rats fed 2,4-DNT in the diet for 14 days (McGown et al. 1983). An acute-duration oral MRL has been derived for 2,4-DNT from a NOAEL for neurotoxicity in dogs (Ellis et al. 1985; Lee et al. 1978). Data were insufficient to derive an acute-duration oral MRL for 2,6-DNT.

There were no acute-duration inhalation or dermal studies in humans available for evaluation. Both 2,4- and 2,6-DNT were shown to be mild primary dermal irritants in rabbits (Ellis et al. 1978; Lee et al. 1975). Acute inhalation and dermal studies would be useful for determination of route-specific toxicity.

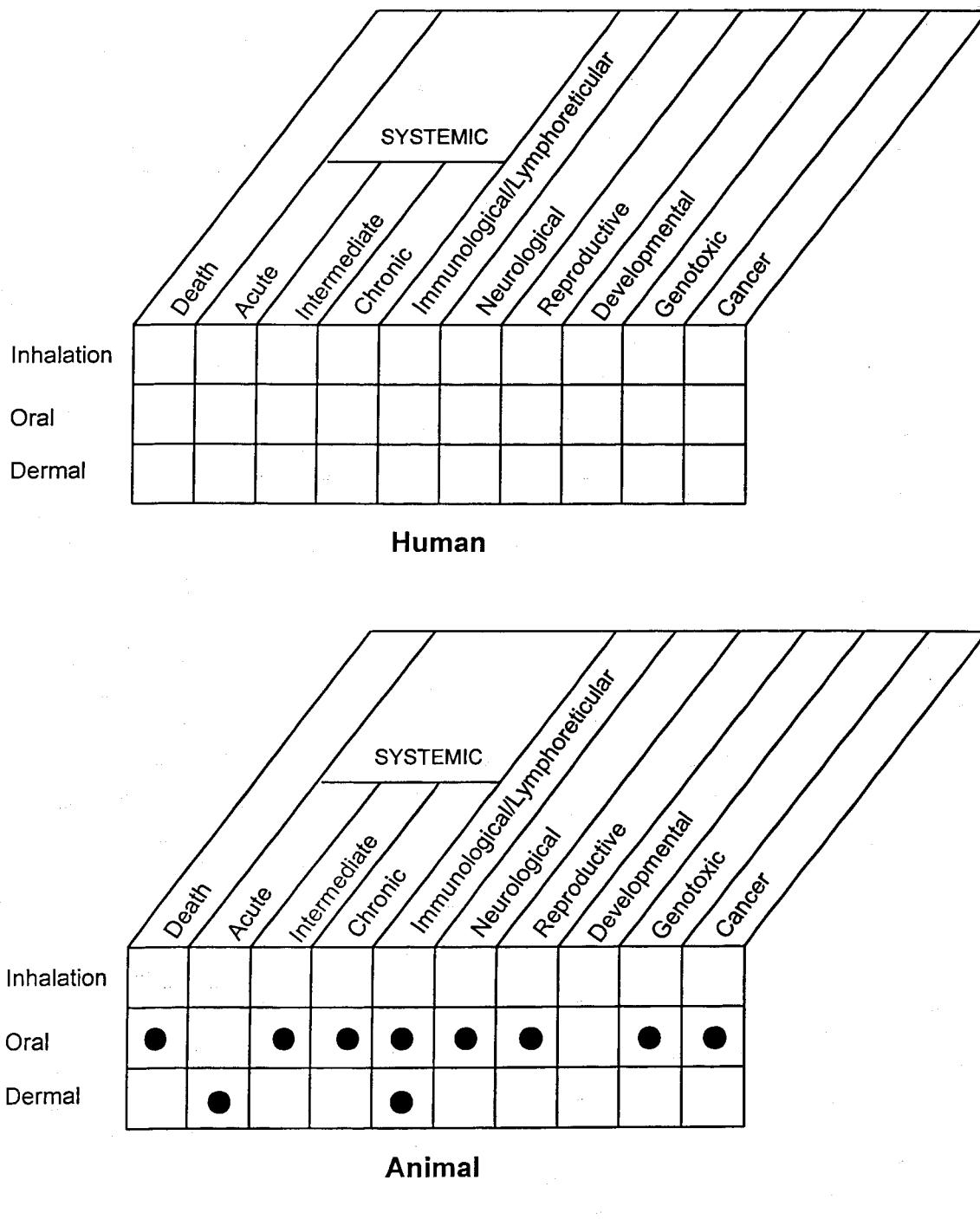
2. HEALTH EFFECTS

FIGURE 2-7. Existing Information on Health Effects of 2,4-DNT



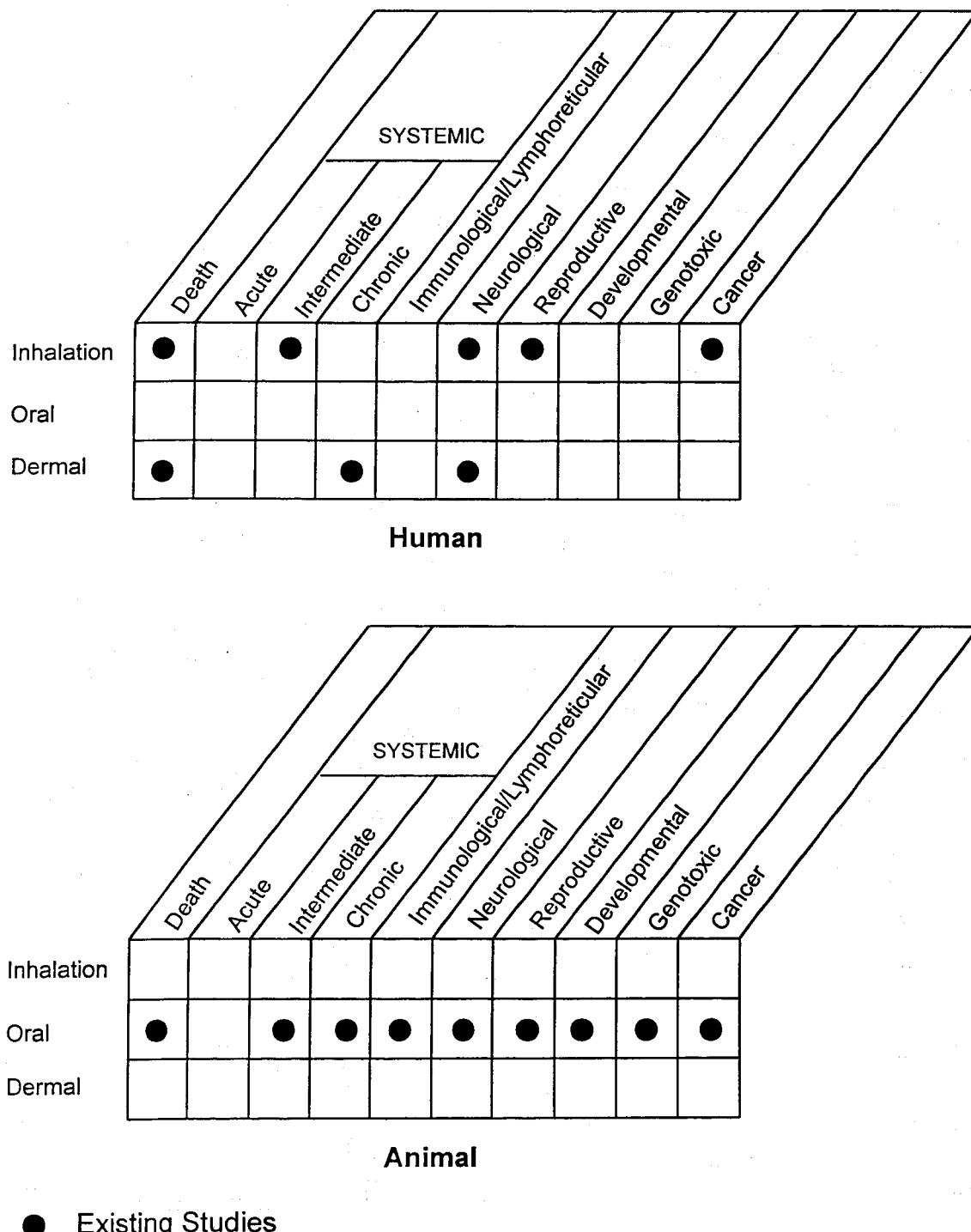
2. HEALTH EFFECTS

FIGURE 2-8. Existing Information on Health Effects of 2,6-DNT



2. HEALTH EFFECTS

FIGURE 2-9. Existing Information on Health Effects of Tg-DNT



2. HEALTH EFFECTS

Intermediate-Duration Exposure. Currently available animal studies using repeated-dose exposure are appropriate for evaluation of oral toxicity for both 2,4- and 2,6-DNT (Ellis et al. 1979, 1985; Hazleton Laboratories 1977, 1982; Hong et al. 1985; Jones-Price et al. 1982; Lee et al. 1976, 1978, 1985; McGown et al. 1983; Smith et al. 1996). Methemoglobinemia and its sequelae (Heinz bodies, anemia, reticulocytosis), hemosiderosis, extramedullary hematopoiesis, and cyanosis have been observed in animals after oral treatment with 2,4-, 2,6-, or Tg-DNT (Hazleton Laboratories 1977, 1982; Lee et al. 1976, 1978). Mild hepatocellular dysplasia was observed in mice fed 2,4-DNT in the diet for 13 weeks (Hong et al. 1985; Lee et al. 1978), but no hepatotoxicity was observed after 2,4-DNT administration to rats or dogs for the same duration (Ellis et al. 1985; Lee et al. 1978). However, treatment with 2,6-DNT did cause bile duct hyperplasia in rats and mice (Lee et al. 1976). This lesion, as well as hepatic degeneration, was observed in dogs dosed with 2,6-DNT (Lee et al. 1976).

Oral administration of 2,4-DNT to rats, mice, or dogs for 13 weeks did not cause any significant adverse renal effects (Hong et al. 1985; Lee et al. 1978). Administration of 2,6-DNT to dogs for 13 weeks caused renal inflammation and degeneration, which were not observed in rats or mice (Lee et al. 1976). Decreased body weight gain or weight loss was observed in rats and mice after administration of 2,4-DNT (Ellis et al. 1979; Hong et al. 1985; Lee et al. 1978, 1985; Leonard et al. 1987; NCI 1978) and in rats, mice, and dogs after administration of 2,6-DNT (Lee et al. 1976). An intermediate-duration oral MRL has been derived for 2,6-DNT based on a LOAEL for hematological effects in dogs (Lee et al. 1976). Subchronic inhalation and dermal studies would be useful for determination of toxic effects in order to derive an MRL for 2,4-DNT and to determine a mechanism of action from routes of exposure that are more characteristic of occupational exposure.

Chronic-Duration Exposure and Cancer. There are no data available in humans regarding the carcinogenicity of 2,4- or 2,6-DNT. A retrospective cohort mortality study performed using data from workers at ammunition plants that used 2,4- or Tg-DNT found no increases in mortality due to either malignant neoplasms as a whole or from particular cancers (Levine et al. 1986b). However, the small cohort examined in this study limited its statistical power. Both 2,4- and 2,6-DNT have been found to cause hepatocellular carcinoma in rats (Ellis et al. 1979; Leonard et al. 1987). Renal cancer was observed in mice after administration of 2,4-DNT in the diet (Ellis et al. 1979). An upperbound qi^* has been derived for oral exposure to 2,4-DNT/2,6-DNT mixture (EPA 1986d, 1998).

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Excessive mortality rates from ischemic heart disease and residual diseases of the circulatory system were observed in ammunition plant workers (Levine et al. 1986a). Because it is expected that these workers would have a lower incidence of cardiovascular disease due to the "healthy worker effect," this finding is unusual. Further epidemiological studies to verify these findings are needed. Studies performed in nonhuman primates to investigate further the potential cardiovascular effects of both chemicals would help to elucidate the mechanism of heart disease observed.

The currently available studies in laboratory animals on the effects of 2,4- and 2,6-DNT after chronic exposure are appropriate for evaluation of chronic oral toxicity (Ellis et al. 1979, 1985; Hazleton Laboratories 1982; Lee et al. 1978, 1985; Leonard et al. 1978; NCI 1978). Hematological effects, including anemia, compensatory anemia, methemoglobinemia, and Heinz bodies have been observed after chronic administration of 2,4-DNT to dogs, mice, and rats (Ellis et al. 1979, 1985; Hong et al. 1985; Lee et al., 1978). A chronic-duration oral MRL has been derived for 2,4-DNT based on a NOAEL of 0.2 mg/kg for hematological and neurological effects and biliary hyperplasia in dogs (Ellis et al. 1979, 1985). Data were insufficient for the derivation of a chronic-duration oral MRL for 2,6-DNT. Severe hepatocellular changes, such as degeneration and vacuolation and dysplasia, were found in rats, mice, and dogs administered 2,4- or 2,6-DNT for chronic durations in oral exposure studies (Ellis et al. 1979, 1985; Hong et al. 1985; Leonard et al. 1987). Renal cystic dysplasia was observed in mice, but not rats or dogs, treated orally with 2,4-DNT for chronic-duration periods (Ellis et al. 1979; Hong et al. 1985). Chronic-duration studies have not been performed in mice using 2,6-DNT to determine whether these findings would also result after administration of this isomer. Although no histopathological effects were found in adrenal, pituitary, or thyroid glands of rats after chronic oral administration of Tg-DNT, increases in parathyroid hyperplasia, fatty metamorphosis, and vascular ectasia were found (Hazleton Laboratories 1982). Further studies may be useful to verify these findings. Effects on body weight, including body weight loss, were reported in almost all chronic-duration oral studies (Ellis et al. 1979, 1985; Hazleton Laboratories 1982; Hong et al. 1985; Leonard et al. 1987; NCI 1978).

A well-controlled chronic inhalation study and dermal studies would be useful for determination of the potential for route-specific toxicity. In addition, for both 2,4- and 2,6-DNT, well-controlled epidemiological evaluations of larger occupationally exposed populations would contribute valuable insights regarding the human relevancy of chronic health effects observed in animal studies.

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Genotoxicity. Both 2,4- and 2,6-DNT cause gene mutations in the reverse mutation assay using *S. typhimurium*. (Couch et al. 1981; Dellarco and Prival 1989; Ellis et al. 1978; Spanggord et al. 1982b; Tokiwa et al. 1981). However, the test system has given variable results because of the need for metabolic activation and the sensitivity of the tester strains. *In vivo* assays using 2,4-DNT have shown unscheduled DNA synthesis and S-phase synthesis using rat hepatocytes (Ashby et al. 1985; Mirsalis and Butterworth 1982; Mirsalis et al. 1989), chromosomal aberrations using human lymphocytes (Huang et al. 1995), and DNA binding in rat hepatocytes (Chadwick et al. 1993; La and Froines 1993). The genotoxicity of Tg-DNT is believed to be due to the potent genotoxicity of 2,6-DNT, as evidenced in an *in vivo-in vitro* hepatocyte UDS system (Mirsalis and Butterworth 1982). Both 2,4- and 2,6-DNT have induced DNA adducts in rat liver (La and Froines 1992, 1993). Studies currently available for 2,4- and 2,6-DNT are considered to be appropriate for evaluation of genotoxicity.

Reproductive Toxicity. The currently available laboratory data on reproductive toxicity are considered appropriate for evaluation of oral exposure of animals to both isomers. Several studies in rats, mice, and dogs with either isomer have shown impairment of the male reproductive system. The effects observed include testicular atrophy, degeneration of the seminal vesicles, and decreased sperm production (Bloch et al. 1988; Ellis et al. 1979; Lee et al. 1976, 1978). *In vitro* studies have shown that the testicular degeneration is due, at least in part, to structural changes in Sertoli cells (Reader and Foster 1990). Animal studies of reproductive toxicity using inhalation exposure would provide information relative to occupational exposure conditions.

Several assessments of reproductive function in exposed workers have been performed that did not detect differences in sperm production or fertility rates as a result of exposure (Ahrenholz and Meyer 1982; Hammill et al. 1982; Levine et al. 1985a). However, an earlier study reported a significant reduction in the sperm counts of exposed workers, as well as an increase, of marginal statistical significance, in the number of spontaneous abortions in their wives (Ahrenholz 1980). These studies were all limited by the small exposure populations studied and the lack of historical individual exposure monitoring. Further epidemiological studies of larger exposed occupational populations with exposure data may be considered useful since questions of potential reproductive effects associated with these exposures have not yet been clearly resolved.

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Developmental Toxicity. No data are available regarding developmental effects in humans after oral exposure to DNT, but animal studies that have been performed show possible developmental effects. The only developmental effect observed in a three-generation reproductive study in rats using 2,4-DNT was a decrease in pup viability. This decrease was attributed to maternal neglect and a high incidence of maternal death during parturition. Tg-DNT administered to pregnant dams caused a decrease in relative liver weight in postpartum pups and possible transient neurotoxicity (Jones-Price et al. 1982). Further studies may be useful to elucidate these effects. Additional animal studies using 2,4- and 2,6-DNT by oral and inhalation routes should analyze fetal and maternal blood for hematological parameters. This is recommended because any factor that could reduce the amount of oxygen to developing tissue is expected to have adverse consequences in the offspring.

Immunotoxicity. Although no data are available regarding immunological or lymphoreticular effects in humans, some data on these endpoints are available in animals. The currently available information on the potential immunotoxic effects of 2,4- and 2,6-DNT is sufficient to describe the sensitizing potential of DNT. Mild sensitization has been reported in guinea pigs after dermal exposure to 2,6-DNT, but not 2,4-DNT (Ellis et al. 1978; Lee et al. 1975). No effects on IgE, the antibody associated with allergic or hypersensitive reactions, were reported in rats or dogs exposed to either the 2,4- or the 2,6-DNT isomer (Ellis et al. 1985; Lee et al. 1976, 1978, 1985). Studies have not been performed that would describe effects on immunocompetence following exposure to DNT. A battery of immunotoxicity tests would provide a better assessment of possible effects in humans.

Neurotoxicity. The nervous system has been shown to be a major target of 2,4- and 2,6-DNT toxicity in animals (Ellis et al. 1979, 1985; Kozuka et al. 1979; Lee et al. 1979, 1985). Clinical signs in dogs have included incoordination and stiffness of the hind legs leading to complete paralysis; cerebellar vacuolation, hypertrophy, and focal gliosis; cerebellar and brain stem hemorrhage. In mice, depression and hyperexcitability were observed, while some rats administered 2,6-DNT showed neuromuscular symptoms. More systematic examination of the neurological effects of these compounds in laboratory animals would be useful to assess fully behavioral abnormalities and morphological damage to the nervous system. The biochemical mechanism of dinitrotoluene neurotoxicity is not known.

Generalized symptoms of neurotoxicity, including headache, sleepiness, dizziness, and tingling pain in the extremities were reported in workers occupationally exposed to 2,4-DNT (McGee et al. 1942; Perkins 19 19). However, the more recent occupational studies performed failed to examine workers for symptoms of

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neurotoxicity (Ahrenholz 1980; Ahrenholz and Meyer 1982; Hammill et al. 1982; Levine et al. 1985a). Because the early reports of potential neurotoxicity in exposed workers have not been followed-up in more recent studies, neurological examination of workers in occupational studies could provide additional information regarding the potential magnitude of neurotoxic effects.

Epidemiological and Human Dosimetry Studies. Epidemiology studies of workers exposed to DNT suggest a potential for heart disease in exposed populations (Levine et al. 1986a). Doses of DNT associated with heart disease in humans have not been determined. Further studies with historical cohort monitoring data would be useful to verify these findings.

Animal studies have indicated that the male reproductive system is a target of DNT toxicity. Epidemiological studies have provided only suggestive evidence of a reproductive effect in workers exposed to DNT. Studies of larger worker populations may help to determine more conclusively the magnitude of the potential for reproductive toxicity in exposed humans.

Other effects that were observed in animal studies but not confirmed in human populations include liver and kidney toxicity, neurotoxicity, and cancer. Well-controlled epidemiological studies examining these endpoints in humans would be useful.

Biomarkers of Exposure and Effect

Exposure. Recently a rapid, accurate method for determining exposure to DNT has been developed using spectrophotometric analysis of complexes of primary arylamines, which result from the reduction of DNT and its metabolites (Smith et al. 1995).

Effect. Epidemiological studies that correlate quantitative estimates of exposure with disease outcomes would be useful. Studies that identify subtle physiological changes, such as altered blood chemistry indices, associated with a particular disease state are not available.

A disease registry is not currently available. The development of a registry of exposures and diseases would provide a useful reference tool for assessing the variations in exposure concentrations and health effects from, for example, geography, season, regulatory actions, presence of hazardous waste landfills, or manufacturing

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and use facilities. These assessments, in turn, would provide a better understanding of the needs for some types of research or data acquisition based on the current exposure concentrations.

Absorption, Distribution, Metabolism, and Excretion. The toxicokinetics of 2, 4- and 2,6-DNT in rats by the oral route have been extensively studied. That Tg-DNT is absorbed and excreted in the urine by humans in an occupational setting, where the main routes of absorption are considered to be inhalation and dermal, has also been documented. There are no data available in animals on the toxicokinetics of DNT by the dermal or inhalation routes. Toxicokinetics studies in rats administered the test materials by the inhalation and dermal routes would be critical in understanding possible differences in the toxicity of DNT by different routes of administration. The main routes of exposure of humans are dermal and inhalation. Understanding the possible differences in toxicity in animals by different routes would be valuable in determining the significance of findings to humans who may be exposed by inhalation or dermal routes.

Comparative Toxicokinetics. Absorption and excretion studies in several species indicate that there are considerable differences between mice and the other species evaluated. More detailed study of the metabolism of DNT by mice, including the role of biliary excretion and enterohepatic cycling, would assist in understanding why the metabolism in mice is different from other species and which species may be the most appropriate model for evaluating hazards and risks to humans.

Methods for Reducing Toxic Effects. The most important method for reducing the toxic effects of DNT is removal of the person from the area of exposure. Skin and eyes should be rinsed copiously (Bronstein and Currance 1994), although absorption through the skin has not been adequately examined. Gastric lavage, with subsequent administration of activated charcoal, and cathartics may be of some benefit in reducing peak absorption after oral exposure to DNT. Methylene blue treatment is used with patients presenting with serious methemoglobinemia (Ellenhorn 1997). No additional studies are considered necessary at this time to examine further methods for reducing body burden of DNT. Further studies on supportive therapy after DNT exposure, such as the use of hemodialysis, forced diuresis, hyperbaric oxygen, or hemoperfusion might be useful.

Children's Susceptibility. No data are available on the health effects of DNT on exposed children. Few usable data are available on whether the developmental process is altered by exposure to DNT; this information is summarized above under the Developmental Toxicity heading.

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There is inadequate experimental evidence to evaluate if the pharmacokinetics of DNT are different in children. There are no studies on whether DNT or its active metabolites can cross the placenta or be excreted in breast milk, so it cannot be determined if fetuses may be exposed *in utero* or if infants may be exposed via breast milk ingestion. There are also no data to show if DNT and its metabolites are stored in maternal tissues and thus might be later mobilized during gestation or lactation; however, DNT and its metabolites are not likely to be stored because of their low octanol-water partition coefficient.

There is little experimental evidence to evaluate whether the metabolism of DNT or its mechanisms of action are different in children. As discussed in section 2.6, newborns are highly sensitive to the methemoglobin-generating effect of DNT because of their deficiency in methemoglobin reductase (Gruener 1976), which reduces methemoglobin back to hemoglobin. In addition, newborns have a high concentration of fetal hemoglobin in their erythrocytes. It will be useful to determine if fetal hemoglobin is more sensitive to the methemoglobin-generating effect of DNT. It will also be helpful to have data on the metabolism and mechanism of action of DNT on children to determine if children are more vulnerable than adults to health effects from exposure to DNT, as some enzymes involved in DNT metabolism are known to have developmental regulation. There are no biomarkers of exposure or effect that have been validated in children or in adults exposed as children. There are no data to determine whether there are any interactions with other chemicals unique to children, or whether interactions observed in adults also occur in children. Although DNT is shown to be genotoxic, it is not known if parental exposure to DNT may affect children via parental germ cells, or if DNT may indirectly affect the fetus during maternal exposure.

Child health data needs relating to exposure are discussed in 5.8.1 Data Needs: Exposures of Children.

2.11.3 Ongoing Studies

No information was located regarding ongoing research related to the potential health effects of 2,4- or Tg-DNT.

A project to develop and apply biomarkers of exposure associated with 2,6-DNT is being conducted by S. Rappaport at the University of North Carolina at Chapel Hill (FEDRIP 1997). The project will ultimately perform epidemiological studies of exposed populations in order to provide exposure-biomarker relationship. It is anticipated that *in vivo* and *in vitro* measurements of the protein adducts will provide data on the disposition of reactive intermediates and mechanisms of toxicity.

3. CHEMICAL AND PHYSICAL INFORMATION

3.1 CHEMICAL IDENTITY

Information regarding the chemical identity of 2,4- and 2,6-DNT is located in Table 3-1.

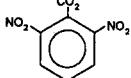
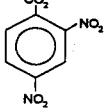
3.2 PHYSICAL AND CHEMICAL PROPERTIES

Information regarding the physical and chemical properties of 2,4- and 2,6-DNT is located in Table 3-2.

Data regarding specific isomers of DNT have been provided whenever possible. However, DNT is generally produced as a technical-grade mixture of the two isomers, with approximately 5% other substances. This 5% contains predominantly the other isomers of DNT, which are not discussed in this profile. Where information pertains to Tg-DNT, it has been so noted.

3. CHEMICAL AND PHYSICAL INFORMATION

TABLE 3-1. Chemical Identity of 2,4- and 2,6-Dinitrotoluene

Characteristic	Information	Information	Reference
Chemical Name	2,4-Dinitrotoluene	2,6-Dinitrotoluene	HSDB 1998
Synonym(s)	1-Methyl-2,4-dinitrobenzene; 2,4-dinitrotoluol; 2,4-DNT	1-Methyl-2,6-dinitrobenzene; 2,6-DNT	HSDB 1998
Registered trade name(s)	No data	No data	
Chemical formula	C ₇ H ₆ N ₂ O ₄	C ₇ H ₆ N ₂ O ₄	HSDB 1998
Chemical structure			
Identification numbers:			
CAS registry	121-14-2	606-20-2	HSDB 1998
NIOSH RTECS	XT1575000	XT1925000	
EPA hazardous waste	U105	U106	HSDB 1998
OHM/TADS	7800118	8300219	HSDB 1998
DOT/UN/NA/IMCO shipping	IMO 6.1 UN 1600 (molten) UN 2038 (solid or liquid)	6.1 UN 1600 (molten) UN 2038 (solid or liquid)	HSDB 1998
HSDB	1144	2931	HSDB 1998
NCI	NCI-C01865	No data	HSDB 1998

CAS = Chemical Abstracts Services; DOT/UN/NA/IMCO = Department of Transportation/United Nations/North America/International Maritime Dangerous Goods Code; EPA = Environmental Protection Agency; HSDB = Hazardous Substances Data Bank; NCI = National Cancer Institute; NIOSH = National Institute for Occupational Safety and Health; OHM/TADS = Oil and Hazardous Materials/Technical Assistance Data System; RTECS = Registry of Toxic Effects of Chemicals Substances

3. CHEMICAL AND PHYSICAL INFORMATION

TABLE 3-2. Physical and Chemical Properties of 2,4- and 2,6-Dinitrotoluene

Property	Information	Information	Reference
Chemical name	2,4-Dinitrotoluene	2,6-Dinitrotoluene	Lide 1993
Molecular weight	182.14	182.14	HSDB 1998
Color	Yellow	Yellow to red	HSDB 1998
Physical state	Solid	Solid	HSDB 1998
Melting point	71°C	66°C	HSDB 1998
Boiling point	300°C (slight decomposition)	285°C	HSDB 1998
Density	1.3208 (71°C)	1.2833 (111°C)	HSDB 1998
Odor	Slight	Slight	HSDB 1998
Odor threshold:			
Water	No data	No data	HSDB 1998
Air	No data	No data	HSDB 1998
Solubility:			
Water, mg/L	0.03g/100g (22°C)	180 (20°C)	HSDB 1998, Mabey et al. 1982
Organic solvents	Soluble in acetone, alcohol, benzene, ethanol, diethyl ether, pyridine, CS ₂	Soluble in alcohol	Lide 1993
Partition coefficients:			
Log K _{ow}	1.98	1.72 (estimated)	HSDB 1998
Log K _{oc}			
Vapor pressure, torr 25°C	1.4 × 10 ⁻⁴	5.67 × 10 ⁻⁴	HSDB 1998
Henry's law constant: atm-cu m/mol	8.79 × 10 ⁻⁸	9.26 × 10 ⁻⁸	HSDB 1998
Autoignition temperature,	No data	No data	HSDB 1998
Flashpoint, CC	404°F	404°F	HSDB 1998
Flammability limits	No data	No data	HSDB 1998
Conversion factors			
ppm (v/v) to mg/m ³ in air (20°C)	1 ppm = 7.40 mg/m ³	1 ppm = 7.40 mg/m ³	HSDB 1998
mg/m ³ to ppm (v/v) in air (20°C)	1 mg/m ³ = 0.13 ppm	1 mg/m ³ = 0.13 ppm	HSDB 1998

4. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

4.1 PRODUCTION

Tables 4-1 and 4-2 list the facilities in each state that manufacture or process 2,4-DNT and 2,6-DNT, respectively; the intended use, and the range of maximum amounts of 2,4- and 2,6-DNT that are stored on site. There are currently 3 facilities that produce or process 2,4-DNT in the United States: First Chemical Corporation in Pascagoula, MS; U.S. Army Radford Army in Radford, VA; and Bayer Corporation in New Martinsville, WV. There is one facility that produces or processes 2,6-DNT in the United States: Bayer Corporation in New Martinsville, WV. The data listed in Tables 4-1 and 4-2 are derived from the Toxics Release Inventory (TR196 1998). Only certain types of facilities were required to report. Therefore, this is not an exhaustive list.

The chemicals 2,4- and 2,6-DNT are generally produced as a mixture called Tg-DNT, which contains approximately 76.5% 2,4-DNT and 18.8% 2,6-DNT (with the remainder consisting of other isomers and minor contaminants such as TNT and mononitrotoluenes) (HSDB 1998). This mixture is commercially prepared by the nitration of toluene with concentrated sulfuric and nitric acid (Etnier 1987).

4.2 IMPORT/EXPORT

No data describing import or export activities for 2,4- or 2,6-DNT were located. Imports of nitrobenzenes and nitrotoluenes into the United States for 1994 totaled 1,676,582 kg (NTDB 1996).

4.3 USE

The most commercially important use of DNT is as a chemical intermediate in the production of toluene diisocyanate, a precursor to polyurethane polymers (HSDB 1998). It has been estimated that 99% of all DNT produced is used for this purpose (CMR 1983). Additionally, DNT is used in the production of TNT, and as a waterproofing, plasticizing, and gelatinizing agent in explosives (HSDB 1998). DNT is also used as an intermediate in the production of dyes and as a modifier for smokeless powders in the munitions industry (HSDB 1998). 2,4-DNT is used in the air bags of automobiles (Ellenhorn 1997).

TABLE 4-1. Facilities that Manufacture or Process 2,4-Dinitrotoluene

FACILITY	LOCATION ^a	RANGE OF MAXIMUM AMOUNTS ON SITE IN POUNDS	ACTIVITIES AND USES
FIRST CHEMICAL CORP.	PASCAGOULA , MS	1,000 - 9,999	PRODUCE , BYPRODUCT
U.S. ARMY RADFORD ARMY	RADFORD , VA	100,000 - 999,999	FORMULATION COMPONENT
BAYER CORP.	NEW MARTINSVILLE , WV	100,000 - 999,999	PRODUCE , ON-SITE USE/PROCESSING , REACTANT

Source: TRI96 1998

^a Post Office state abbreviations used

TABLE 4-2. Facilities that Manufacture or Process 2,6-Dinitrotoluene

FACILITY	LOCATION ^a	RANGE OF MAXIMUM AMOUNTS ON SITE	
		IN POUNDS	ACTIVITIES AND USES
BAYER CORP.	NEW MARTINSVILLE, WV	100,000 - 999,999	PRODUCE, ON-SITE USE/PROCESSING, REACTANT

Source: TRI96 1998

^aPost Office state abbreviations used

4. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

4.4 DISPOSAL

2,4-DNT and 2,6-DNT are listed as toxic substances under Section 313 of the Emergency Planning and Community Right to Know Act (EPCRA) under Title III of the Superfund Amendments and Reauthorization Act (SARA) (EPA 1995). Disposal of wastes containing 2,4-DNT and 2,6-DNT is controlled by a number of federal regulations (see Chapter 7).

Only limited information is available regarding the appropriate disposal of DNT. NTOSH recommends small quantities be swept onto paper or other suitable material and incinerated in a suitable combustion chamber. Larger quantities should be reclaimed; if this is not practical, then they should be dissolved in fuel oil and atomized in a suitable combustion chamber (HSDB 1998). DNT has also been proposed as a potential candidate for rotary kiln incineration at 820-1,600°C or fluidized bed incineration at 450-980°C, with residence times of seconds for gases and liquids and longer for solids (HSDB 1998).

5. POTENTIAL FOR HUMAN EXPOSURE

5.1 OVERVIEW

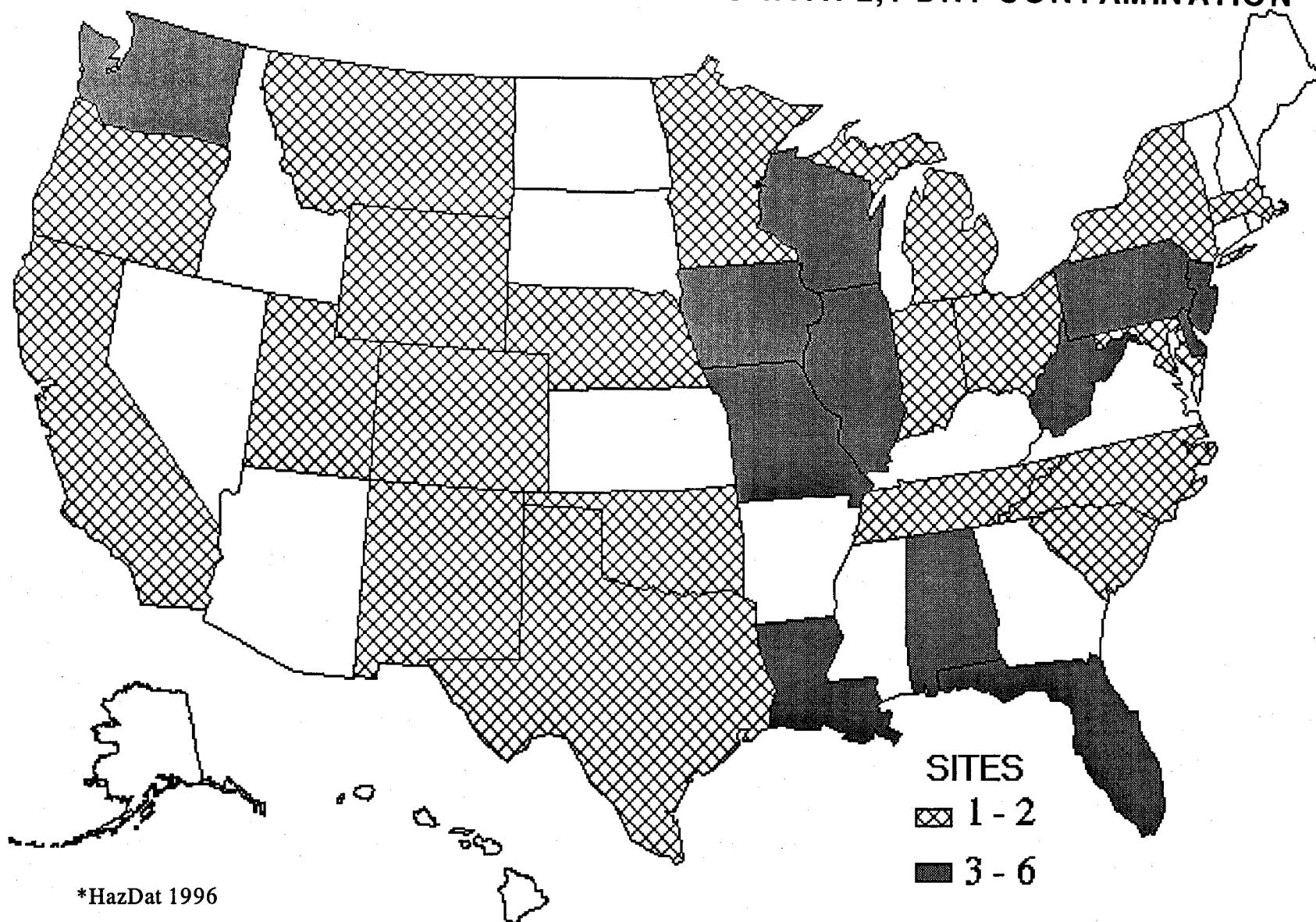
2,4-DNT and 2,6-DNT have been identified in at least 69 and 53 of the 1,467 current or former EPA National Priorities List (NPL) hazardous waste sites, respectively (HazDat 1998). However, the number of sites evaluated for 2,4-DNT and 2,6-DNT is not known. The frequency of these sites within the United States can be seen in Figures 5-1 and 5-2.

The available data provide a complex and incomplete view of the overall potential for human exposure to isomers of DNT. Little direct knowledge of the magnitude of environmental exposure pathways exists. Data regarding exposure of humans to 2,4- and 2,6-DNT have been obtained primarily from the workplace.

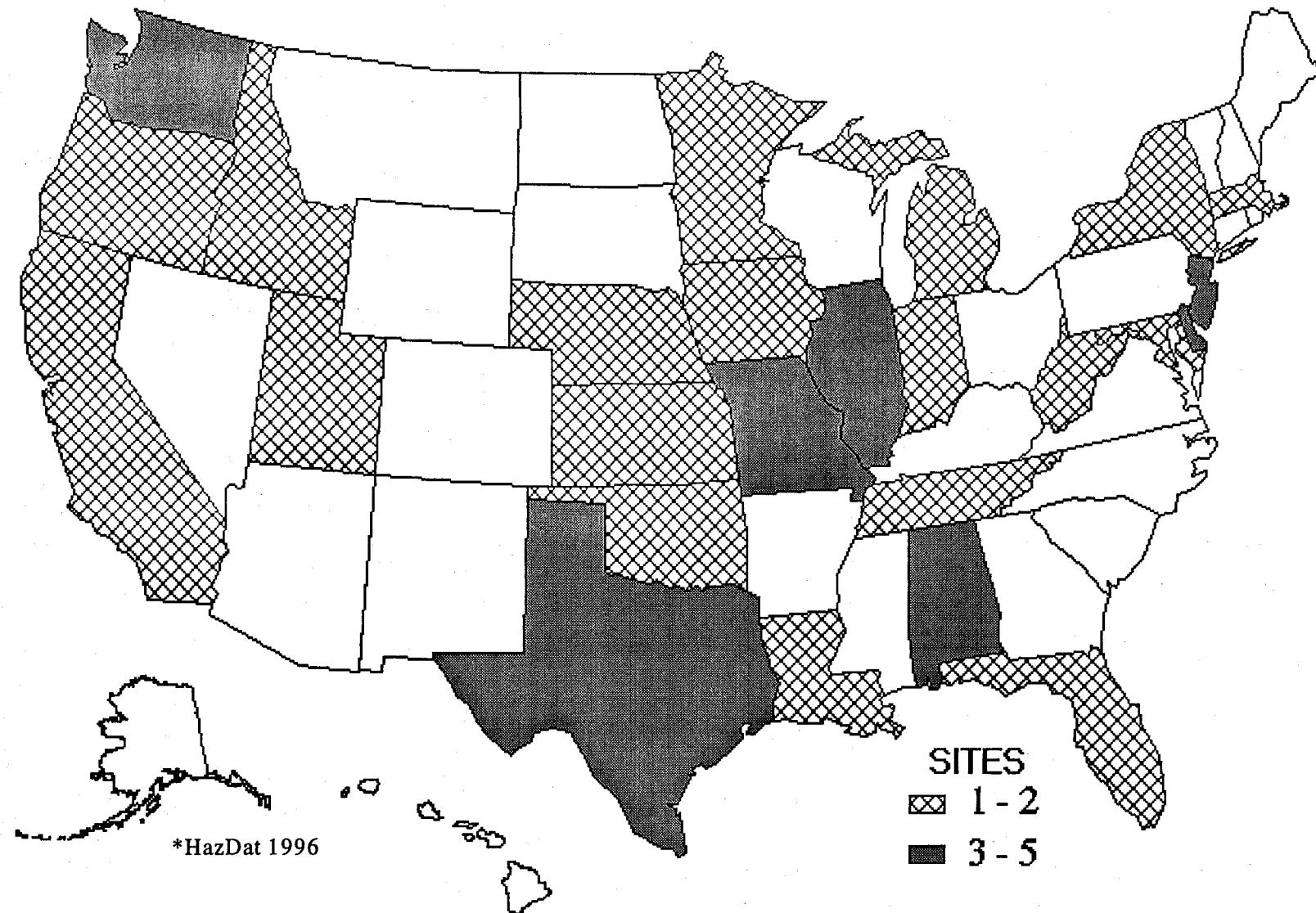
DNT has been found in waste water and groundwater in and around munitions sites (Jenkins et al. 1986), and 4-nitrotoluene and dinitrobenzene, structural analogues of DNT, are taken up by plants (McFarlane et al. 1987). However, predictions of environmental exposure pathways based on measurements of structural analogues of DNT are severely limited by the complex abiotic reactions of DNT in the environment and by the different pathways, rates, and products of biological reduction and/or oxidation of 2,4-DNT and 2,6-DNT.

The relatively low log octanol-water partition coefficients of the DNT isomers (1.98 and 1.72 for 2,4-DNT and 2,6-DNT, respectively) predict that DNT released to the environment would not bioaccumulate. However, the log octanol-water partition coefficients are sufficiently large to indicate adsorption to soil organic matter. The bioavailability and potential toxicity of soil- and sediment-bound products are also unknown. DNT is degraded by oxidation, photolysis, and biotransformation in water or soil, but a variety of degradation products, about which very little is known, is formed. DNT may bind to clays and clay colloids and facilitate its oxidation. It is otherwise expected to be stable in water because it is not hydrolyzed. The relatively low volatility and high solubility of DNT indicates that it will tend to remain in water for long periods of time, unless acted upon by light, oxygen, or biota. As a result, DNT can be transported to groundwater or surface waters.

FIGURE 5-1. FREQUENCY OF NPL SITES WITH 2,4-DNT CONTAMINATION*



*HazDat 1996

FIGURE 5-2. FREQUENCY OF NPL SITES WITH 2,6-DNT CONTAMINATION*

5. POTENTIAL FOR HUMAN EXPOSURE

Studies of occupational exposures to DNT indicate that inhalation and dermal contact can result in absorption of DNT into the body. However, inhalation and dermal contact are less likely to be critical environmental exposure pathways for the general population. Individuals exposed outside the workplace would be more likely to ingest DNT via contaminated drinking water and food.

5.2 RELEASES TO THE ENVIRONMENT

According to the Toxics Release Inventory (TRI), in 1996, a total of 6,844 pounds (5,080 kg) of 2,4-DNT from 3 large processing facilities and 1,315 pounds (592 kg) 2,6-DNT from 2 large processing facilities were released to the environment (TR196 1998). Tables 5-1 and 5-2 list amounts released from these facilities. In addition, an estimated 1,866 pounds (840 kg) of 2,4-DNT and 58 pounds (26 kg) 2,6-DNT were transferred offsite (TR196 1998). The TRI data should be used with caution because only certain types of facilities are required to report. This is not an exhaustive list.

2,4-DNT and 2,6-DNT have been identified in a variety of environmental media (air, surface water, groundwater, soil, and sediment) collected at 69 and 53 of the 1,467 NPL hazardous waste sites, respectively (HazDat 1998).

5.2.1 Air

According to the Toxics Release Inventory, in 1996, the estimated releases of 4,202 pounds (1,891 kg) 2,4-DNT from 3 large processing facilities and 1,048 pounds (472 kg) 2,6-DNT from 1 large processing facility to air accounted for about 77% of total environmental releases (TR196 1998). Tables 5-1 and 5-2 list amounts released from these facilities. The TRI data should be used with caution, however, since only certain types of facilities are required to report. This is not an exhaustive list.

2,4-DNT has been identified in air samples collected at 1 of the 69 NPL hazardous waste sites, and 2,6-DNT has been identified in air samples collected at 1 of the 53 NPL hazardous waste sites where they were detected in some environmental media (HazDat 1998).

The DNT isomers have relatively low vapor pressures (2,4-DNT, 5.1×10^{-3} torr [20°C]; 2,6-DNT, 0.018 torr [20°C]) (Maksimov 1968) and quite low Henry's Law constants (2,4-DNT, 4.5×10^{-6} atm-m³ mol⁻¹ [calc., 20°C]; 2,6-DNT, 7.9×10^{-6} atm-m³ mol⁻¹ [calc., 20°C]) (Mabey et al. 1982). As a result, DNT is not

TABLE 5-1. Releases to the Environment from Facilities that Manufacture or Process 2,4-Dinitrotoluene

STATE ^b	CITY	FACILITY	Reported amounts released in pounds per year ^a					TOTAL ENVIRONMENT ^d
			AIR ^c	WATER	LAND	UNDER GROUND INJECTION	POTW TRANSFER	
MS	PASCAGOULA	FIRST CHEMICAL CORP.	3	0	0	0	0	0
VA	RADFORD	U.S. ARMY RADFORD ARMY	5	250	0	0	0	840
WV	NEW MARTINSVILLE	BAYER CORP.	1,883	99	0	0	0	1,982
TOTALS			1,891	349	0	0	0	3,080

Source: TRI96 1998

^aData in TRI are maximum amounts released by each facility^bPost office state abbreviations used^cThe sum of fugitive and stack releases are included in releases to air by a given facility^dThe sum of all releases of the chemical to air, land, and water, and underground injection wells; and transfers off-site by a given facility

POTW = publicly owned treatment works

TABLE 5-2. Releases to the Environment from Facilities that Manufacture or Process 2,6-Dinitrotoluene

Reported amounts released in pounds per year ^a									
STATE ^b	CITY	FACILITY	AIR ^c	WATER	LAND	UNDERGROUND INJECTION	POTW TRANSFER	OFF-SITE WASTE TRANSFER	TOTAL ENVIRONMENT ^d
WV	NEW MARTINSVILLE	BAYER CORP.	472	94	0	0	0	26	592
		TOTALS	472	94	0	0	0	26	592

Source: TRI96 1998

^aData in TRI are maximum amounts released by each facility^bPost office state abbreviations used^cThe sum of fugitive and stack releases are included in releases to air by a given facility^dThe sum of all releases of the chemical to air, land, and water, and underground injection wells; and transfers off-site by a given facility

POTW = publicly owned treatment works

5. POTENTIAL FOR HUMAN EXPOSURE

expected to volatilize from water. However, DNT may be released to or transported in the air as dusts or aerosols or adsorbed to other suspended particles. Exposure to airborne DNT may occur in the workplace. Environmental exposure to DNT in the air may occur if contaminated surface soils are eroded and reentrained.

Minute amounts of nitrotoluene are formed by the photochemical reaction of toluene, nitrogen oxides, and sunlight (Atkinson et al. 1980). Although DNT could be formed subsequently, it would be subject to photolysis and would not be likely to accumulate enough to contribute significantly to human exposure.

5.2.2 Water

According to the Toxics Release Inventory, in 1996, the estimated releases of 2,4-DNT and 2,6-DNT of 776 pounds (349 kg) from 2 large processing facilities and 209 pounds (94 kg) from 1 large processing facility, respectively, to water accounted for about 12% of total environmental releases (TRI96 1998). Tables 5-1 and 5-2 list amounts released from these facilities. The TRI data should be used with caution, however, since only certain types of facilities are required to report. This is not an exhaustive list.

2,4-DNT has been identified in surface water at 19 of 69 NPL hazardous waste sites, and in groundwater at 54 of 69 NPL hazardous waste sites where it was detected in some environmental media (HazDat 1998). 2,6-DNT has been identified in surface water at 18 of 53 NPL hazardous waste sites, and in groundwater at 32 of 53 NPL hazardous waste sites where it was detected in some environmental media (HazDat 1998).

The occurrence of DNT in water has been reported (Feltes et al. 1990; Shackelford and Keith 1976; Staples et al. 1985). Both 2,4- and 2,6-DNT are recognized as major components in waste waters from TNT manufacturing facilities (Spanggord and Suta 1982; Spanggord et al. 1982a). DNT occurs in samples of TNT waste waters at concentrations of 0.04-48.6 mg/L (2,4-DNT) and 0.06-14.9 mg/L (2,6-DNT). The occurrence of DNT in waste waters from other manufacturing uses (e.g., polyurethane forms) has not been reported. The frequency of detection of DNT in surface waters, as indicated in the STORET database (Staples et al. 1985), is low. Slightly over 1% of the stations reported detectable quantities of DNT, and the median of positive samples was less than 10 $\mu\text{g}/\text{L}$. The presence of DNT was not detected in samples of sediment or biota.

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The water solubility of 2,4- and 2,6-DNT (270 ppm and 180 ppm, respectively) (Mabey et al. 1982) is sufficient to permit transport in surface or groundwaters. Thus, releases to water are very important sources of potential human exposure. Prior to introduction of discharge controls, washings from munitions manufacturing and demilitarization facilities could produce thousands of gallons of contaminated effluent per day at a single facility. Moreover, water contamination may persist. For example, an old effluent lagoon contained detectable quantities of DNT about 12 years after use had been discontinued (Jenkins et al. 1986).

Hashimoto et al. (1982) reported the release of as much as 150 kg/day of DNT isomers in the effluent from a single coastal industrial drain in Dokai Bay, Japan, with a daily average release of 76 kg/day over 7 months. However, DNT concentrations in the bay decreased with distance from the site more rapidly than predicted by simple dilution and tidal action. The salting-out effect of sea water on nitroaromatic compounds was probably responsible for this effect (Hashimoto et al. 1984).

5.2.3 Soil

According to the Toxics Release Inventory, in 1996, there were no reported releases of 2,4-DNT or 2,6-DNT to soil (TR196 1998). The TRI data should be used with caution, however, since only certain types of facilities are required to report. This is not an exhaustive list

2,4-DNT has been identified in soil at 35 of 69 NPL hazardous waste sites, and in sediment at 16 of 69 NPL hazardous waste sites where it was detected in some environmental media (HazDat 1998). 2,6-DNT has been identified in soil at 20 of 53 NPL hazardous waste sites, and in sediment at 9 of 53 NPL hazardous waste sites where it was detected in some environmental media (HazDat 1998).

No information on releases of DNT to soil were located. The extensive use of DNT as an intermediate in the synthesis of toluenediisocyanate and polyurethane foam is not a reported source of releases to soil.

5.3 ENVIRONMENTAL FATE

5.3.1 Transport and Partitioning

The water solubilities of DNT are moderate (Callahan et al. 1979), and the octanol-water partition coefficients are low (Mabey et al. 1982). As a result, there is a potential for transport of DNT by surface or

5. POTENTIAL FOR HUMAN EXPOSURE

groundwater. Spanggord et al. (1980) determined partitioning of 2,4-DNT onto environmental media and found that the soil organic carbon partition coefficient (K_{OC}), the octanol-water partition coefficient ($\log K_{ow}$) and the partition bioconcentration factor (KB) were 364, 2, and 64, respectively. Mabey et al. (1982) calculated sediment-water partitioning coefficients of 45 and 92 for 2,4-DNT and 2,6-DNT, respectively. Depending on the nature of the sediment load, the total concentration of DNT carried in the soil and water column could be high. DNT in buried munitions wastes could potentially be released to groundwater or transported as contaminated soil and sediment.

The expected bioaccumulation of DNT in animal tissues is negligible. The low octanol-water partition coefficients do not indicate a concern for biological lipid accumulation (Trabalka and Garten 1982). Values of K_{ow} lower than 2.5 for nonionized organic chemicals predict bioconcentration solely dependent on the magnitude of exposure. The $\log K_{ow}$ values for 2,4-DNT and 2,6-DNT are 1.98-2.01 and 2.05-2.28, respectively (Callahan et al. 1979; Mabey et al. 1982), indicating that bioaccumulation is not likely to occur.

Direct measurement of plant uptake of DNT has not been made, but plant uptake is predicted to occur based on its low octanol-water partition coefficient. Structural analogy with 1,3-dinitrobenzene and 4-nitrotoluene (McFarlane et al. 1987; Nolt 1988) suggests that 2,4- and 2,6-DNT would be readily taken up by plants. However, plant uptake of related nitroaromatic compounds such as 2,4,6-TNT and its byproduct 4-amino-2,6-DNT has been observed and is inversely proportional to soil organic carbon content (Pennington 1988). The relative concentrations in the plants was root > stem > leaves > seed and food (Cataldo et al. 1989).

The Pre-Biologic Screen (PBS) model for ecotoxicologic effects (Gillett 1983) estimates a score (heavy concern, concern, or no concern) for a compound determined by the octanol-water partition coefficient, the Henry's Law constant, and the half-life in the medium of interest. The score indicates the compound's potential for (a) bioaccumulation and multi-media/multispecies effects, (b) bioaccumulation and long-term effects, (c) persistence and interactions in the water column, including plant uptake and leaching, and (d) direct and indirect effects in the atmosphere (e.g., smog formation, plant fumigation, stratospheric modification). Both 2,4- and 2,6-DNT are of concern or heavy concern only for (c), persistence and interactions in the water column, depending on the value used for half-life. Since the degradation of DNT is so dependent on environmental conditions and the presence of effective microorganisms, the protective view that DNT is of heavy concern for persistence in water, plant uptake, and leaching to groundwater may be warranted. The lack of concern for bioaccumulation, multimedia/multispecies action, and atmospheric action also appears to be justified.

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5.3.2 Transformation and Degradation

5.3.2.1 Air

Based on its rapid photolysis in water, DNT is presumed to be subject to oxidation of its methyl group, decarboxylation, ring oxidation, and/or nitroreduction in air and sunlight. Indirect photolysis must be considered as well. Although the literature implies that these routes lead to destruction of DNT, no experimental measurements were located in the available studies.

5.3.2.2 Water

DNT may be degraded in water through several mechanisms, including photolysis, microbial biodegradation, ozonation and chlorination, and oxidation by strong oxidants such as hydrogen peroxide, ozone, or oxone (Andrews and Osmon 1977; Bausum et al. 1992; Bradley et al. 1995; Freedman et al. 1996; Ho 1986; Noguera and Freedman 1996; Roth and Murphy 1979). Ho (1986) studied photooxidation of 2,4-DNT in aqueous solution in the presence of hydrogen peroxide and suggested that the degradation pathway of 2,4-DNT was, although it includes other intermediates, as follows: 2,4-DNT -- 1,3-dinitrobenzene - hydroxynitrobenzene derivatives + carboxylic acids – CO₂ H₂O, and HNO₃. Oxidation of aqueous DNT with hydrogen peroxide or UV irradiation alone was very slow, and elimination was not complete. Dillert et al. (1995) reported that degradations of DNT and several other nitroaromatics were accelerated in irradiated TiO₂, suspensions and that the degradation rates were dependent on time, solution pH, and light intensity. At given temperature, pH, and photo intensity, degradation rates were shown in the order of 2-nitrotoluene > nitrobenzene > DNT > 1,3-dinitrobenzene > TNT > trinitrobenzene, and the degradations followed first order kinetics. There was very little influence on degradation rates by changes in pH (Dillert et al. 1995; Kumar and Davis 1997).

The photocatalytic oxidation of 2,6-DNT in aqueous suspension of TiO₂ produces ammonium and nitrate ions as the predominant species (Kumar and Davis 1997).

The presence and potential toxicity of DNT in waste water have spurred considerable study of the abiotic and biotic fate of 2,4- and 2,6-DNT. Spanggord et al. (1980) reported that the half-lives of DNT in 3 sunlit natural waters were 3-10 hours, whereas the photolysis half-life in distilled water was 43 hours. Simmons

5. POTENTIAL FOR HUMAN EXPOSURE

and Zepp (1986) found that dissolved or suspended humic substances greatly enhance (10-17 times) indirect photolysis of nitroaromatic compounds with a nitro group ortho to a methyl group. Since the calculated rates of oxidation are less by a factor of about 10^{-5} , photolysis is probably the major route of degradation of DNT in oxygenated waters.

DNT may be also degraded by ozonation and chlorination. Lee and Hunter (1985) reported that both ozone and chlorine produced less than 17% reduction of 2,6-DNT, whereas 2,4-DNT was more vulnerable, yielding about 35% reduction by chlorine and 60% reduction by ozone. Contact time did not appear to have any impact on the reduction rates.

In the absence of sunlight and oxygen, any losses of DNT would apparently be dependent on biodegradation. Spanggord et al. (1980) observed biodegradation in an aerobic environment, with a half-life of less than 1 hour. Under anaerobic conditions, the half-life of 2,6-DNT in non-acclimated sewage is found to be 28 days, with no loss of the compound under aerobic conditions during the same period (Hallas and Alexander 1983). Similarly, Liu et al. (1984) reported no loss of 2,4-DNT in aerobic municipal activated sludge to which benzene was added as a carbon source for bacterial growth, but complete biotransformation of 2,4-DNT within 14 days under anaerobiosis. The intermediates of biotransformation were identified as 2-amino-4-nitrotoluene, 4-amino-2-nitrotoluene, 2-nitroso-4-nitrotoluene, and 4-nitroso-2-nitrotoluene. Parrish (1977) investigated 190 fungal species from 98 genera but found only 5 capable of 2,4-DNT biotransformation. Valli et al. (1992) reported degradation of 2,4-DNT as the sole source of carbon and energy by the lignin-degrading fungus *Phanerochaete chrysosporium* under aerobic conditions, resulting in stoichiometric release of nitrate.

Several additional studies have shown biodegradation of DNT from microorganisms taken from areas that are frequently exposed to DNT (Bausum et al. 1992; Bradley et al. 1995; Freedman et al. 1996). Bausum et al. (1992) found complete degradation of 20 ppm 2,4-DNT and 20 ppm 2,6-DNT in water samples taken downstream a short distance from the Radford Army Ammunition Plant in Radford, Virginia. A lag time was noted prior to the breakdown for both of the two compounds with 2,4-DNT exhibiting the shorter lag time. Microbial enrichment cultures were developed from the collected water samples by exposing the cultures to increasing concentrations of 2,4- and 2,6-DNT. Degradation and visible turbidity in the suspension medium were noted up to a level of 130 ppm. In a separate but related study, degraded DNT was shown to be converted to CO₂ with 2,4-DNT conversion occurring at a greater rate than that of 2,6-DNT; concentrations

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ranged from 0.004 to 10.0 ppm (Bausum et al. 1992). The rate of mineralization to CO₂ was concentration dependant and increased with increasing concentration.

In the Bradley et al. (1995) study, a culture of microorganisms taken from aquifer sediments at an explosivescontaminated site was observed to be capable of removing added 2,4- and 2,6-DNT. The removal rate of 2,4-DNT after 70 days was greater than 99% after application of 100 µM of 2,4-DNT. Removal of 2,6-DNT at the same concentration was less efficient, with 60% of the compound being removed after 70 days. Breakdown products from 2,4-DNT degradation included 4-amino-2-nitrotoluene and 2-amino-4-nitrotoluene. Carbon dioxide was released during the degradation process. Aminonitrotoluene isomers were also detected as breakdown products of a solution of 2,4-DNT and ethanol (Freedman et al. 1996). The Freedman et al. (1996) study exposed an inoculum from a wastewater treatment plant at an ammunition plant to a solution of 2,4-DNT and ethanol and a solution of 2,4-DNT and ether. The concentration of 2,4-DNT at each application was 0.55 mM and the concentrations of ethanol and ether were 600 mg/L and 142 mg/L, respectively. Ethanol and ether were chosen because they are often found in munitions manufacturing wastewater streams along with 2,4-DNT. As stated above, aminonitrotoluene isomers were detected as products of the solution containing 2,4-DNT and ethanol. The degradation of ethanol was believed to be driving a partial reduction of 2,4-DNT before the oxidation of 2,4-DNT took place. In contrast, ether at the applied concentration slowed the rate of 2,4-DNT degradation. Low chemical oxygen demand during the studies suggests that DNT was mineralized to a significant degree.

In a culture using a continuous flow laboratory fermentor under anaerobic conditions with both 2,4-DNT and ethanol as substrates, 2,4-DNT was completely transformed to 2,4-diaminotoluene (DAT) (Cheng et al. 1996). During the biotransformation, two intermediates were formed: 2-amino-4-nitrotoluene and 4-amino-2-nitrotoluene. The products formed from anoxic biotransformation of 2,4-DNT by two denitrifying enrichment cultures with ethanol provided as a primary substrate were characterized in one study (Noguera and Freedman 1997). One culture was developed with inoculum acclimated to DNT, the other with activated sludge that was not routinely exposed to nitroaromatic compounds. The acclimated culture consumed DNT twice as fast as the unacclimated culture, with reduction of DNT to aminonitrtoluenes as the initial pathway. The principal metabolites identified in the acclimated culture were 6-nitroindazole, 2-nitrotoluene, 4 nitrotoluene, as well as products from acetylation at the pm-u position (4-acetamide-2-nitrotoluene and 4 acetamidetoluene). Reduction of aminonitrotoluenes to 2,4-diaminotoluene also occurred, and its subsequent disappearance results in accumulation of significant amount of nonfilterable material in both cultures. The soluble metabolites formed from the unacclimated culture were more hydrophilic. Initial

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characterization of the highly hydrophilic metabolites indicated approximately equal amounts of negatively charged and neutral compounds.

Biotransformation of DNT by a *Pseudomonas aeruginosa* strain, which was isolated from a propellant wastewater treatment plant, was observed under both aerobic and anoxic conditions (Noguera and Freedman 1996). The biotransformation was mainly reductive under both of these conditions. The primary breakdown products were 4-amino-2-nitrotoluene and 2-amino-4-nitrotoluene, with small amounts of 2,4-diaminotoluene also formed. Several DNT metabolites from acetylation of the arylaminos were also identified, including 4-acetamide-2-nitrotoluene, Zacetamide-4-nitrotoluene, 4-acetamide-2-aminotoluene, and 2,4-diacetamidetoluene.

5.3.2.3 Sediment and Soil

Little information was located regarding transformation of DNT in soil. Microorganisms indigenous to surface soils collected at a munitions-contaminated site were reported to transform 2,4- and 2,6-DNT to amino-nitro intermediates within 70 days (Bradley et al. 1994). Another study showed that composting can decrease the concentrations of explosives, such as TNT, in contaminated soil, but neither 2,4- nor 2,6-DNT was detected in the compost (Griest et al. 1993). A study of soil sample handling times indicated that lower temperatures retard the breakdown of 2,4-DNT (Grant et al. 1995). 2,4-DNT was observed to be more stable than TNT in contaminated soils (Grant et al. 1995).

5.4 LEVELS MONITORED OR ESTIMATED IN THE ENVIRONMENT

Reliable evaluation of the potential for human exposure to 2,4-DNT and 2,6-DNT depends in part on the reliability of supporting analytical data from environmental samples and biological specimens. In reviewing data on 2,4-DNT and 2,6-DNT levels monitored or estimated in the environment, it should also be noted that the amount of chemical identified analytically is not necessarily equivalent to the amount that is bioavailable.

5.4.1 Air

Measurements of DNT in ambient air were not located. In an occupational environment, Ahrenholz (1980) measured breathing zone air concentrations of Tg-DNT that ranged from undetected to 23 $\mu\text{g}/\text{m}^3$ (timeweighted average [TWA]). Tg-DNT concentrations in the area air samples ranged from undetected to 420

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$\mu\text{g}/\text{m}^3$ (TWA). Ahrenholz and Meyer (1982) reported that area air samples in a manufacturing facility contained TWA concentrations of Tg-DNT that ranged from undetected to $890 \mu\text{g}/\text{m}^3$. Levine et al. (1985b) also measured personal air samples of 2,4-DNT at $10\text{--}440 \mu\text{g}/\text{m}^3$ and 2,4- and 2,6-DNT at $50\text{--}590 \mu\text{g}/\text{m}^3$ in the workplace.

5.4.2 Water

As priority pollutants, 2,4- and 2,6-DNT are monitored routinely in U.S. waters, but both the number of positive samples (1.1 - 1.6%) and the mean value ($<10 \mu\text{g}/\text{L}$) of those samples indicate that release of the compounds to water is neither widespread nor very great. No residues of 2,4- or 2,6-DNT were reported in sediments or biota (Staples et al. 1985). Residues of 2,4-DNT were detected in the groundwater of 1.5% of 800-900 hazardous waste sites (CLPSD 1988). The geometric mean concentration in groundwater was $58 \mu\text{g}/\text{L}$. In surface waters, 2,4-DNT was detected at 1.0% of hazardous waste sites at a geometric mean concentration of $104 \mu\text{g}/\text{L}$. In surface waters of the river Elbe in Germany, concentrations of 2,4-DNT ranged from 0.1 to $1.3 \mu\text{g}/\text{L}$, while concentrations of 2,6-DNT ranged from 0.08 to $0.5 \mu\text{g}/\text{L}$ (Feltes et al. 1990). Sohr et al. (1995) reported 2,4-DNT and 2,6-DNT concentrations of $0.7 \mu\text{g}/\text{L}$ and $3.1 \mu\text{g}/\text{L}$, respectively, at contaminated warfare sites in Germany. Other less contaminated sites contained 28 ng/L 2,4-DNT and 19 ng/L 2,6-DNT (Sohr et al. 1995).

Residues of 2,6-DNT were detected in the groundwater of 1.0% of waste sites at a geometric mean concentration of $18 \mu\text{g}/\text{L}$. 2,6-DNT was detected in the surface water of only 0.3% of the waste sites (CLPSD 1988).

5.4.3 Sediment and Soil

Hoke et al. (1993) reported that only low concentrations of 2,4-DNT and 2,6-DNT were detected in sediment of the Great Calumet River-Indian Harbor. Concentrations of 2,4-DNT in sediments ranged from the detection limit of $0.01 \mu\text{g}/\text{L}$ to $0.07 \mu\text{g}/\text{L}$. 2,4-DNT concentrations in sediment pore water ranged from $0.1 \mu\text{g}/\text{L}$ to $1.7 \mu\text{g}/\text{L}$. 2,6-DNT was not detected in sediment samples (LOD = $0.01 \mu\text{g}/\text{L}$) and subsequently was not analyzed for in sediment pore water.

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Concentrations ranging from less than 0.1 mg/kg (detection limit) to 117 mg/kg of 2,4-DNT were found at the Joliet Army Ammunition Plant, in Joliet, IL, an NPL site. 2,6-DNT was detected on this site at concentrations ranging from less than 0.1 mg/kg to 8 mg/kg (Simini et al. 1995).

5.4.4 Other Environmental Media

Neither 2,4- nor 2,6-DNT were detected in samples of fish obtained from Lake Michigan tributaries and Grand Traverse Bay (Camanzo et al. 1987). DNT was not detected in fish from Great Lakes harbors and tributaries in Ohio and Wisconsin (DeVault 1985).

5.5 GENERAL POPULATION AND OCCUPATIONAL EXPOSURE

Sources of exposure to DNT for the general population include processing facilities that manufacture or process DNT, as well as hazardous waste sites that release this chemical. There are 3 large processing facilities that release a total of 3,080 kg of 2,4- and 2,6-DNT to the environment annually (TR196 1998), and 2,4- and 2,6-DNT has been found in at least 69 and 53 waste sites, respectively (HazDat 1998). The general population may be exposed to DNT via inhalation, dermal contact, and incidental ingestion pathways. Occupational exposure to DNT may occur from its use in the manufacture of toluene diisocyanate, in the production of explosives, in the manufacture of azo dye intermediates, and in organic synthesis in the preparation of toluidines and dyes (IARC 1996). Exposure may also occur at facilities that store or dispose the substance. Occupational exposure pathways will also involve inhalation, dermal contact, and incidental ingestion.

Studies on occupational exposure to DNT are limited. Levine et al. (1985b) evaluated the 7-hour Time-Weighted Average (TWA) personal exposure of workers to DNT technical grade (Tg-DNT) and measured urinary metabolites of DNT at a DNT manufacturing plant. Breathing zone exposure level of production unit operators to both 2,4- and 2,6-DNT averaged 0.26 mg/m³. Air exposure concentrations of loaders, who load storage tanks, collect samples, and perform cleaning tasks, averaged 0.32 mg/m³. Exposure of maintenance mechanics averaged 0.12 mg/m³, and the exposure of acid-stripper operators was 0.06 mg/m³. The highest personal air monitoring concentrations and levels of urinary metabolites were found to be for loaders, followed by process operators. The levels of urinary metabolites of DNT in loaders and operators exceeded those that would have resulted from the inhaled concentrations, although the workers wore gloves for operations that might have led to dermal exposure. In another study of occupational absorption in an

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explosives manufacturing facility (Woollen et al. 1985), personal airborne exposure to DNT ranged from undetectable to 0.1 mg/m³. Area samples collected near dusty parts of the process ranged from 0.02 to 2.68 mg/m³. However, air concentrations could not account for the observed excretion levels of the metabolite 2,4-dinitrobenzoic acid, indicating probable dermal uptake.

Two major studies of occupational exposure to DNT (Levine et al. 1985b; Woollen et al. 1985) reported the following: maximum excretion of metabolites occurred at or after the end of a working shift; excretion was practically complete by the start of the next shift; and rates of 2,4-dinitrobenzoic acid excretion in the urine made a plausible marker of exposure, provided that sampling patterns were adequate to compensate for the wide variation over time, both between individuals and for the same person. In both studies, difficulties in detecting metabolites and frequent occurrence of samples with values near the limits of detection hampered interpretation of actual exposure and disposition. Higher exposures in the study by Levine et al. (1985b) permitted more extensive analysis. The results of both studies indicated that dermal contact and inadvertent ingestion were contributing routes of exposure for male and female workers.

The Occupational Safety and Health Administration (OSHA) established an 8-hour Time-Weighted Average (TWA) Permissible Exposure Limit (PEL) for dinitrotoluene as 1.5 mg/m³, with skin designation to indicate the potential significant contribution to the overall exposure by the cutaneous route (OSHA 1998). The American Conference of Governmental Industrial Hygienist (ACGIH)’s Threshold Limit Value (TLV) for dinitrotoluene is 0.2 mg/m³, with skin notation (ACGIH 1998). TLV is the time-weighted average concentration for a conventional 8-hour workday and a 40-hour workweek to which it is believed that nearly all workers may be repeatedly exposed without adverse effect. The National Institute of Occupational Safety and Health (NIOSH) determined the Recommended Exposure Limit (REL) for dinitrotoluene as 1.5 mg/m³, with skin designation (NIOSH 1997).

5.6 EXPOSURES OF CHILDREN

This section focuses on exposures from conception to maturity at 18 years in humans and briefly considers potential pre-conception exposure to germ cells. Differences from adults in susceptibility to hazardous substances are discussed in 2.6 Children’s Susceptibility.

Children are not small adults. A child’s exposure may differ from an adult’s exposure in many ways. Children drink more fluids, eat more food, and breathe more air per kilogram of body weight, and have a

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larger skin surface in proportion to their body volume. A child's diet often differs from that of adults. The developing human's source of nutrition changes with age: from placental nourishment to breast milk or formula to the diet of older children who eat more of certain types of foods than adults. A child's behavior and lifestyle also influence exposure. Children crawl on the floor; they put things in their mouths; they may ingest inappropriate things such as dirt or paint chips; they spend more time outdoors. Children also are closer to the ground, and they do not have the judgement of adults in avoiding hazards (NRC 1993).

No studies are available that monitor the level of exposure of children to DNT. No measurements have been made of DNT or its metabolite levels in amniotic fluid, meconium, cord blood, or neonatal blood to test for prenatal exposure, nor have measurements been made of DNT or metabolite levels in breast milk. However, because of the low octanol-water partition coefficient of DNT and excretion in the urine, it is not expected to accumulate in maternal tissues.

Although DNT can degrade in water by several mechanisms, it has the potential to be transported in surface water or groundwater due to its moderate water solubility. Therefore, children playing in DNT contaminated surface water have the potential to be more exposed than adults, both because of this behavior and because of their larger skin surface area in proportion to their body weight for dermal absorption. Also, children drinking well or municipal water contaminated with DNT might be exposed to more of the chemical than adults would be due to the fact that children drink more fluids per kilogram of body weight than adults. Significant dietary exposure is unlikely as DNT is not expected to accumulate in animal tissues. However, ingestion of vegetables and crops grown in DNT contaminated areas could be a source of exposure.

There were no studies that examine potential exposure of children from parents' work clothes, skin, hair, tools, or other objects removed from the workplace. No information is available concerning exposure from consumer products because DNT is used mainly for military and industrial purposes.

Although DNT is genotoxic in *in vivo* test systems, it is found to be negative in dominant lethal mutations (Ellis et al. 1979) and spermatocyte DNA repair (Working and Butterworth 1984). There is no evidence that exposure of parental germ cells to the active form of DNT could plausibly occur since DNT does not accumulate in tissue.

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5.7 POPULATIONS WITH POTENTIALLY HIGH EXPOSURES

In addition to individuals who are occupationally exposed to 2,4-DNT and 2,6-DNT (see Section 5.5), there are several groups within the general population that have potentially high exposures (higher than background levels) to 2,4-DNT and 2,6-DNT. These populations include individuals living in proximity to sites where 2,4-DNT and 2,6-DNT were produced or sites where 2,4-DNT and 2,6-DNT were disposed, and individuals living near one of the NPL hazardous waste sites where 2,4-DNT and 2,6-DNT have been detected in some environmental media (HazDat 1998).

Based on the available information, it appears that the highly-exposed populations would be workers exposed in manufacturing facilities.

Members of the general population are likely to be exposed only if they are near a local source of contamination, such as an industrial discharge or an abandoned waste site. 2,4-DNT and 2,6-DNT do not appear to be widespread in the environment, and they were not frequently detected at hazardous waste sites.

5.8 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of 2,4- and 2,6-DNT is available. Where adequate information is not available, ATSDR, in conjunction with the NTP, is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of 2,4- and 2,6-DNT.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

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5.8.1 Identification of Data Needs

Physical and Chemical Properties. Information regarding the physical and chemical properties of a chemical is essential for estimating the partitioning of the chemical in the environment. Information on the physical and chemical properties of DNT is presented in Chapter 3 and the data appear to be adequate (HSDB 1998; Lide 1993). The isomers of DNT have many similar traits, including identical molecular weights), but 2,4-DNT has higher melting and boiling points than 2,6-DNT and a greater solubility in water (HSDB 1998). DNT is generally produced as a technical-grade mixture comprised of 95% 2,4- and 2,6-DNT, and 5% other substances. The other substances are predominantly isomers of DNT not discussed in this profile.

Production, Import/Export, Use, Release, and Disposal. According to the Emergency Planning and Community Right-to-Know Act of 1986,42 U.S.C. Section 11023, industries are required to submit chemical release and off-site transfer information to the EPA. The Toxics Release Inventory (TRI), which contains this information for 1996, became available in May of 1998. This database is updated yearly and should provide a list of industrial production facilities and emissions. The TRI reported that DNT was produced at 3 sites within the United States (TR196 1998).

USITC statistics on synthetic organic chemical production (USITC 1987) do not describe DNT production. Uses of DNT appear to be well characterized (CMR 1983; HSDB 1998). The most commercially important use of DNT is as a chemical intermediate in the production of toluene diisocyanate, a precursor to polyurethane polymers (HSDB 1998). It has been estimated that 99% of all DNT produced is used for this purpose (CMR 1983). DNT is recognized as a potentially hazardous chemical and is subject to a variety of regulations (see Chapter 7), but disposal practices and restrictions are not adequately documented.

Environmental Fate. The low octanol-water partition coefficients of the DNT isomers predict that DNT released to the environment would not bioaccumulate and would be weakly bound to soil organic matter. The relatively low volatility and high solubility of DNT indicate that it will tend to remain in water for long periods of time unless acted upon by light, oxygen, or biota, creating the potential for transportation to groundwater or surface water (Jenkins et al. 1986). DNT has been found in wastewater and groundwater in and around munitions sites (Jenkins et al. 1986; Spanggord and Suta 1982). The occurrence of DNT in wastewater from other manufacturing uses such as polyurethane forms has not been reported.

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Given the importance of information about the behavior of DNT in the water column, and the extensive range of available information relative to that topic (Gillett 1983; Hashimoto et al. 1982, 1984; Jenkins et al. 1986; Mabey et al. 1982; Spanggord et al. 1980), the absence of substantive information about DNT releases to, or fate in, soils and air is less troublesome than it might be for many chemicals. Data on the persistence of DNT in soil, the vadose zone (the unsaturated zone lying between the ground level and the top of the groundwater), and groundwater are needed, as well as measured rates of plant uptake and metabolism. Because of the structurally specific nature of biotransformations, more information on the fate of DNT metabolites would be welcome.

Bioavailability from Environmental Media. No information is currently available that describes the bioavailability of DNT in environmental media or in food. Data on bioavailability of soil/sediment residues would be helpful. Neither 2,4- nor 2,6-DNT were detected in samples of fish obtained from Lake Michigan tributaries and Grand Traverse Bay (Camanzo et al. 1987). DNT was not detected in fish from Great Lakes harbors and tributaries in Ohio and Wisconsin (DeVault 1985).

Food Chain Bioaccumulation. Limited information indicates that DNT is not widely distributed in the environment. Residues of DNT have not been observed in fish samples and have a low frequency of detection in water samples. These data indicate that bioaccumulation may not be an area of concern (Callahan et al. 1979; Mabey et al. 1982; Trabalka and Garten 1982). The expected bioaccumulation of DNT in animal tissues is negligible. The log K_{ow} values for 2,4- and 2,6-DNT are 1.98-2.01 and 2.05-2.28, respectively (Callahan et al. 1979; Mabey et al. 1982), indicating that bioaccumulation is not likely to occur. The bioavailability and potential toxicity of soil- and sediment-bound products are unknown. Degradation of DNT forms a variety of products, about which very little is known. Additional information would help to confirm or refute indications of low potential for bioaccumulation of DNT in foods.

Exposure Levels in Environmental Media. Reliable monitoring data for the levels of 2,4- and 2,6-DNT in contaminated media at hazardous waste sites are needed so that the information obtained on levels of 2,4- and 2,6-DNT in the environment can be used in combination with the known body burdens of 2,4- and 2,6-DNT to assess the potential risk of adverse health effects in populations living in the vicinity of hazardous waste sites.

The sparse information base suggests that widespread contamination by DNT has not occurred (Staples et al. 1985). Monitoring for priority pollutants in wastewater, drinking water, and biota has failed to detect

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residues in sediments or biota (Staples et al. 1985). DNT was detected at very low frequencies in drinking water, the number of positive samples ranging from 1.1 - 1.6% and the mean value of those samples reported at less than 10 µg/L. Analyses of wastewaters indicate that local contamination may occur (Feltes et al. 1990; Shackelford and Keith 1976; Spanggord and Suta 1982; Spanggord et al. 1982a). Residues of 2,4-DNT were detected in the groundwater of 1.5% of 800-900 hazardous waste sites (CLPSD 1988). The geometric mean concentration in groundwater was 58 µg/L. In surface waters, 2,4-DNT was detected at 1.0% of hazardous waste sites at a geometric mean concentration of 104 g/L.

Exposure Levels in Humans. This information is necessary for assessing the need to conduct health studies on these populations. No studies of exposure of the general population were found, and the occupational studies (Levine et al. 1985b; Woollen et al. 1985) are inadequate to ascertain "background" or nonoccupational exposure. Based on available information, the highly-exposed populations are those workers exposed in manufacturing facilities. Members of the general population are likely to be exposed only in that they are near a local source of contamination. Toxicokinetic data on occupationally- and environmentally-exposed humans will be helpful. Measurements of DNT and its metabolite levels in blood and urine will be useful to provide an estimate of internal dose of exposure.

Exposures of Children. No exposure and body burden studies have been conducted on children; consequently, it is not known if children differ from adults in their weight-adjusted intake of DNT, or if unique exposure pathways for children exist. Since DNT is not a widespread environmental contaminant, there are only two likely potential sources of exposure for children. Children living near a DNT contaminated site might be exposed if DNT has moved offsite in contaminated environmental media. If such a situation were identified, further site specific studies of children's exposure could be conducted. Children whose parents work in manufacturing facilities that produce or use DNT and are occupationally exposed to significant quantities of DNT might potentially be exposed to DNT transported home on their parents' work clothes, skin, hair, tools, or other objects removed from the workplace. If such a significant occupational exposure setting were identified, they might be the subject of a take-home exposure study.

Exposure Registries. No exposure registries for DNT (2,4- or 2,6-DNT) were located. Neither of these substances is currently one of the compounds for which a subregistry has been established in the National Exposure Registry. The substance will be considered in the future when chemical selection is made for subregistries to be established. The information that is amassed in the National Exposure Registry facilitates

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the epidemiological research needed to assess adverse health outcomes that may be related to exposure to this substance.

The development of an exposure registry would provide valuable data on exposure levels and frequency. In addition to providing information on exposure levels and duration, a registry would be useful in identifying sources of exposure such as hazardous waste sites and manufacturing and use facilities. Knowledge about exposure levels and sources would be valuable in developing strategies to control unnecessary sources and these exposures. The ability to correlate sources and exposure levels with health effects would be useful in identifying disease conditions that may result from exposure to the chemical.

5.8.2 Ongoing Studies

Photochemical reactions of 2,4-DNT in aqueous solutions containing cationic surfactant, nonionic surfactant, a hydrogen donor, or a base are being investigated by C.A. Diehl et al. (1995) at Purdue University.

A project to develop and apply biomarkers of exposure associated with 2,6-DNT is being conducted by Dr. Steve Rappaport at University of North Carolina at Chapel Hill (FEDRIP 1998).

A project to develop an anaerobic biodegradative process for the biodegradation of DNT in aqueous streams is being conducted by D. Munnecke at Environmental Biotechnologies, Inc. in Montara, CA (FEDRIP 1997).

6. ANALYTICAL METHODS

The purpose of this chapter is to describe the analytical methods that are available for detecting, and/or measuring, and/or monitoring 2,4-DNT and 2,6-DNT, its metabolites, and other biomarkers of exposure and effect to 2,4-DNT and 2,6-DNT. The intent is not to provide an exhaustive list of analytical methods. Rather, the intention is to identify well-established methods that are used as the standard methods of analysis. Many of the analytical methods used for environmental samples are the methods approved by federal agencies and organizations such as EPA and the National Institute for Occupational Safety and Health (NIOSH). Other methods presented in this chapter are those that are approved by groups such as the Association of Official Analytical Chemists (AOAC) and the American Public Health Association (APHA). Additionally, analytical methods are included that modify previously used methods to obtain lower detection limits, and/or to improve accuracy and precision.

6.1 BIOLOGICAL SAMPLES

The need to determine DNT in biological materials could arise from occupational exposure in the manufacture and processing of 2,4,6-TNT and from exposure to waste water and waste disposal sites associated with TNT manufacture. It has been noted (Jenkins et al. 1986) that “one of the Army’s most serious water pollution problems is the disposal of wash waters used to clean equipment and interior surfaces at munitions manufacturing and demilitarization facilities.” The same reference mentions the generation of large quantities of waste water from these facilities.

Although there are numerous occupational monitoring studies, a limited number of methods regarding the determination of DNT in biological samples is available in the literature. DNT has been determined in ocean floor fauna using thin layer chromatography (TLC) (Hoffsommer et al. 1972). Procedures have been described for the examination of swabs for traces of explosives, including 2,6-DNT using high-performance liquid chromatography (HPLC) with electrochemical detection at a pendent drop electrode (Lloyd 1983a). These techniques can be applied to biological materials such as skin surfaces exposed to explosives. DNT and its metabolites were determined in blood and urine by gas chromatography (GC) techniques (Turner et al. 1985; Woollen et al. 1985) and in urine by TLC (Woollen et al. 1985). Qualitative determination of DNT and its metabolites can also be performed after reduction to primary arylamines and subsequent coupling of diazo compounds to produce a colored complex (Smith et al. 1995).

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Dichloromethane is the solvent of choice for extracting DNT from water samples (EPA 1982a) and from wastes (EPA 1986a). Reversed-phase high-performance liquid chromatography (RP-HPLC) is attractive for the determination of DNT in waste water because it enables direct analysis of aqueous samples (Jenkins et al. 1986). A medium similar to the mobile phase used in this HPLC separation, i.e., 50/38/12 (v/v/v) water/methanol/acetonitrile, should be suitable for extracting DNT from low-lipid biological samples and for subsequent HPLC determination after sample cleanup.

Methods for the determination of the DNT in biological samples are given in Table 6-1.

6.2 ENVIRONMENTAL SAMPLES

The basic method for collecting DNT from the ambient atmosphere is adsorption on a solid phase, such as granular adsorbents (silica gel), filters, and impingers, followed by removal with solvents such as chloroform. Bubbler collectors can also be used for direct collection of analyte in a non-volatile solvent such as ethylene glycol. New instrumentation for the detection of 2,4-DNT and other explosives has recently been developed that will accept both air and surface particulate samples (Nacson et al. 1994). The instruments consist of capillary GC columns terminating in an electron-capture detector; the detection limit for 2,4-DNT is 20 ppt (Nacson et al. 1994). Also available is a portable version that is useful for a wide variety of applications, such as security checks, mail or passports, or in high-risk facilities (Nacson et al. 1994).

DNT is most commonly extracted with dichloromethane from water samples (EPA 1982a) and from wastes (EPA 1986a). A continuous countercurrent liquid-liquid extraction method is useful in extracting DNT from surface water samples (Deroux et al. 1996). The advantage of this method is that it is capable of extractions from large sample volumes and unfiltered natural water samples (Deroux et al. 1996). Supercritical fluid extraction (SFE) has been used with GC to reduce the preparation time of DNT in solids, such as soil (Francis et al. 1995). A sonic extraction-liquid chromatographic method has also been used for detection of 2,4-DNT in soils (Bauer et al. 1990; Griest et al. 1993). A simple screening method has been developed for the detection of 2,4-DNT in field soil samples that utilizes the spectrophotometer for identification by colorimetrics after an initial reaction of the extract with potassium hydroxide and sodium sulfite (Jenkins and Walsh 1992).

The analysis of DNT is normally done by GC with a variety of detectors, including flame ionization detector (FID), electron capture detector (ECD), Hall electrolytic conductivity detector (HECD), thermionic specific

TABLE 6-1. Analytical Methods for Determining Dinitrotoluene in Biological Samples

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Urine containing dinitrotoluene and metabolites	Hydrolysis of metabolites, extraction, derivatization	GC/MS	0.1 mg/L	NR	Turner et al. 1985
Urine containing dinitrotoluene and metabolites	Zinc-catalyzed reduction of DNT with hydrochloric acid to primary arylamines; diazotize and couple with <i>N</i> -(1-naphthyl)ethylene diamine to produce complex	SP	100 ng/mL	NR	Smith et al. 1995
Urine containing metabolites	Extraction with ethyl acetate	GC	0.1 mg/L	NR	Woollen et al. 1985
Urine	Extraction with ethyl acetate	TLC	0.1 mg/L ^a	NR	Woollen et al. 1985
Tissue/fluids	Extract with acetonitrile and filter with nylon membrane.	HPLC UV/VIS	0.4-0.5 ppm	-	Caton and Griest 1996
Plant tissue	Extract with sonication. Clean up on flonsil; alumina (liq. chrom.). Inject.	Reverse-phase LC UV/VIS	0.02-10 mg/mL	-	Larson 1998
Skin	Swab with ethanol	HPLC/ED	5.6 ng/mL ^a	97 (2,6-DNT); 93 (2,4-DNT)	Lloyd 1983a
Blood	Extraction with toluene	GC	0.00001 mg/mL ^a	NR	Woollen et al. 1985
Ocean floor fauna	NR	TLC	NR		Hoffsommer et al. 1972

TABLE 6-1. Analytical Methods for Determining Dinitrotoluene in Biological Samples (continued)

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Skin	Swab with ethanol	HPLC/ED	5.6 ng/mL ^a	97 (2,6-DNT); 93 (2,4-DNT)	Lloyd 1983a
Skin	Swab surface and insert into outlet port	GC/ECD	20 ppt (2,4-DNT)		Nacson et al. 1994

^aLowest detected concentration

ECD = electron capture detection; ED = electrochemical detection; GC = gas chromatography; HPLC = high-performance liquid chromatography; MS = mass spectrometry; NR = not reported; SP = Spectrophotometry; TLC = thin layer chromatography

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detector (TSD), fourier transform infrared (FT-IR), or mass spectrometry (MS). It has been noted (EPA 1986b) that 2,4-DNT is “subject to erratic (gas) chromatographic behavior.” When mass spectrometry is used to analyze water samples for DNT, electron impact (EI) is preferentially used because many structurespecific fragments will be formed, which can be used for identification of isomers (Feltes et al. 1990). To improve the accuracy of mass spectroscopic techniques in the identification of pollutants in aqueous and solid matrices, EPA has developed the method of isotope dilution (EPA 1989). Isotope dilution employs stable, isotopically labeled analogs of both 2,4- and 2,6-DNT to be used as internal standards in GUMS analysis (EPA 1989). Negative-ion chemical ionization has been shown to have a higher sensitivity and selectivity than EI, however, and should be used when determining traces of nitroaromatic compounds in complex aqueous mixtures (Feltes et al. 1990).

A sensitive and selective technique that has been used to identify trace amounts of 2,4-DNT and other explosive vapors is negative ion mobility spectrometry (Clark et al. 1995). This technique makes use of a tunable laser ionization source to produce characteristic negative ions, which can be used to identify the chemicals present (Clark et al. 1995). TLC and high-performance thin layer chromatography (HPTLC) have also been used to identify and quantify 2,4- and 2,6-DNT in soil and water samples from contaminated waste sites (Griest et al. 1993; Sohr et al. 1995; Steuckart et al. 1994). GC analysis is difficult because of the large amounts of humic acids present which cause overlap of matrix signals without cleanup; therefore, HPTLC can be a more advantageous method (Steuckart et al. 1994). Cleanup is not necessary with HPTLC, except in the analysis of soil samples (Steuckart et al. 1994).

A sensitive method for the analysis of DNT in drinking water has been developed using wide-bore fused silica capillary column GC with an ECD (Hable et al. 1991). The detection limits of this method are 0.04 $\mu\text{g/L}$ for 2,4-DNT and 0.003 $\mu\text{g/L}$ for 2,6-DNT; these detection limits are sensitive enough to meet the suggested requirements for EPA health advisories and water quality criteria.

For the determination of 2,4-DNT in munitions manufacture waste water, RP-HPLC was chosen by Jenkins et al. (1986) because it enables direct analysis of samples in aqueous solution without prior extraction, attains adequate detection limits without preconcentration, and avoids problems with analyte thermal instability. The detection limit for 2,4-DNT was 10 $\mu\text{g/L}$ with a standard deviation of 3.4 $\mu\text{g/L}$ for concentrations up to 250 $\mu\text{g/L}$. HPLC with photodiode array detection was used by Bouvier et al. (1995) to separate and quantitate 2,4- and 2,6-DNT in water samples. Recoveries ranged from 95 to 100%, and minimum detection limits were 0.041- 0.160 $\mu\text{g/L}$ (Bouvier et al. 1995). A convenient method for analysis of 2,4- and 2,6-DNT in

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contaminated soils used high performance liquid chromatography with minimal sample preparation (Preslan et al. 1993).

Methods for the determination of DNT in environmental samples are summarized in Table 6-2.

6.3 ADEQUACY OF THE DATABASE

Section 104(I)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of 2,4-DNT and 2,6-DNT is available. Where adequate information is not available, ATSDR, in conjunction with the NTP, is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of 2,4-DNT and 2,6-DNT.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

6.3.1 Identification of Data Needs

Methods for Determining Biomarkers of Exposure and Effect. The available methods for the determination of DNT and its metabolites in biological samples are inadequate. Although one method exists for determination of DNT and its metabolites in urine (Smith et al. 1995; Turner et al. 1985; Woolen et al. 1985), blood (Woolen et al. 1985), and skin (Lloyd 1983a; Nacson et al. 1994), there is a need for modern validated standard methods of analysis for such data in plant and animal tissues and exudates. Methods do exist for water and waste water (EPA 1982a, 1982b) and for solid wastes (EPA 1986a, 1986b, 1986c). The need also exists for good methods to determine DNT biomarkers in biological materials. The determination of this compound in plant and animal tissues and exudates would be useful to help determine exposure.

TABLE 6-2. Analytical Methods for Determining Dinitrotoluene in Environmental Samples

Sample matrix	Preparation method ^a	Analytical method ^a	Sample detection limit ^a	Percent recovery	Reference
Air	Ethylene glycol bubbler, filter, impinger	HPLC/UV	0.9 mg/m ³		NIOSH 1977
Air	Silica gel, desorb with chloroform	GC	0.1 mg/m ³	70-84	Hunt et al. 1980
Air	Collect sample and insert into analyzer	GC/ECD	20 ppt (2,4-DNT)		Nacson et al. 1994
Water	Extraction with dichloromethane	GC/ECD or TEA	ECD: 3.8×10^{-14} g/s (2,4-DNT); TEA: 1.77×10^{-11} g/s (2,4-DNT)		Feltes et al. 1990
Water	Extraction with dichloromethane	GC/MS (electron impact-full scan)	47 pg (2,6-DNT)		Feltes et al. 1990
Water	Extraction with dichloromethane; add methanol	GC/MS	5 ppb		Yinon 1996
Water	Adjust pH of spiking solution to >11 with NaOH; extraction with dichloromethane; add anhydrous sodium sulfate; filter; rotary evaporate	Liquid-liquid extraction with GC/MS	0.8 µg/L (2,4-DNT); 1.4 µg/L (2,6-DNT)	96 (2,4-DNT); 100 (2,6-DNT)	Yook et al. 1994
Water	Extraction with acetonitrile	HPLC/PDA	0.04-0.07 µg/L (2,4-DNT); 0.08-0.16 µg/L (2,6-DNT)	97-100 (2,4-DNT); 95-99 (2,6-DNT)	Bouvier et al. 1995

TABLE 6-2. Analytical Methods for Determining Dinitrotoluene in Environmental Samples (continued)

Sample matrix	Preparation method ^a	Analytical method ^a	Sample detection limit ^a	Percent recovery	Reference
Water	Counter current liquid-liquid extraction method	GC/MS (EI and PICI)	2 ng/L	89	Deroux et al. 1996
Water	Adjust to pH 12 and pump into column. Remove organic phase. Add to re-tirnate and adjust to pH 2 before pumping into column.	GC/MS	2 ng/L	89	Deroux et al. 1996
Water	Spike H ₂ O sample with standards. Conduct solid-phase extraction (SPE). Elute and dry under nitrogen.	TLC (254 nm)	20 ng (scanner) 40 ng (visually)	diol-119-115.3 RP-18-100.6-102	Kessel and Hauch 1996
Aqueous	Mix sample with sodium chloride until salt dissolves. Concentrate extracts. Combine with 5.0 g/L CaCl ₂ solution and inject.	HPLC (254 nm)	0.13 µg/L	—	Weisberg and Ellickson 1998 (modification to EPA Method 8330)
Drinking water	Extraction with 0.5 mL toluene; rotate 30 minutes at 15 RPM	GC/ECD	0.003 µg/L (2,6-DNT); 0.04 µg/L (2,4-DNT)	93–103 (2,6-DNT); 93–96 (2,4-DNT)	Hable et al. 1991
Drinking water	Elute initially with ethyl acetate, then with dichloromethane	GC/MS			Munch et al. 1993

TABLE 6-2. Analytical Methods for Determining Dinitrotoluene in Environmental Samples (continued)

Sample matrix	Preparation method ^a	Analytical method ^a	Sample detection limit ^a	Percent recovery	Reference
Groundwater and surface water	Extraction with dichloromethane; acidify water sample with HCl and extract with isobutyl methyl ketone; concentrate in rotary evaporator; dissolve residues in dichloromethane	TLC	10–20 ng/spot	—	Sohr et al. 1995
Groundwater	Extraction with dichloromethane	HPTLC-AMD	20 ng (2,4- and 2,6-DNT)	—	Steuckart et al. 1994
Groundwater	Expose three samples to different amounts of sunlight. Inject sample, loop.	HPLC/UV (254 nm)	0.01–0.1 µg/L	—	Spiegel and Welsch 1997
Groundwater	Extract with dichloromethane and dry over sodium sulfate. Extract with dichloromethane and dry over sodium sulfate. Add 1 mL acetonitrile.	HPLC NMR	1–10 µgm/L	—	Preiß et al. 1996
Waste water (for 2,4-DNT)	Diluted directly with methanol and acetonitrile	HPLC/UV	4.6 µg/L	NR	Jenkins et al. 1986
Waste water	Extraction with dichloromethane	GC/IDMS	10 µg/L	10 (2,4-DNT); 17 (2,6-DNT)	EPA 1980a

TABLE 6-2. Analytical Methods for Determining Dinitrotoluene in Environmental Samples (continued)

Sample matrix	Preparation method ^a	Analytical method ^a	Sample detection limit ^a	Percent recovery	Reference
Waste water	Extraction with dichloromethane, exchange to hexane	GC/ECD	NR		EPA 1982a
Waste water	Extraction with dichloromethane; EPA Method 8090	GC/MS	5.7 µg/L (2,4-DNT); 1.9 µg/L (2,6-DNT)		EPA 1986a
Waste water	Extraction with hexane	GC/ECD	25.0×10^{-5} µg (2,4-DNT); 62.5×10^{-6} µg (2,6-DNT) ^b	80 (2,4-DNT); 26 (2,6-DNT)	Hartley et al. 1981
Non-water miscible waste	Extraction with dichloromethane; EPA Method 8090	GC/ECD	2,000 µg/L (2,4-DNT); 1,000 µg/L (2,6-DNT)		EPA 1986a
Vapor	Solid samples placed in gas flow system	Negative ion mobility spectrometry (laser ionization source)	NR	NR	Clark et al. 1995
Biosludge	Extraction with sulfuric acid and dichloromethane	GC/TEA	0.05 mg/L	84	Phillips et al. 1983
Soil	Supercritical fluid extraction (SFE) with neat CO ₂ and CO ₂ with organic modifiers	GC/TDM	2.6 ppb		Francis et al. 1995

TABLE 6-2. Analytical Methods for Determining Dinitrotoluene in Environmental Samples (continued)

Sample matrix	Preparation method ^a	Analytical method ^a	Sample detection limit ^a	Percent recovery	Reference
Soil	Extraction with acetonitrile in ultrasonic bath; flocculate supernatant with CaCl_2 ; filter	SE/LC		95-97	Bauer et al. 1990
Soil	Dilution directly with methanol	HPLC/PDA	40-80 $\mu\text{g}/\text{mL}$		Emmrich et al. 1993
Soil (2,4-DNT)	Extraction with acetone; filter supernatant; react with potassium hydroxide and sodium sulfite	Spectrophotometry	2 $\mu\text{g}/\text{g}$		Jenkins and Walsh 1992
Soil	Grind soil; extraction with acetone in ultrasonic bath; centrifuge; add 5 mL toluene and remove acetone; dry toluene extract over anhydrous sodium sulfate	HPTLC/AMD	20 ng (2,4- and 2,6-DNT)		Steuckart et al. 1994
Soil	Extract with toluene, acetonitrile, methanol and collect in solid-liquid trap.	SFE/HPLC GC	—	100.6-101.5	Deuster et al. 1996
Soil, sediment, solid waste	Extraction	GC/MS	660 $\mu\text{g}/\text{kg}$		EPA 1986b
Soil, sediment, solid waste	Extraction with dichloromethane	GC/FT-IR	10 $\mu\text{g}/\text{L}$		EPA 1986c

TABLE 6-2. Analytical Methods for Determining Dinitrotoluene in Environmental Samples (*continued*)

Sample matrix	Preparation method ^a	Analytical method ^a	Sample detection limit ^a	Percent recovery	Reference
Soil, sediment, solid waste	Extraction	GC/FT-IR	10 µg/L	NR	Gurka et al. 1987
Soil/sediment	Air dry sample and homogenize and pass through sieve. Extract subsample with acetonitrile ultrasonically. Combine with 5.0 g/L CaCl ₂ solution and inject.	HPLC (214 nm)	0.05 mg/kg	—	Weisberg and Ellickson 1998 (modification to EPA Method 8330)
Soil, water, and municipal sludges	Extraction with dichloromethane; addition of isotopically labeled analog	GC/MS	10 µg/mL	NR	EPA 1989
Soil/compost	Acid leaching followed by sonic extraction	HPLC	0.055–0.248 mg/L ^c	NR	Griest et al. 1993
Soil/compost	Extraction with acetonitrile. Combine with CaCl ₂ . Derivatize with TFAA, then deactivate with H ₂ O.	HPLC	—	NR	Preslan et al. 1993
Soil/compost, leachates	Extract ultrasonically with acetonitrile. Filter with nylon membrane syringe filter.	HPLC uv/vis	0.4–0.5 ppm	—	Caton and Griest 1996
Materials exposed to DNT (bomb debris)	Swab with ethanol	HPLC/ED	5.6 µg/L	97 (2,6-DNT); 93 (2,4-DNT)	Lloyd 1983b

TABLE 6-2. Analytical Methods for Determining Dinitrotoluene in Environmental Samples (continued)

Sample matrix	Preparation method ^a	Analytical method ^a	Sample detection limit ^a	Percent recovery	Reference
Materials exposed to DNT	Swab surface and insert into outlet port	GC/ECD	20 ppt (2,4-DNT)	-	Nacson et al. 1994
Phenolic and nitroaromatic compounds	Inject sample, loop-separate using supercritical CO ₂ as mobile phase.	SFC (230 nm)	oxidative-250 pg reductive-100 pg	-	Wallenborg et al. 1997
Explosives	Extract in acetonitrile and dilute in pH 7 buffer. Inject hydrostatistically and detect by uv.	MECC (214 nm)	0.55–0.74 mg/L	-	Oehrle 1996

2,6-dinitrotoluene (2,6-DNT) unless otherwise noted

^aAnalyses for both 2,4-dinitrotoluene (2,4-DNT) and^bMinimum detection to ECD^cVaried over course of experiment

AMD = automated multiple development; CaCl₂ = calcium chloride; CO₂ = carbon dioxide; ECD = electron-capture detection; ED = electrochemical detection; EI = electron ionization; FT-IR = fourier transform infrared; GC = gas chromatography; HCl = hydrochloric acid; HPLC = high-performance liquid chromatography; HPTLC = high-performance thin layer chromatography; IDMS = isotope dilution mass spectrometry; LC = liquid chromatography; MECC = micellar electrokinetic capillary chromatography; MS = mass spectrometry; NaOH = sodium hydroxide; NR = not reported; PDA = photodiode array detection; PICI = positive ion chemical ionization; RPM = revolutions per minute; SE = solid extraction; SFC = super critical fluid chromatography; TDM = thermal desorption modulator interface; TEA = thermal energy analysis; TLC = thin layer chromatography; UV = ultraviolet absorption

6. ANALYTICAL METHODS

Methods for Determining Parent Compounds and Degradation Products in Environmental

Media. DNT can be analyzed in water, air, and waste samples with reasonable selectivity and sensitivity (EPA 1986a, 1986b, 1986c, 1989; Gurka et al. 1987; Nacson et al. 1994; NIOSH 1977; Yinon 1996; Yook et al. 1994). Therefore, there is a reasonable database in this area.

There exists an ongoing effort to develop a “Master Analytical Scheme” for organic compounds in water (Michael et al. 1988). The overall goal is to detect and measure quantitatively organic compounds at 0.1 $\mu\text{g/L}$ in drinking water, 1 $\mu\text{g/L}$ in surface waters, and 10 $\mu\text{g/L}$ in effluent waters. Analytes well include numerous semivolatile compounds and some compounds that are only “semi-soluble” in water, as well as volatile compounds (boiling point $<150^\circ\text{C}$). A comprehensive review of the literature leading up to these efforts has been published (Pellizzari et al. 1985). It may be anticipated that improved methods for the determination of semivolatile DNT isomers in environmental samples may be developed as part of this effort.

6.3.2 Ongoing Studies

No ongoing studies were identified on analytical methods for DNT and its metabolites.

7. REGULATIONS AND ADVISORIES

The international, national, and state regulations and guidelines regarding 2,4-DNT and 2,6-DNT in air, water, and other media are summarized in Table 7-1.

ATSDR has derived two MRL values for 2,4-DNT and one MRL value for 2,6-DNT. An acute-duration oral MRL of 0.05 mg/kg/day was derived for 2,4-DNT based on neurotoxicity in dogs (Ellis et al. 1985; Lee et al. 1978). A chronic-duration oral MRL of 0.002 mg/kg/day was derived for 2,4-DNT based on a NOAEL of 0.2 mg/kg/day for neurotoxicity, Heinz bodies, and biliary tract hyperplasia in dogs (Ellis et al. 1979, 1985). For 2,6-DNT, an intermediate-duration oral MRL of 0.004 mg/kg/day was derived based on hematological effects of splenic extramedullary erythropoiesis and lymphoid depletion in dogs (Lee et al. 1976).

The International Agency for Research on Cancer (IARC) classifies 2,4-DNT and 2,6-DNT as Group 2B carcinogens (possibly carcinogenic to humans) (IARC 1996). The U.S. EPA assigns Class 2B (human carcinogen) (EPA 1996).

2,4-DNT and 2,6-DNT are on the list of chemicals in "The Emergency Planning and Community Right-to-Know Act of 1986" (EPA 1989a, 1988a). Section 313 of Title III of the Super-fund Amendments and Reauthorization Act (SARA) requires owners and operators of certain facilities that manufacture, import, process, or otherwise use the chemicals on this list to report annually any release of those chemicals to any environmental media over a specified threshold level.

OSHA requires employers of workers who are occupationally exposed to 2,4-DNT and 2,6-DNT to institute engineering controls and work practices to reduce and maintain employee exposure at or below permissible exposure limits (PEL). If the employer can document that 2,4-DNT and 2,6-DNT are used in the workplace less than 30 days per year, the employer can use any combination of engineering controls, work practice controls, or respirators to reduce employee exposure to or below the (PEL) of 1.5 mg/m³. PELs are 8-hour time-weighted averages (TWA). Respirators must be provided and used during the time period necessary to install or implement feasible engineering and work practice controls, or where controls are not yet sufficient. Respirators are also required when the employer determines that compliance with the PEL is not feasible with engineering or work practice controls, such as maintenance and repair activities, vessel cleaning, or other operations where exposures are intermittent and limited in duration, and in emergencies (OSHA 1987).

7. REGULATIONS AND ADVISORIES

Table 7-1. Regulations and Guidelines Applicable to 2,4-DNT and 2,6-DNT

Agency	Description	Information	References
<u>INTERNATIONAL</u>			
IARC	Carcinogenic classification	Group 2B	IARC 1996
<u>NATIONAL</u>			
Regulations:			
a. Air:			
EPA	NAAQS	None listed	EPA 1995
OSHA	Permissible Exposure Limit-Time-Weighted Average	1.5 mg/m ³ (skin)	OSHA 1998 (29 CFR 1910.1000)
b. Non-specific media:			
EPA	Designated as a hazardous substance	Yes	EPA 1998b (40 CFR 116.4)
	CERCLA Reportable Quantity (2,4- and 2,6-DNT)	1,000 pounds	EPA 1998c (40 CFR 302.4)
	RCRA Reportable Quantity 2,4-DNT (U105)	10	EPA 1998c (40 CFR 302.4)
	2,6-DNT (U106)	100	
	OSW Hazardous Waste Constit- uent	Yes	EPA 1998e (40 CFR 261 Appendix VIII)
	Ground Monitoring List	Yes	EPA 1998e (40 CFR 261 Appendix IX)
Guidelines:			
a. Air:			
EPA	RfC (inhalation)	None listed	IRIS 1998
ACGIH	Threshold Limit Values (TWA)	0.2 mg/m ³ (skin)	ACGIH 1998
NIOSH	Recommended Exposure Limit for Occupational Exposure- TWA	1.5 mg/m ³ (skin)	NIOSH 1997
	Potential Occupational Carcinogen	Yes	NIOSH 1997
b. Water:			
EPA	Ambient Water Quality Criteria to Protect Human Health (2,4-DNT)		IRIS 1996
	Water and fish consumption	0.11 µg/L	
	Fish consumption	9.1 µg/L	
	Drinking Water Advisory Values 2,4-DNT-10-kg Child	EPA 1998d	
	1-Day	0.5 mg/L	
	10-Day	0.5 mg/L	
	Longer term	0.3 mg/L	

7. REGULATIONS AND ADVISORIES

**Table 7-1. Regulations and Guidelines Applicable to 2,4-DNT and 2,6-DNT
(continued)**

Agency	Description	Information	References
NATIONAL (cont'd)			
	2,4-Dinitrotoluene—Adult		
	Longer term	1.0 mg/L	
	RfD	0.002 mg/kg/day	
	DWEL	0.1 mg/L	
	Lifetime	None listed	
	2,6-Dinitrotoluene—Child		
	1-Day	0.4 mg/L	
	10-Day	0.4 mg/L	
	Longer term	0.4 mg/L	
	2,6-Dinitrotoluene—Adult		
	Longer term	1.0 mg/L	
	RfD	0.001 mg/kg/day	
	DWEL	0.04 mg/L	
	Lifetime	None listed	
c. Non-specified Media:			
ACGIH	Biological Exposure Index	None listed	ACGIH 1998
	Group (cancer ranking)	A2 ^a	ACGIH 1998
EPA	RfD (oral)		
	2,4-Dinitrotoluene	0.002 mg/kg/day	IRIS 1998
EPA	Carcinogenic Classification		EPA 1996c
	2,4-Dinitrotoluene	B2 ^b	
	2,6-Dinitrotoluene	B2 ^b	
	q ₁ * 2,4-dinitrotoluene (oral)	0.68 (mg/kg/day) ⁻¹	EPA 1992

STATERegulations and
Guidelines:

a. Air:

	Threshold Ambient Limits and Significant Emission Levels (2,4- or 2,6-DNT if not specified)		
Kentucky	(8-hour)	3.827x10 ⁻⁴ lb/hr	KY NREPC 1998
South Carolina	(24-hour)	1.50 µg/m ³	SC DHEC 1998
Vermont	(annual)	0.011 µg/m ³	VT NRA 1998
Connecticut (2,4-DNT)	(8-hour)	15 µg/m ³	NATICH 1994
Florida (2,4-DNT)	(8-hour)	15 µg/m ³	NATICH 1994
	(24-hour)	3.6 µg/m ³	
	(annual)	0.011 µg/m ³	

7. REGULATIONS AND ADVISORIES

**Table 7-1. Regulations and Guidelines Applicable to 2,4-DNT and 2,6-DNT
(continued)**

Agency	Description	Information	References
<u>STATE (cont'd)</u>			
Louisiana (2,4-DNT)	(8-hour)	35.7 $\mu\text{g}/\text{m}^3$	NATICH 1994
(2,6-DNT)	(8-hour)	35.7 $\mu\text{g}/\text{m}^3$	
Nevada (2,4-DNT)	(8-hour)	36 $\mu\text{g}/\text{m}^3$	NATICH 1994
North Dakota	(8-hour)	15 $\mu\text{g}/\text{m}^3$	ND DHCL 1998
Oklahoma (2,4-DNT)	(8-hour)	15 $\mu\text{g}/\text{m}^3$	NATICH 1994

^aGroup A2: suspected human carcinogen

^bGroup B2: probable human carcinogen, based on animal data

ACGIH = American Conference of Governmental Industrial Hygienists; CERCLA = Comprehensive Environmental Response, Compensation, and Liability Act; DWEL = drinking water equivalency level; EPA = Environmental Protection Agency; IARC = International Agency for Research on Cancer; NAAQS = National Ambient Air Quality Standards; NIOSH = National Institute for Occupational Safety and Health; OSHA = Occupational Safety and Health Administration; OSW = Office of Solid Wastes; RCRA = Resource Conservation Recovery Act; RfC = Reference Concentration; RFD = Reference Dose; TWA = Time-Weighted Average

7. REGULATIONS AND ADVISORIES

2,4-DNT and 2,6-DNT are regulated by the Clean Water Effluent Guidelines in 40 CFR Part 401. For each point source category, 2,4-DNT and 2,6-DNT may be regulated as part of a group of chemicals controlled as total toxic organics, or may have a specific regulatory limitation. The point source categories for which 2,4-DNT and 2,6-DNT are controlled as total toxic organics include electroplating and metal finishing (EPA 1998f). The point source category for which 2,4-DNT and 2,6-DNT have a specific regulatory limitation is organic chemicals, plastics, and synthetic fibers (EPA 1998g).

EPA has developed oral reference dose (RfD) of 0.002 mg/kg/day and 0.001 mg/kg/day for 2,4- and 2,6-DNT, respectively (EPA 1998d).

An oral slope factor (q_1^*) of 6.8×10^{-1} (mg/kg/day) $^{-1}$ has been derived for the 2,4-/2,6-DNT mixture and is used for the cancer risk assessment of both 2,4- and 2,6-DNT (EPA 1998).

8. REFERENCES

*Abemethy DJ, Couch DB. 1982. Cytotoxicity and mutagenicity of dinitrotoluenes in Chinese hamster ovary cells. *Mutat Res* 103:53-59.

ACGIH. 1986. Documentation of the threshold limit values and biological exposure indices. 5th ed. Cincinnati, OH: American Conference of Governmental Industrial Hygienists, Inc.

*ACGIH. 1998. Threshold limit values and biological exposure indices. 6th edition. Cincinnati, OH: American Conference of Governmental Industrial Hygienists.

*Adinolfi M. 1985. The development of the human blood-csf-brain barrier. *Dev Med Child Neurol* 27:532-537.

*Ahrenholz SH. 1980. Health hazard evaluation determination. Report no. HE 79-1 13-728. Olin Chemical Co., Brandenberg, KY. Hazard Evaluations and Technical Assistance Branch, NIOSH.

*Ahrenholz SH, Meyer CR. 1982. Health Hazard Evaluation Report, No. HETA-81-295-1155, Olin (formerly Allied) Chemical Co., Moundsville, WV. Cincinnati, OH: Hazard Evaluations and Technical Assistance Branch, NIOSH. 31 pp.

*Ahman PK, Dittmer DS. 1974. Biological handbooks: Biology data book. Volume III, 2nd ed. Bethesda, MD: Federation of American Societies for Experimental Biology, 1987-2008, 204l.

*Andersen ME, Krishnan K. 1994. Relating *in vitro* to *in vivo* exposures with physiologically based tissue dosimetry and tissue response models. In: Salem H, ed. Animal test alternatives: Refinement, reduction, replacement. New York, NY: Marcel Dekker, Inc., 9-25.

*Andersen ME, Clewell HJ III, Gargas ML, et al. 1987. Physiologically based pharmacokinetics and the risk assessment process for methylene chloride. *Toxicol Appl Pharmacol* 87:185-205.

Andersson K, Levin JO, Nilsson CA. 1983. Evaluation of solid sorbents for sampling aliphatic and aromatic nitro compounds in workroom air. *Chemosphere* 12:377-384.

*Andrews CC, Osmon JL. 1977. The effects of light on TNT and other explosives in aqueous solutions. Weapons Quality Engineering Center, Naval Weapons Support Center, Crane, Indiana. WQEC/C 77-32. NTIS AD-A0361 32.

Anonymous. 1992. Dinitrotoluene. Notice of intended change. *Appl Occup Environ Hyg* 7:62-67.

*Ashby J, Burlinson B, Lefevre PA, et al. 1985. Non-genotoxicity of 2,4,6-trinitrotoluene (TNT) to the mouse bone marrow and the rat liver: Implications for its carcinogenicity. *Arch Toxicol* 58:14-19.

*Cited in text

8. REFERENCES

Ashby J, Tennant RW. 1988. Chemical structure, *Salmonella* mutagenicity and extent of carcinogenicity as indicators of genotoxic carcinogenesis among 222 chemicals tested in rodents by the U.S. NCI/NTP. *Mutat Res* 204:17-155.

Atherton SJ, Craig BB. 1986. Laser photolysis of 2,6-dinitrotoluene in solution. *Chem Phys Lett* 127:7-12.

Atkinson R. 1987. A structure-activity relationship for the estimation of rate constants for the gas-phase reactions of OH radicals with organic compounds. *Int J Chem Kinet* 19:799-828.

*Atkinson R, Carter WPL, Damall KR, et al. 1980. A smog chamber and modeling study of the gas phase nitrogen oxides air photo-oxidation of toluene and the cresols. *Int J Chem Kinet* 12:779-836.

*ATSDR. 1989. Decision guide for identifying substance-specific data needs related to toxicological profiles. Agency for Toxic Substances and Disease Registry, Division of Toxicology, Atlanta, GA.

*ATSDR/CDC. 1990. Subcommittee report on biological indicators of organ damage. Agency for Toxic Substances and Disease Registry, Centers for Disease Control and Prevention, Atlanta, GA.

Banerjee H, Dutta SK. 1997. 2-Amino-4,6-dinitrotoluene exposure to mammalian cells causes P53 gene mutation and induces apoptotic changes [Abstract]. The Annual Meeting of the American Society for Biochemistry and Molecular Biology. *Faseb J* 11(9).

Banerjee S, Howard PH, Lande SS. 1990. General structure-vapor pressure relationships for organics. *Chemosphere* 21:1173-1180.

Barnes D, Bellin J, DeRosa C, et al. 1987. Reference dose (RfD): Description and use in health risk assessments. Volume I, Appendix A: Integrated risk information system supportive documentation. Washington, DC: U.S. Environmental Protection Agency, Office of Health and Environmental Assessment. EPA/600/8-86/032a.

*Barnes DG, Dourson M. 1988. Reference dose (RfD): Description and use in health risk assessment. *Regul Toxicol Pharmacol* 8:471-486.

Barrows SE, Cramer CJ, Truhlar DG. 1996. Factors controlling regioselectivity in the reduction of polynitroaromatics in aqueous solution. *Environ Sci Technol* 30:3028-3038.

Batz ML, Garland PM, Reiter RC, et al. 1997. Explosion and ion association chemistry of the anion radicals of 2,4,6-trinitrotoluene, 2,6-dinitrotoluene, and trinitrobenzene. *J Org Chem* 62:2045-2049.

Bauer CF, Grant CL, Jenkins TF. 1986. Interlaboratory evaluation of high performance liquid chromatographic determination of nitroorganics in munitions plant wastewater. *Anal Chem* 58:176-182.

*Bauer CF, Koza SM, Jenkins TF. 1990. Liquid chromatographic method for determination of explosives residues in soil: Collaborative study. *J Assoc Off Anal Chem* 73:541-552.

*Bausum HT, Mitchell WR, Major MA. 1992. Biodegradation of 2,4- and 2,6-dinitrotoluene by freshwater microorganisms. *J Environ Sci Health A* 27: 663-695.

8. REFERENCES

Belkin F, Bishop RW, Sheely MV. 1985. Analysis of explosives in water by capillary gas chromatography. *J Chromatogr Sci* 23:532-534.

Bermudez E, Tiller-y D, Butterworth BE. 1979. The effect of 2,4-dinitrotoluene and isomers of dinitrotoluene on unscheduled DNA synthesis in primary rat hepatocytes. *Environ Mutagen* 1:391-398.

Best EPH, Zappi ME, Fredrickson I-IL et al. 1997. Screening of aquatic and wetland plant species for phytoremediation of explosives-contaminated groundwater from the Iowa Army Ammunition Plant. *Ann NY Acad Sci* 829:179-194.

*Bloch E, Gondos B, Gatz M, et al. 1988. Reproductive toxicity of 2,4-dinitrotoluene in the rat. *Toxicol Appl Pharmacol* 94:466-472.

Bond JA, Medinsky MA, Dent JG, et al. 1981. Sex dependent metabolism and biliary excretion of carbon-14 labeled dinitrotoluene in isolated perfused rat livers. *J Pharmacol Exp Ther* 219:598-603.

Bond JA, Rickert DE. 1981. Metabolism of 2,4-dinitrotoluene by freshly isolated Fischer-344 rat primary hepatocytes. *Drug Metab Dispos* 9:10-14.

*Bouvier ES, Oehrle SA. 1995. Analysis and identification of nitroaromatic and nitramine explosives in water using HPLC and photodiode-array detection. *LC-GC* 13:120-130.

Boyd EM, Killham K, Wright J et al. 1997. Toxicity assessment of xenobiotic contaminated groundwater using lux modified *Pseudomonas fluorescens*. *Chemosphere* 35:1967-1985.

Bradley PM, Chapelle FH, Landmeyer JE et al. 1994. Microbial transformation of nitroaromatics in surface soils and aquifer materials. *Appl Environ Microbial* 60:2170-2175.

*Bradley PM, Chapelle FH, Landmeyer JE. 1995. Degradation of 2,4-DNT, 2,6-DNT, and 2,4,6-TNT by indigenous aquifer microorganisms. *Bioremediation of Recalcitrant Organics* 7:267-271.

Briggs GG, Bromilow RH, Evans AA. 1982. Relationships between lipophilicity and root uptake and translocation of non-ionized chemicals by barley. *Pestic Sci* 13:495-504.

*Bronstein AC, Currnace PL, eds. 1994. Aniline and related compounds. In: *Emergency care for hazardous materials exposure*. St. Louis, MO: CV Mosby Company, 205-207.

Bums DT, Lewis RJ. 1995. Analysis and characterization of nitroglycerin-based explosives by gas chromatography-mass spectrometry. *Anal Chim Acta* 307:89-95.

Burrows EP. 1994. Dimethyl ether and dimethyl-d6 ether chemical ionization mass spectrometry of nitramines, nitroaromatics and related compounds. *Organ Mass Spectrom* 29:315-320.

*Callahan MA, Slimak MW, Gabel N, et al. 1979. Water-related environmental fate of 129 priority pollutants. Volume II. Washington, DC: Monitoring and Data Support Division (WH-553), U.S. Environmental Protection Agency, EPA-440/4-79-029b, PB80-204381,81-1 to 82-8.

*Camanzo J, Rice CP, Jude DG, et al. 1987. Organic priority pollutants in near-shore fish from 14 Lake Michigan (USA) tributaries and embayments, 1983. *J Great Lakes Res* 13:296-309.

8. REFERENCES

*Cataldo DA, Harvey SD, Fellows RJ, et al. 1989. An evaluation of the environmental fate and behavior of munitions material (TNT, RDX) in soil and plant systems; environmental fate and behavior of TNT. Final report. Final report to U.S. Army Medical Research and Development Command. Pacific Northwest Laboratories, Battelle. NTIS ADA223546.

Caton JE, Griest WH. 1996. Determination of explosives and some metabolites of TNT in biological and environmental samples by liquid chromatography on a mixed-mode &anion column. *J Liq Chrom Rel Technol* 19:661-667.

*CDC. 1981. Reproductive abnormalities in male Chemical Workers-Kentucky. *MMWR* 30:199-205.

*Chadwick RW, George SE, Chang J, et al. 1990. Comparative gastrointestinal enzyme activity and activation of the promutagen 2,6-dinitrotoluene in male CD-I mice and male Fischer 344 rats. *Cancer Lett* 52:13-19.

Chadwick RW, George SE, Chang J, et al. 1991. Potentiation of 2,6-dinitrotoluene genotoxicity in Fischer 344 rats by pretreatment with pentachlorophenol. *Pestic Biochem Physiol* 139:168-181.

*Chadwick RW, George SE, Kohan MJ, et al. 1993. Potentiation of 2,6-dinitrotoluene genotoxicity in Fischer-344 rats by pretreatment with Aroclor 1254. *Toxicology* 80:153-171.

Chadwick RW, George SE, Kohan MJ, et al. 1995. Potentiation of 2,6-dinitrotoluene genotoxicity in Fischer-344 rats by pretreatment with coal-tar creosote. *J Toxicol Environ Health* 44:319-336.

*Chapman DE, Michener SR, Powis G. 1993. In vitro metabolism of (3H)2,6-dinitrotoluene by human and rat liver. *Toxicol in Vitro* 7:213-220.

Cheng J, Kanjo Y, Suidan MT, et al. 1996. Anaerobic biotransformation of 2,4-dinitrotoluene with ethanol as primary substrate: mutual effect of the substrates on their biotransformation. *Water Research* 30:307-314.

Chism JP, Turner MJ, Jr., Rickert DE. 1984. The metabolism and excretion of mononitrotoluenes by Fischer-344 rats. *Drug Metab Dispos* 12:596-602.

*Chiu CW, Lee LH, Wang CY, et al. 1978. Mutagenicity of some commercially available nitro compounds for *Salmonella typhimurium*. *Mutat Res* 58:11-22.

*Clark A, Deas MR, Kosmidis C, et al. 1995. Explosives vapor identification in ion mobility spectrometry using a tunable laser ionization source: a comparison with conventional 63Ni ionization. *JAERI-Conf* 95-005:521-529.

*Clewell HJ III, Andersen ME. 1985. Risk assessment extrapolations and physiological modeling. *Toxicol Ind Health* 1:111-131.

*CLPSD. 1988. Contract laboratory program's statistical database. VIAR & Company. Alexandria, VA. August 10.

CMA. 1991. Initial submission from Chemical Manufacturers Association to U.S. EPA submitting information on 2,6-dinitrotoluene acute (6-hour) inhalation toxicity study in rats with attachments. Chemical Manufacturers Association. NTIS OTS0533663.

8. REFERENCES

*CMR. 1983. Chemical Marketing Reporter, February 21: 14.

Cossum PA, Rickert DE. 1985. Metabolism of dinitrobenzenes by isolated rat hepatocytes. *Drug Metab Disp* 13:664-668.

*Couch DB, Allen PF, Abernethy DJ. 1981. The mutagenicity of dinitrotoluenes in *Salmonella typhimurium*. *Mutat Res* 90:373-383.

CRIS/USDA. 1997. Current Research Information Systems/U.S. Department of Agriculture. Beltsville, MD: U.S. Department of Agriculture.

David F, Sandra P, Stafford SS, et al. 1994. Analysis of semivolatiles by CGS-MS using pressure electronic control for increasing sensitivity. *Tee Labo* 16:938-944.

Davidova IA, Sufhta JM. 1997. Transformation of 2,4-dinitrotoluene by anaerobic bacteria [Abstract]. 97th General Meeting of the American Society for Microbiology.

Davis EM, Murray HE, Liehr JG, et al. 1981. Basic microbial degradation rates and chemical byproducts of selected organic compounds. *Water Res* 15:1125-1127.

DeBethizy JD, Rickert DE. 1984. Metabolism of nitrotoluenes by freshly isolated Fischer 344 rat hepatocytes. *Drug Metab Disp* 12:45-50.

*De Vault DS. 1985. Contaminants in fish from Great Lakes harbors and tributary mouths. *Arch Environ Contam Toxicol* 14:587-594.

*Dellarco VL, Prival MJ. 1989. Mutagenicity of nitro compounds in *Salmonella typhimurium* in the presence of flavin mononucleotide in a preincubation assay. *Environ Mol Mutagen* 13:116-127.

*Deroux JM, Gonzalez C, Le Cloirec P, et al. 1996. Analysis of extractable organic compounds in water by gas chromatography mass spectrometry: applications to surface water. *Talanta* 43:365-380.

Deuster, Ralphy, Lubahn, Natascha, Friedrich, Carsten, Kleibohmer, Wolfgang. 1997. Supercritical CO₂ assisted liquid extraction of nitroaromatic and polycyclic aromatic compounds in soil. *J Chromatog A*. 785:227-238.

Dey S, Godbole SH. 1986. Enzymological aspects of dinitrobenzene degradation. In: Kon 01, et al. eds. International Council of Scientific Unions Short Reports. Volume 6. Contemporary Themes in Biochemistry. Cambridge, England, UK, Cambridge University Press. 6:23-26.

Dey S, Godbole SH. 1987. Toxicity of m-dinitrobenzene wastes. A bioassay study. *J Environ Biol* 8(2 Supp1):201-206.

Dey S, Kanekar P, Godbole SH. 1986. Aerobic microbial degradation of dinitrobenzene. *Indian J Environ Health* 29:118-128.

*Diehl CA, Jafvert CT, Larson RA. 1995. Photochemical reactions of 2,4-dinitrotoluene in surfactant solutions. *Abs Papers American Chemical Society* 210 (1-2).

8. REFERENCES

*Dillert R, Brandt M, Fomefett I, et al. 1995. Photocatalytic degradation of trinitrotoluene and other nitroaromatic compounds. *Chemosphere* 30:2333-2341.

Dixit R, Schut HAJ, Klaunig JE, et al. 1986. Metabolism and DNA binding of 2,6-dinitrotoluene in Fischer-344 Rats and A/J mice. *Toxicol Appl Pharmacol* 82:53-61.

Duarte-Davidson R, Jones KC. 1996. Screening the environmental fate of organic contaminants in sewage sludge applied to agricultural soils: II. The potential for transfers to plants and grazing animals. *Sci Total Environ* 185:59-70.

Duester R, Lubahn N, Friedrich C. 1997. Supercritical CO₂ assisted liquid extraction of nitroaromatic and polycyclic aromatic compounds in soil. *J Chromatogr A* 785:227-238.

Dunkel VC, Zeiger E, Brusik D, et al. 1985. Reproducibility of microbial mutagenicity assays: II. Testing of carcinogens and noncarcinogens in *Salmonella typhimurium* and *Escherichia coli*. *Environ Mutagen* 7:1-248.

Dunlap KL. 1981. Nitrobenzene and nitrotoluenes. In: Kirk-Othmer encyclopedia of chemical technology. Volume 15,3rd ed. Grayson M, Eckroth D, eds. John Wiley and Sons, Inc., NY. 931.

*Einsto P, Watanabe M, Ishidate M Jr., et al. 1991. Mutagenicity of 30 chemicals in *Salmonella typhimurium* strains possessing different nitroreductase or *O-acetyltransferase* activities. *Mutat Res* 259:95-102.

*Ellenhorn MJ. 1997. Ellenhorn's medical toxicology: Diagnosis and treatment of human poisoning, 2nd ed. Baltimore, MD: Williams and Wilkins, 1366-1368.

*Ellis HV, Hagensen JH, Hodgson JR, et al. 1979. Mammalian toxicity of munitions compounds. Phase BI: Effects of lifetime exposure. Part I. 2,4-Dinitrotoluene. Final report no. 7. Midwest Research Institute, Kansas City, MO. Contract no. DAMD 17-74-C-4073, ADA077 692.

*Ellis HV, Hong CB, Lee CC, et al. 1985. Subchronic and chronic toxicity studies of 2,4-dinitrotoluene. Part I. Beagle dog. *J Am Coll Toxicol* 4:233-242.

*Ellis HV, Hodgson JR, Hwang SW, et al. 1978. Mammalian toxicity of munitions compounds. Phase I. Acute oral toxicity, primary skin and eye irritation, dermal sensitization, disposition and metabolism and Ames tests of additional compounds. Progress report no. 6. Midwest Research Institute. Kansas City, MO. Contract no. DAMD 17-74-C-4073, AD A069 444

*Emmrich M, Kaiser M, Rueden H, et al. 1993. Determination of RDX, 2,4,6-trinitrotoluene and other nitroaromatic compounds by high-performance liquid chromatography with photodiode-array detection. *J Chromatogr* 645:89-94.

EPA. 1978. U.S. Environmental Protection Agency. Chemical hazard information profile: 2,4-dinitrotoluene. EPA-560/1 l-80-011.

*EPA. 1980a. U.S. Environmental Protection Agency. Semivolatile organic compounds by isotope dilution GC-IDMS-method 1625.

8. REFERENCES

EPA. 1980b. U.S. Environmental Protection Agency. Hazardous waste; identification and listing; final and interim rules. Federal Register. May 19.45:33084-33133.

EPA. 1980~. U.S. Environmental Protection Agency. Ambient water quality criteria for dinitrotoluene. Office of Water Regulation and Standards. Washington, DC. EPA-440/5-80-045.

EPA. 1980d. U.S. Environmental Protection Agency. Guidelines and methodology used in the preparation of health effect assessment chapters of the consent decree water criteria documents. Federal Register 45:79347-79357.

EPA. 1980e. U.S. Environmental Protection Agency. Federal Register 45:79318-79379.

EPA. 1980f. U.S. Environmental Protection Agency. Chemical hazard information profiles (CHIPS) August 1, 1976, to November 20,1979. Office of Pesticides and Toxic Substances. EPA 560/1 l-80-011.

*EPA. 1982a. U.S. Environmental Protection Agency. Nitroaromatics and Isophorone-method 609. Methods for organic chemical analysis of municipal and industrial wastewater. Environmental Monitoring and Support Laboratory, Cincinnati, OH. pp. 609-1 to 609-8.

*EPA. 1982b. U.S. Environmental Protection Agency. Base/ neutrals and acids-method 625. Methods for organic chemical analysis of municipal and industrial wastewater. Environmental Monitoring and Support Laboratory, Cincinnati, OH. pp. 625-1 to 625-19.

EPA. 1983. U.S. Environmental Protection Agency. Treatability manual. Volume I. Treatability data. Office of Research and Development. February 1983.

EPA. 1985. U.S. Environmental Protection Agency. Part II. Notification requirements; reportable quantity adjustments; final rule and proposed rule. Federal Register 50:13456-13522.

*EPA. 1986a. U.S. Environmental Protection Agency. Nitroaromatics and cyclic ketones-method 8090. Test methods for evaluating solid wastes, SW-846. 3rd ed. Office of Solid Waste and Emergency Response. Washington, DC. Pp. 8090-1 to 8090-15.

*EPA. 1986b. U.S. Environmental Protection Agency. Gas chromatography/mass spectrometry for semivolatile organics: capillary column technique-method 8270. Test methods for evaluating solid wastes, SW-846. 3rd ed. Office of Solid Waste and Emergency Response. Washington, DC. Pp. 8270-1 to 8270-32.

*EPA. 1986~. U.S. Environmental Protection Agency. Capillary column analysis of semivolatile organic compounds by gas chromatography/fourier transform infrared (GC/FT-IR) spectrometry-method 8410. Test methods for evaluating solid wastes, SW-846. 3rd ed. Office of Solid Waste and Emergency Response. Washington, DC. Pp. 8410-1 to 8410-17.

*EPA. 1986d. U.S. Environmental Protection Agency. Health and environmental effects profile for dinitrotoluenes. Prepared by Office of Health and Environmental Assessment, Environmental Criteria and Assessment Office, Cincinnati, OH for Office of Solid Waste and Emergency Response, Washington, DC. ECAO-CINOP183. EPA/600/X-86/159.

8. REFERENCES

EPA. 1986e. U.S. Environmental Protection Agency. Quality criteria document for water 1986: Dinitrotoluene. EPA 440/5-86-001.

EPA. 1987a. U.S. Environmental Protection Agency. Hazardous substances; reportable quantity adjustments; proposed rule. Federal Register 50:8140.

EPA. 1987b. U.S. Environmental Protection Agency. Part II. List (Phase 1) of hazardous constituents for 6-round-water monitoring; final rule. Federal Register 52:25942-25953.

EPA. 1987c. U.S. Environmental Protection Agency. Health assessment document for beryllium. Office of Health and Environmental Assessment. Washington, DC. EPA/600/8-84-026F.

EPA. 1988a. U.S. Environmental Protection Agency. Evaluation of the potential carcinogenicity of 2,6-dinitrotoluene. Final Report. Office of Health and Environmental Assessment. Washington, DC. EPA/600/8-91/124. NTIS PB93-185411.

EPA. 1988b. U.S. Environmental Protection Agency. Methodology for evaluating potential carcinogenicity in support of reportable quantity adjustments pursuant to CERCLA section 102 (Final). EPA/600/8-89/053.

*EPA. 1989. U.S. Environmental Protection Agency. Method 1624: Volatile organic compounds by isotope dilution GCMS; Method 1625: Semivolatile organic compounds by isotope dilution GCMS. Office of Water Regulations and Standards, Industrial Technology Division. EPA 440/I-89-023.

*EPA. 1990a. Interim methods for development of inhalation reference doses. U.S. Environmental Protection Agency. EPA-600/8-90/066A.

*EPA. 1990b. U.S. Environmental Protection Agency. Interim methods for development of inhalation reference doses. Office of Research and Development. Washington, DC. EPA 600/8-90-066A.

*EPA. 1992. U.S. Environmental Protection Agency. Health advisory for 2,4- and 2,6-dinitrotoluene (DNT). Office of Water, Office of Science and Technology, Health and Ecological Criteria Division. Washington, DC. PB92-189315.

*EPA. 1995. U.S. Environmental Protection Agency. Health effects assessment summary tables. Office of Research and Development, Office of Emergency and Remedial Response, Washington, DC. EPA 540/R-95-036. NTIS publication no. PB95-921199.

*EPA. 1996. U.S. Environmental Protection Agency. Drinking water regulations and health advisories, Office of Water, Washington, DC. EPA 822-R-96-001.

*EPA. 1997a. U.S. Environmental Protection Agency. Ground-water monitoring list. Code of Federal Regulations 40 CFR 264 Appendix IX.

*EPA. 1997b. U.S. Environmental Protection Agency. Hazardous constituents. Code of Federal Regulations 40 CFR 261 Appendix VIII.

EPA. 1997c. U.S. Environmental Protection Agency. Toxic chemical release reporting: Community right-to-know. Code of Federal Regulations 40 CFR 372.

8. REFERENCES

*EPA 1998a. U.S. Environmental Protection Agency. Applicability; description of the bulk organic chemicals subcategory. 40 CFR 414.70.

*EPA. 1998b. U.S. Environmental Protection Agency. Designation of hazardous substances. Code of Federal Regulations 40 CFR 116.4.

*EPA. 1998~. U.S. Environmental Protection Agency. Designation, reportable quantities, and notification. Code of Federal Regulations 40 CFR 302.4.

*EPA 1998d. U.S. Environmental Protection Agency. Drinking water regulations and health advisories. EPA-822-B-96-002.

*EPA 1998e. U.S. Environmental Protection Agency. Identification and listing of hazardous wastes. Code of Federal Regulations 40 CFR 26 1 Appendix VIIt and IX.

*EPA 1998f. U.S. Environmental Protection Agency. Electroplating point source category. 40 CFR 413.

*EPA 1998g. U.S. Environmental Protection Agency. Synthetic fibers, organic chemicals, and plastics. 40 CFR 414.70.

*Etnier EL. 1987. Water quality criteria for 2,4-dinitrotoluene and 2,6-dinitrotoluene. Oak Ridge, TN: Oak Ridge National Laboratory. U.S. Army Medical Research and Development Command. Project order no. 84PP4845.

*FEDRIP. 1997. Federal Research in Progress: Dinitrotoluene. Dialog Information Services, Inc. February 1997.

*FEDRIP. 1998. Federal Research in Progress: Dinitrotoluene. Dialog Information Services, Inc. May 12, 1998.

*Feltes J, Levsen K, Volmer D, et al. 1990. Gas chromatographic and mass spectrometric determination of nitroaromatics in water. *J Chromatogr* 5 18:21-40.

Floret F. 1929. Medical opinions on industrial poisonings. *Zentr Gewerbehyg Unfallverhut* 16:280. (Cited in USEPA 1980c).

*Fornan SJ. 1966. Body composition of the infant. Part I: The male reference infant. Falkner F, ed. *Human development*. Philadelphia, PA: WB Saunders, 239-246.

*Foman SJ, Haschke F, Ziegler EE, Nelson SE. 1982. Body composition of reference children from birth to age 10 years. *Am J Clin Nutr* 35:1169-1175.

*Ford LS. 1981. Eye irritation test in rabbits. E.I. du Pont de Nemours and Co., Inc. Haskell Laboratory Report No. 713-81.

*Francis ES, Wu M, Farnsworth PB, et al. 1995. Supercritical fluid extraction/gas chromatography with thermal desorption modular interface and nitro-specific detection for the analysis of explosives. *Journal of Microcolumn Separations* 7:23-28.

8. REFERENCES

*Freedman DL, Shanley RS, Scholze RJ. 1996. Aerobic biodegradation of 2,4-dinitrotoluene, aminonitrotoluene isomers, and 2,4-diaminotoluene. *J Haz Mater* 49:1-14.

*FSTRAC. 1988. Summary of state and federal drinking water standards and guidelines. Federal-State Toxicology and Regulatory Alliance Committee. March 1988.

FSTRAC. 1990. Summary of state and federal drinking water standards and guidelines. Federal-State Toxicology and Regulatory Alliance Committee. February, 1990.

George SE, Allison JC, Kohan MJ, et al. 1995. Effect of arachlor on 2,6-dinitrotoluene intestinal activation in Fischer 344 rats. 26th Annual Meeting Of The Environmental Mutagen Society, St. Louis, Missouri, USA, March 12-16, 1995. *Environ Mol Mutagen* 25 (suppl. 25):18.

*George SE, Chadwick RW, Chang JJ, et al. 1992. 2,4,5-Trichlorophenoxyacetic acid influence on 2,6-dinitrotoluene-induced urine genotoxicity in Fischer 344 rats: Effect on gastrointestinal microflora and enzyme activity. *Fundam Appl Toxicol* 18:240-246.

George SE, Chadwick RW, Creason JP, et al. 1991. Effect of pentachlorophenol on the activation of 2,6-dinitrotoluene to genotoxic urinary metabolites in CD-1 mice: A comparison of GI enzyme activities and urine mutagenicity. *Environ Mol Mutagen* 18:92-101.

George SE, Kohan MJ, Warren SH. 1996. Hepatic DNA adducts and production of mutagenic urine in 2,6-dinitrotoluene-treated B6C3Fl male mice. *Cancer Lett* 102: 107-111.

*Gillett JW. 1983. A comprehensive pre-biologic screen for ecotoxicologic effects. *Environ Toxicol Chem* 2:463-476.

Gillett JW, Sedlak DA, eds. 1987. Pre-biologic screen (PBS) version 2.0. Institute for Comparative and Environmental Toxicology, Cornell University, Ithaca, NY.

Goldsworthy TL, Hamm Jr. TE, Rickert DE et al. 1986. The effect of diet on 2,6-dinitrotoluene hepatocarcinogenesis. *Carcinogenesis* 7:1909-1915.

Grant CL, Jenkins TF, Mudambi AR. 1997. Comparison of environmental chemical results for split samples analyzed in different laboratories. *J AOAC Int* 80:1129-1138.

*Grant CL, Jenkins TF, Myers KF, et al. 1995. Holding-time estimates for soils containing explosive residues: comparison of fortification vs. field contamination. *Environ Toxicol Chem* 14:1865-1874.

*Griest WH, Stewart AJ, Tyndall RL, et al. 1993. Chemical and toxicological testing of composted explosives-contaminated soil. *Environ Toxicol Chem* 12:1105-1116.

*Gruener N. 1976. Ontogenetic development of NADH-dependent methemoglobin reductase in erythrocytes of man and rat. *J Toxicol Environ Health* 1:787-791.

*Guest D, Schnell SR, Rickert DE, et al. 1982. Metabolism of 2,4-dinitrotoluene by intestinal microorganisms from rat, mouse and man. *Toxicol Appl Pharmacol* 64: 160-168.

8. REFERENCES

*Gurka DF, Titus R, Griffiths PR, et al. 1987. Evaluation of an improved single-beam gas chromatography/fourier transform infrared interface for environmental analysis. *Anal Chem* 59:2362-2369.

*Guzelian PS, Henry CJ, Olin SS. 1992. Similarities and differences between children and adults: Implications for risk assessment. Washington, DC: International Life Sciences Institute Press.

*Hable M, Stem C, Asowata C, et al. 1991. The determination of nitroaromatics and nitroamines in ground and drinking water by wide-bore capillary gas chromatography. *J Chromatogr Sci* 29: 131-135.

Haderlein SB, Weismahr KW, Schwarzenbach RP. 1996. Specific adsorption of nitraromatic explosives and pesticides to clay minerals. *Environ Sci Technol* 30:612-622.

Haidour L, Ramos JL. 1996. Identification of products resulting from the biological reduction of 2,4,6-trinitrotoluene, 2,4-dinitrotoluene, and 2,6-dinitrotoluene by *Pseudomonas* sp. *Environ Sci Technol* 30:2365-2370.

*Hallas LE, Alexander M. 1983. Microbial transformation of nitro aromatic compounds in sewage effluent. *Appl Environ Microbial* 45: 1234-1241.

*Hamill PVV, Steinberger E, Levine RJ, et al. 1982. The epidemiologic assessment of male reproductive hazard from occupational exposure to TDA and dinitrotoluene. *J Occup Med* 24:985-993.

Hamilton AS, Nixon CE. 1981. Optic atrophy and multiple neuritis developed in the manufacture of explosives. (Binitrotoluene). *JAMA* 70:2004-2006.

*Hartley WR, Anderson AC, Reimers RS, et al. 1981. Separation and determination of dinitrotoluene isomers in water by gas chromatography. *Trace Substances in Environmental Health* 15:298-302.

*Hashimoto A, Sakino H, Kojima T, et al. 1982. Sources and behavior of dinitrotoluene isomers in sea water. *Water Res* 16:891-898.

*Hashimoto Y, Tokura K, Kishi H, et al. 1984. Prediction of sea-water solubility of aromatic compounds. *Chemosphere* 13:881-888.

*HazDat. 1998. Agency for Toxic Substances and Disease Registry (ATSDR), Atlanta, GA.

*Hazleton Laboratories. 1977. A thirty-day toxicology study in Fischer-344 rats given dinitrotoluene, technical grade. Full report. Submitted to Chemical Industry Institute of Toxicology, Research Triangle Park, NC.

*Hazleton Laboratories. 1982. 104-week chronic study in rats. Dinitrotoluene. Final report Volume I of II. Submitted to Chemical Industry Institute of Toxicology, Research Triangle Park, NC.

Heinze L, Brosius M, Wiesmann U. 1995. Biological degradation of 2,4-dinitrotoluene in a continuous bioreactor and kinetic studies. *Acta Hydrochim Hydrobiol* 23:254-263.

*Heny JE. 1982. Eye irritation test in rabbits. E.I. du Pont de Nemours and Co., Inc. Haskell Laboratory Report No. 31-82.

8. REFERENCES

Hildenbrand M, Luckner L. 1995. Laboratory experiments for describing the migration of explosives in sandy aquifers. *Acta Hydrochim Hydrobiol* 23:111-120.

*Ho PC. 1986. Photooxidation of 2,4-dinitrotoluene in aqueous solution in the presence of hydrogen peroxide. *Environ Sci Technol* 20:260-267.

*Hodgson JR, Kowalski MA, Glennon JP, et al. 1976. Mutagenicity studies on 2,4-dinitrotoluene [Abstract]. *Mutat Res* 38:387.

Hoff MC. 1983. Toluene. In: Kirk-Othmer encyclopedia of chemical technology. Volume 23,3rd ed. Grayson M, Eckroth D, eds. John Wiley and Sons, Inc. 265.

*Hoffsommer JC, Glover DJ, Rosen JM. 1972. Analysis of explosives in sea water and in ocean floor sediment and fauna. Report 72-215. Naval Ordnance Laboratory, White Oak, Silver Spring, MD.

*Hake RA, Giesy JP, Zabik M, et al. 1993. Toxicity of sediments and sediment pore waters from the Grand Calumet River-Indiana Harbor are of concern. *Ecotoxicol Environ Saf* 26:86-112.

*Holen I, Mikalsen SO, Sanner T. 1990. Effects of dinitrotoluenes on morphological cell transformation and intercellular communication in Syrian hamster embryo cells. *J Toxicol Environ Health* 29:89-98.

*Hong CB, Ellis JV, Lee CC, et al. 1985. Subchronic and chronic toxicity studies of 2,4-dinitrotoluene. Part III. CD-1 mice. *J Am Co11 Toxicol* 4:257-269.

HSDB. 1988. Hazardous Substances Data Bank. On-line, 8-88. National Library of Medicine. Bethesda, MD.

*HSDB. 1998. Hazardous Substances Data Bank. National Library of Medicine. Bethesda, MD.

*Huang Q, Wang L, Han S. 1995. The genotoxicity of substituted nitrobenzenes and the quantitative structure-activity relationship studies. *Chemosphere* 30:915-923.

Huang QG, Kong LR, Liu YB et al. 1996. Relationships between molecular structure and chromosomal aberrations in in vitro human lymphocytes induced by substituted nitrobenzenes. *Bull Environ Contam Toxicol* 57:349-353.

*Hunt RJ, Neubauer NR, Picone RF. 1980. An improved procedure for sampling and analysis of dinitrotoluene vapor concentrations in workplace air. *Am Ind Hyg Assoc J* 41:592-594.

*IARC. 1996. IARC monographs on the evaluation of carcinogenic risks to humans. Vol. 65: 2,4-Dinitrotoluene and 2,6-Dinitrotoluene. Lyon, France: World Health Organization, International Agency for Research on Cancer.

IRIS. 1993. Integrated Risk Information System. U.S. Environmental Protection Agency, Washington, DC.

*IRIS. 1996. Integrated Risk Information System. US Environmental Protection Agency, Washington, DC.

*IRIS. 1998. Integrated Risk Information System. US Environmental Protection Agency, Washington, DC.

8. REFERENCES

*Jenkins TF, Leggett DC, Grant CL, et al. 1986. Reversed-phase high-performance liquid chromatographic determination of nitroorganics in munitions wastewater. *Anal Chem* 58:170-175.

*Jenkins TF, Walsh ME. 1992. Development of field screening methods for TNT, 2,4-DNT and RDX in soil. *Talanta* 39:419-428.

*Johanson CE. 1980. Permeability and vascularity of the developing brain: cerebellum vs cerebral cortex. *Brain Res* 190:3-16.

*Jokinen MP, Clakson TB, Prichard RW. 1985. Animal models in atherosclerosis research. *Exp Mol Pathol* 42: 1-28.

*Jones-Price C, Marks TA, Ledoux TA, et al. 1982. Teratological and postnatal evaluation of dinitrotoluene in Fischer-344 rats. Final report. Research Triangle Institute. Submitted to the Chemical Industry Institute of Toxicology, Research Triangle Park, NC.

Kaplan DL, Kaplan AM. 1982. 2,4,6-Trinitrotoluene surfactant complexes decomposition mutagenicity and soil leaching studies. *Environ Sci Technol* 16:566-571.

*Kedderis GL, Dyroff MC, Rickert DE. 1984. Hepatic macromolecular binding of the hepatocarcinogen 2,6-DNT and its 2,4-isomer *in vivo*; modulation by the sulfotransferase inhibitors pentachlorophenol and 2,6-dichloro-4-nitrophenol. *Carcinogenesis* 5: 1199-1204.

Kedderis GL, Rickert DE. 1985. Characterization of the oxidation of amine metabolites of nitrotoluenes by rat hepatic microsomes n and c hydroxylation. *Mol Pharmacol* 28:207-214.

Kessel S, Hauck HE. 1996. Qualitative and quantitative determination of 2,4,6-TNT, hexogen, octogen, aminonitro- and nitrocompounds in ammunition wastes with modified TLC and HPTLC precoated layers, *Chromatographia* 43:401-404.

King RD, Srinivasan A. 1996. Prediction of rodent carcinogenicity bioassays from molecular structure using inductive logic programming. *Environ Health Perspect* 104: 103 1-1040.

Klaassen CD, Amdur MO, Doull J, eds. 1986. Casarett and Doull's toxicology: The basic science of poisons. 3rd ed. New York, NY: Macmillan Publishing Company, 239.

Klaassen CD, Amdur MO, Doull J. 1996. Casarett and Doull's toxicology: The basic science of poisons. 5th ed. New York, NY: The McGraw-Hill Companies, Inc.

Kleiner AI, Stovpivskaya YUR. 1981. Digestive function of the small intestine in patients chronically intoxicated with toluene nitro derivatives. *Gig Tr Prof Zabol* 1981:23-26.

Kohan MJ, George SE, Brooks LR. 1997. Effect of alachlor treatment on the formation of 2,6-dinitrotoluene hepatic DNA adducts in Fischer 344 rats [Abstract]. *Environ Mol Mutagen* 29(Suppl 28).

*Komori M, Nishio K, Kitada M, et al. 1990. Fetus-specific expression of a form of cytochrome P-450 in human liver. *Biochemistry* 29:4430-4433.

8. REFERENCES

Korolev AA, Voitsekhovskaya TV, Bogdanov MV, et al. 1977. [Experimental data for hygienic standardization of dinitrotoluene and trinitrobenzene in surface waters.] *Gig Sanit* 10:17-20. (Russian).

*Kozuka H, Mori M, Nause Y. 1979. Studies on the metabolism and toxicity of dinitrotoluenes. *Toxicological study of 2,4-dinitrotoluene (2,4-DNT) in rats in long-term feeding.* *J Toxicol Sci* 4:221-228.

*Krishnan K, Andersen ME. 1994. Physiologically based pharmacokinetic modeling in toxicology. In: Hayes W, ed. *Principles and methods of toxicology*. 3rd ed. New York, NY: Raven Press, Ltd., 149-188.

*Krishnan K, Andersen ME, Clewell HJ III, et al. 1994. Physiologically based pharmacokinetic modeling of chemical mixtures. In: Yang, RSA, ed. *Toxicology of chemical mixtures*. New York, NY: Academic Press, 399-437.

*Kumar S, Davis AP. 1997. Heterogeneous photocatalytic oxidation of nitrotoluenes. *Water Environment Research* 69:1238-1245.

*KY NREPC. 1998. Threshold ambient limits and significant emission levels of toxic air pollutants. 401 KAR 63:022.

*La DK, Froines JR. 1992. Comparison of DNA adduct formation between 2,4 and 2,6-dinitrotoluene by 32P-postlabelling analysis. *Arch Toxicol* 66:633-640.

*La DK, Froines JR. 1993. Comparison of DNA binding between the carcinogen 2,6-dinitrotoluene and its noncarcinogenic analog 2,6-diaminotoluene. *Mutat Res* 301:79-85.

*Lane RW, Simon GS, Dougherty RW, et al. 1985. Reproductive toxicity and lack of dominant lethal effects of 2,4-dinitrotoluene in the male rat. *Drug Chem Toxicol* 8:265-280.

Larson SL. 1997. Fate of explosive contaminants in plants. *Ann NY Acad Sci* 829: 195-201.

*Lee CC, Dilley JV, Hodgson JR, et al. 1975. Mammalian toxicity of munition compounds: Phase I. Acute oral toxicity, primary skin and eye irritation, dermal sensitization, and disposition and metabolism. Report no. 1. Contract DAMD17-74-c-4073; Midwest Research Institute Project no. 3900-B.

*Lee CC, Ellis HV, Kowalski JJ, et al. 1976. Mammalian toxicity of munitions compounds. Phase II: Effects of multiple doses. Part IIh 2,6-Dinitrotoluene. Progress report no. 4. Midwest Research Institute Project no. 3900-B. Contract no. DAMD-17-74-C-4073.

*Lee CC, Ellis HV, Kowalski JJ, et al. 1978. Mammalian toxicity of munitions compounds. Phase II: Effects of multiple doses. Part II: 2,4-Dinitrotoluene. Progress report No. 3. Midwest Research Institute, Kansas City, MO. Contract no. DAMD 17-74-C-4073.

*Lee CC, Hong CB, Ellis HV, et al. 1985. Subchronic and chronic toxicity studies of 2,4-dinitrotoluene. Part II. CD rats. *J Am Coll Toxicol* 4:243-256.

*Lee YS, Hunter JV. 1985. Effect of ozonation and chlorination of Environmental Protection Agency priority pollutant. In: Jolley RL, Bull RJ, Davis WP, et al., eds. *Water chlorination: Chemistry, environmental impact and health effects*. Vo15. Chelsea, MI: Lewis Publishers, Inc. 1515-1526.

8. REFERENCES

*Leeder JS, Keams, GL. 1997. Pharmacogenetics in pediatrics: Implications for practice. *Ped Clin North America* 44:55-77.

*Leonard TB, Adams T, Popp JA. 1986. Dinitrotoluene isomer-specific enhancement of the expression of diethylnitrosamine-initiated hepatocyte foci. *Carcinogenesis* 7:1797-1803.

*Leonard TB, Graichen ME, Popp JA. 1987. Dinitrotoluene isomer-specific hepatocarcinogenesis in F344 rats. *JNCI* 79:1313-1319.

*Leonard TB, Lyght O, Popp JA. 1983. Dinitrotoluene structure-dependent initiation of hepatocytes *in vivo*. *Carcinogenesis* 4:1059-1061.

Leone JA, Flagan RC, Grosjean D, et al. 1985. An outdoor smog chamber and modeling study of tolueneneitrogen oxides photooxidation. *Int J Chem Kinet* 17:177-216.

*Leung H-W. 1993. Physiologically-based pharmacokinetic modeling. In: Ballantyne B, Marrs T, Turner P, eds. *General and applied toxicology*. New York, NY: Stockton Press, I:153-164.

*Levine RJ, Andjelkovich DA, Kersteter SL, et al. 1986a. Heart disease in workers exposed to dinitrotoluene. *J Occup Med* 28:811-816.

*Levine RJ, Andjelkovich DA, Kersteter SL, et al. 1986b. Mortality of munitions workers exposed to dinitrotoluene. Final Report. Research Triangle Park, NC: Chemical Industry Institute of Toxicology. Government Accession No. ADA 167600.

*Levine RJ, Corso RDD, Blunden PB. 1985a. Fertility of workers exposed to dinitrotoluene and TDA at three chemical plants. In: Rickert DE, ed. *Toxicity of nitroaromatic compounds*. Chemical Industry Institute of Toxicology Series. Washington, DC: Hemisphere Publishing Corp: 243-254.

*Levine RJ, Turner MJ, Crume YS, et al. 1985b. Assessing exposure to dinitrotoluene using a biological monitor. *J Occup Med* 27:627-638.

Li W, Yin P, Yang Y. 1987. Properties of TNT-degrading enzymes in intact cells of *Citrobacter freundii*. *Acta Microbial Sin* 27:257-263.

Lichtenberg JJ, Longbottom JE, Bellar TA. 1987. Analytical methods for the determination of volatile nonpolar chemicals in water and water-related environments. *Advanced Chemistry Series* 2 14:63-81.

*Lide DR, ed. 1993. *CRC handbook of chemistry and physics*. London: CRC Press, 3-489 - 3-490.

*Liu D, Thomson K, Anderson AC. 1984. Identification of nitroso compounds from biotransformation of 2,4-dinitrotoluene. *Appl Environ Microbial* 47: 1295-1298.

*Lloyd JBF. 1983a. Clean-up procedures for the examination of swabs for explosive traces by highperformance liquid chromatography with electrochemical detection at a pendant drop electrode. *J Chromatogr* 261:391-406.

*Lloyd JBF. 1983b. High-performance liquid chromatography of organic explosives components with electrochemical detection at a pendant mercury drop electrode. *J Chromatogr* 257:227-236.

8. REFERENCES

Lochmuller CH, Hui M. 1998. Calculated $\log K_{ow}$ as a guide for key-set mobile phase selection in retention prediction. *J Chromatogr Sci* 36:11-18.

*Long LM, Rickert DE. 1982. Metabolism and excretion of 2,6-dinitro-[14C]toluene *in vivo* and in isolated perfused rat livers. *Drug Metab Dispos* 10:455-458.

*Loveday KS, Lugo MH, Resnick MA, et al. 1989. Chromosome aberration and sister chromatid exchange tests in Chinese hamster ovary cells *in vitro*: II. Results with 20 chemicals, *Environ Mol Mutagen* 13:60-94.

*Mabey WR, Smith JH, Podoll RT, et al. 1982. Aquatic fate process data for organic priority pollutants. Washington, DC: U.S. Environmental Protection Agency, EPA-440/4-81-014,239-243.

*Maksimov YY. 1968. Vapor pressures of aromatic nitrocompounds at various temperatures. *Russian J Phys Chem* 42:1550-1552.

Martin JL, Comfort SD, Shea PJ. 1997. Denitration of 2,4,6-trinitrotoluene by *Pseudomonas savastanoi*. *Can J Microbial* 43:447-455.

McCormick NG, Cornell JH, Kaplan AM. 1978. Identification of biotransformation products from 2,4-dinitrotoluene. *Appl Environ Microbial* 35:945-948.

McCormick NG, Feeherry FE, Levinson HS. 1976. Microbial transformation of 2,4,6-trinitrotoluene and other aromatic compounds. *Appl Environ Microbial* 31:949-958.

*McFarlane C, Nolt C, Wickliff C, et al. 1987. The uptake, distribution and metabolism of four organic chemicals by soybean plants and barley roots. *Environ Toxicol Chem* 6:847-856.

*McGee LC, McCausland A, Plume CA, et al. 1942. Metabolic disturbances in workers exposed to dinitrotoluene. *Am J Digest Dis* 9:329-331.

*McGee LC, Reed HL, Nereim TJ, et al. 1947. Metabolic disturbances in workers exposed to dinitrotoluene during World War II. *Gastroenterology* 8:293-295.

*McGown EL, Knudsen JJ, Makovec GT, et al. 1983. Fourteen-day feeding study of 2,4-dinitrotoluene in male and female rats. U.S. Army Medical Research and Development Command, Division of Research Support, Letterman Army Institute of Research. AD-A126069.

*Medinsky MA, Dent JG. 1983. Biliary excretion and enterohepatic circulation of 2,4-dinitrotoluene metabolites in Fischer-344 Rats. *Toxicol Appl Pharmacol* 68:359-366.

Meylan WM, Howard PH. 1991. Bond contribution method for estimating Henry's Law Constants. *Environ Toxicol Chem* 10:1283-1293.

*Michael LC, Pellizari ED, Wiseman RW. 1988. Development and evaluation of a procedure for determining volatile organics in water. *Environ Sci and Technol* 22:565-570.

Michelsen OB, Ostem S. 1979. Removal of nitroglycerol and nitroglycol from a nitration plant effluent by means of solvent extraction. *Environ Sci Technol* 13:735-738.

8. REFERENCES

*Mirsalis JC, Butterworth BE. 1982. Induction of unscheduled DNA synthesis in rat hepatocytes following *in vivo* treatment with dinitrotoluene. *Carcinogenesis* 3:241-245.

*Mirsalis JC, Tyson CK, Steinmetz KL, et al. 1989. Measurement of unscheduled DNA synthesis and S-phase synthesis in rodent hepatocytes following *in vivo* treatment: Testing of 24 compounds. *Environ Mol Mutagen* 14:155-164.

Mitchell WR, Burrows EP. 1995. Nitroreduction of 2,4-dinitrotoluene *in vitro* by cytochrome P-450 induced H4IIE cells. *Chemosphere* 31:2767-77.

Mitchell WR, Dennis WH, Jr. 1982. Biodegradation of 1,3-dinitrobenzene. *J Environ Sci Health A* 17:837-853.

*Mori M, Kudo Y, Nunozawa T, et al. 1985. Intestinal metabolism of 2,4-dinitrotoluene in rats. *Chem Pharm Bull* 33:327-332.

Mori M, Kawajiri T, Sayama M, et al. 1989. Metabolism of 2,4-dinitrotoluene and 2,6-dinitrotoluene, and their dinitrobenzyl alcohols and dinitrobenzaldehydes by Wistar and Sprague-Dawley rat-liver microsomal and cytosol fractions. *Chem Pharm Bull* 37:1904-1908.

Mori M, Kawajiri T, Sayama M, et al. 1989. Metabolism of 2,6-dinitrotoluene in male Wistar rat. *Xenobiotica* 19:731-741.

Mori M, Matsuhashi T, Miyahara T, et al. 1984. Reduction of 2,4-dinitrotoluene by Wistar rat liver microsomal and cytosol fractions. *Toxicol Appl Pharmacol* 76:105-112.

*Mori M, Miyahara T, Taniguchi K, et al. 1982. Mutagenicity of 2,4-dinitrotoluene and its metabolites in *Salmonella typhimurium*. *Toxicol Lett* 13:1-5.

*Mori MA, Sayama M, Shoji M et al. 1997. Bihar-y excretion and microfloral transformation of major conjugated metabolites of 2,4-dinitrotoluene and 2,6-dinitrotoluene in the male Wistar rat. *Xenobiotica* 27:1225-1236.

*Mori MA, Shoji M, Dohrin M, et al. 1996. Further studies on the urinary metabolites of 2,4-dinitrotoluene and 2,6-dinitrotoluene in the male Wistar rat. *Xenobiotica* 26:79-88.

*Morselli PL, France-Morselli R, Bossi L. 1980. Clinical pharmacokinetics in newborns and infants. *Clin Pharmacokin* 5:485-527.

*Munch JW, Shoemaker JA, Flores P, et al. 1993. U.S. EPA method 525.1 update: Improved sample preparation and additional method analytes. *Proc Water Qual Technol Conf PT* 1:449-462.

*Nacson S, Legrady O, Siu T, et al. 1994. Improved and novel approaches for the detection of explosives. *Proc SPIE Int Sot Opt Eng* 2276(Cargo Inspection Technologies):69-78.

*NAS/NRC. 1989. Biologic markers in reproductive toxicology. National Academy of Sciences/National Research Council. Washington, DC: National Academy Press, 15-35.

8. REFERENCES

NATICH. 1987. NATICH data base report on state, local and EPA air toxics activities. Research Triangle Park, NC: National Air Toxics Information Clearinghouse. Office of Air Quality Planning and Standards, U.S. Environmental Protection Agency. July 1987.

*NATICH. 1994. National Air Toxics Information Clearinghouse: Data base report on acceptable ambient air concentration guidelines/standards concentrations, units, and averaging times report. US Environmental Protection Agency, Office of Air Quality Planning and Standards. Washington, DC. January 29, 1997.

Naumova RP, Amerkhanova NN, Belousova TO. 1982. Reductive transformation of aromatic nitro compounds by bacteria. *Mikrobiologiiia* 5 1:735-739.

Naumova RP, Amerkhanova NN, Zolotukhina LM. 1983. Nitro reduction as a key stage in microbial destruction of aromatic nitro compounds. *Prikl Biokhim Mikrobiol* 19:507-512.

*NCI. 1978. Bioassay of 2,4-dinitrotoluene for possible carcinogenicity. CAS No. 121-14-2. Washington, DC: National Cancer Institute, U.S. Department of Health, Education, and Welfare, Public Health Service, National Institutes of Health. NCI-CG-TR-54.

*ND DHCL. 1998. Maximum acceptable ambient levels. North Dakota Department of Health and Consolidated Laboratory.

Neumann HG, Van Dorp C, Zwimer-Baier I. 1995. The implications for risk assessment of measuring the relative contribution to exposure from occupation, environment and lifestyle: hemoglobin adducts from amino- and nitro-arenes. *Toxicol Lett* 82-83:771-778.

*NIOSH. 1977. National Institute for Occupational Safety and Health. NIOSH manual of analytical methods. 2nd ed. Publication no. 77-157A.

NIOSH. 1985. Current intelligence bulletin 44. Dinitrotoluenes (DNT). Cincinnati, OH: National Institute for Occupational Safety and Health. DHHS Publication No. 85-109. NTIS PB86-105913.

NIOSHb. 1985. Pocket guide to chemical hazards. Washington, DC: U.S. Department of Health and Human Services, National Institute for Occupational Safety and Health.

*NIOSH. 1992. Recommendations for occupational safety and health. Cincinnati, OH: US Department of Health and Human Services, Public Health Service, Centers for Disease Control, National Institute for Occupational Safety and Health. NIOSH publication No. 92-100.

*NIOSH. 1997. NIOSH pocket guide to chemical hazards. Cincinnati, OH: US Department of Health and Human Services, Public Health Service, Centers for Disease Control, National Institute for Occupational Safety and Health. NIOSH publication No. 94-116.

Nishino SF, Spain JC. 1996. Degradation of 2,6-dinitrotoluene by bacteria [Abstract]. Abstracts of The Annual Meeting of the American Society for Microbiology 96(0):452.

Noguera DR, Freedman DL. 1995. Characterization of the metabolites produced during the biotransformation of 2,4-dinitrotoluene under nitrate reducing conditions. 95th General Meeting Of The American Society For Microbiology, Washington, DC, USA, May 21-25, 1995. Abs Meet Am Soc Microbial 95:431.

8. REFERENCES

*Noguera DR, Freedman DL. 1996. Reduction and acetylation of 2,4-dinitrotoluene by a *Pseudomonas aeruginosa* strain. *Applied Environ Microbiol* 62:2257-2263.

*Noguera DR, Freedman DL. 1997. Characterization of products from the biotransformation of 2,4-dinitrotoluene by denitrifying enrichment cultures. *Water Environment Research* 69:260-268.

*NoIt CL. 1988. Uptake and translocation of six organic chemicals in a newly-designed plant exposure system and evaluation of plant uptake aspects of the prebiologic screen for ecotoxicologic effects. Master's thesis. Cornell University, Ithaca, NY.

*NRC. 1993. Pesticides in the diets of infants and children. National Research Council, Washington DC: National Academy Press.

*NRC. 1997. Fax transmission from Kathy Iverson regarding current EEGL and CEGL levels. National Research Council, Board on Environmental Studies and Toxicology, Washington, DC.

*NTDB. 1996. National Trade Data Bank: The export connection. U.S. Department of Commerce, Economics and Statistics Administration, Washington, DC.

Oehrle SA. 1996. Analysis of nitramine and nitroaromatic explosives by capillary electrophoresis. *J Chromatogr A* 745:233-237.

*OSHA. 1998. Table Z-1 limits for air contaminants. U.S. Department of Labor, Occupational Safety & Health Administration. Code of Federal Regulations 29 CFR 1910.1000.

*OTA. 1990. Neurotoxicity: Identifying and controlling poisons of the nervous system. Office of Technology Assessment, Washington, DC. OTA-BA-438.

*Owen GM, Brozek J. 1966. Influence of age, sex, and nutrition on body composition during childhood and adolescence. In: Falkner, ed. *Human development*. Philadelphia, PA: Saunders, 222-238.

Parker LV, Jenkins TF. 1986. Suitability of polyvinyl chloride well casings for monitoring munitions in groundwater. *Groundwater Monit Rev* 6:92-98.

Parker RG, McOwen JM, Cherolis JA. 1975. Analysis of explosives and explosive residues. 2. Thin-layer chromatography. *J Forensic Sci* 20:254.

*Parrish FW. 1977. Fungal transformation of 2,4-dinitrotoluene and 2,4,6-trinitrotoluene. *Appl Environ Microbiol* 34:232-233.

*Pearson JG, Glennon JP, Barkley JJ, et al. 1979. An approach to the toxicological evaluation of a complex industrial wastewater. *Annual Symposium on Aquatic Toxicology* 2:284-301.

*Pellizzari ED, Shelton LS, Bursey JT, et al. 1985. Master scheme for the analysis of organic compounds in water, state-of-the-art review of analytical operations. U.S. Environmental Protection Agency. Washington, DC. USEPA contract no. 68-03-2704.

8. REFERENCES

*Pennington JC. 1988. Plant uptake of 2,4,6-trinitrotoluene, 4-amino-2,6-dinitrotoluene, and 2-amino-4,6-dinitrotoluene using ¹⁴C-labeled and unlabeled compounds. Army Engineer Waterways Experiment Station, Vicksburg, MS. Report No. WES/TR/EL-88-20. NTIS AD-A203-690.

*Perkins RG. 1919. A study of the munitions intoxications in France. US Pub Health Rep 34:2335-2374.

*Phillips JH, Coraor RJ, Prescott SR. 1983. Determination of nitroaromatics in biosludges with a gas chromatograph/thermal energy alanyzer. Anal Chem 55:889-892.

*Preslan JE, Hatrel BB, Emerson M, et al. 1993. An improved method for analysis of 2,4,6-trinitrotoluene and its metabolites from compost and contaminated soils. Journal of Hazardous Materials 33:329-337.

Price CJ, Tyl RW, Marks TA, et al. 1985. Teratologic evaluation of dinitrotoluene in the Fischer 344 rat. Fund Appl Toxicol 5:948-961.

Preiß A, Levsen K, Humpfer E et al. 1996. Application of high-field proton nuclear magnetic resonance (¹H NMR) spectroscopy for the analysis of explosives and related compounds in groundwater samples - a comparison with the high-performance liquid chromatography (HPLC) method. Fresenius J Anal Chem 356:445-451.

Puacz W, Szahun W, Linke K. 1995. Catalytic determination of sulfide in blood. Analyst 120:939-941.

Rajnik S, Mitchell W. 1996. Effects of 2,4,6-trinitrotoluene and associated munitions on HSP72/73 production in a human lymphoblast cell line. In Vitro Toxicol 9: 183-190.

Ramos K, McMahon K, Alipui C, et al. 1991 a. Modulation of aortic smooth muscle cell proliferation by dinitrotoluene. Adv Exp Med Biol 283(Biological Reactive Intermediates 4):805-808.

Ramos KS, McMahon KK, Alipui C, et al. 1991b. Modulation of DNA synthesis in aortic smooth muscle cells by dinitrotoluenes. Cell Biol Toxicol 7:111-128.

*Reader SC, Foster P. 1990. The in vitro effects of four isomers of dinitrotoluene on rat Sertoli and Sertoli-germ cell cocultures: Germ cell detachment and lactate and pyruvate production. Toxicol Appl Pharmacol 106:287-294.

Richard JJ, Junk GA. 1986. Determination of munitions in water using macroreticular resins. Anal Chem 58:723-725.

Rickert DE. 1982. Metabolism and excretion of 2,6-dinitro[¹⁴C] toluene *in vivo* and in isolated perfused rat livers. Drug Metab Dispos 10:455-458.

*Rickert DE, Long RM. 1980. Tissue distribution of 2,4-dinitrotoluene and its metabolites in male and female Fischer-344 rats. Toxicol Appl Pharmacol 56:286-293.

*Rickert DE, Long RM. 1981. Metabolism and excretion of 2,4-dinitrotoluene in male and female Fischer-344 rats after different doses. Drug Metab Dispos 9:226-232.

*Rickert DE, Long RM, Krakowka S, et al. 1981. Metabolism and excretion of 2,4-(¹⁴C)dinitrotoluene in conventional and axenic Fischer-344 rats. Toxicol Appl Pharmacol 59:574-579.

8. REFERENCES

*Rickert DE, Schnell SR, Long RM. 1983. Hepatic macromolecular covalent binding and intestinal disposition of 2,4-(14C)dinitrotoluene. *J Toxicol Environ Health* 11:555-568.

*Rickert DE, Butterworth BE, Popp JA. 1984. Dinitrotoluene: Acute toxicity, oncogenicity, genotoxicity, and metabolism. *CRC Crit Rev Toxicol* 13:217-234.

Rickert DE, Irons RD, Popp JA, et al. 1986. The effects of diet on the toxicity of nitroaromatic chemicals in rodents. *Dev Toxicol Environ Sci* 12:107-14.

Rippe JM, Irwin RS, Fink MP, et al., eds. 1996. *Intensive care medicine*. 3rd ed. Vol II. Boston, MA: Little, Brown and Company, 1606-1607

Roberts WC, Abernathy CO, Commons BJ. 1995. U.S. drinking water health advisories. Nitrated munitions chemicals. *Toxicol Ecotoxicol News* 2:114-119

Rosenkrantz M, Rosenkrantz HS, Klopman G. 1997. Intercellular communication, tumor promotion and non-genotoxic carcinogenesis: relationships based upon structural considerations. *Mutat Res* 381:171-188.

*Roth M, Murphy JM. 1979. Evaluation of the ultraviolet-ozone and ultraviolet-oxidant treatment of pink water. Cincinnati, OH: U.S. Environmental Protection Agency, Office of Research and Development, Industrial Environmental Research Laboratory. EPA/600/12. NTIS PB300763.

Rowland IR, Mallett AK, Wise A, et al. 1983. Effect of dietary carrageenan and pectin on the reduction of nitrocompounds by the rat cecal micro flora. *Xenobiotica* 13:251-256.

Ryon MG, Pal BC, Talmage SS, et al. 1984. Database assessment of the health and environmental effects of munition waste products. Oak Ridge, TN: Oak Ridge National Laboratory. ORNL-60118. NTIS DE84-016512.

*Sayama M, Mori M, Shirokawa T, et al. 1989b. Mutagenicity of 2,6-dinitrotoluene and its metabolites, and their related compounds in *Salmonella typhimurium*. *Mutat Res* 226: 181-184.

Sayama M, Mori MA, Ishida M, et al. 1989a. Enterohepatic circulation of 2,4-dinitrobenzaldehyde, a mutagenic metabolite of 2,4-dinitrotoluene, in male Wistar rat. *Xenobiotica* 19:83-92.

*Sayama M, Mori MA, Maruyama Y, et al. 1993. Intestinal transformation of 2,6-dinitrotoluene in male Wistar rats. *Xenobiotica* 23:123-131.

SC DHEC. 1998. South Carolina Department of Health and Environmental Control, Bureau of Air Quality. Regulation no. 62.5 air pollution control standards, standard no. 8 toxic air pollutants.

Scheibner K, Hofrichter M, Fritsche W. 1997. Mineralization of 2-amino-4,6-dinitrotoluene by manganese peroxidase of the white-rot fungus *Nematoloma frowardii*. *Biotechnology Letters* 19(9):835-839.

Schiff LJ, Sommer HZ, Davis GT. 1978. Selective reduction of dinitrotoluene isomers by ascorbate ion. Relative rates in homogeneous solution. NTIS AD-A05 1292.

Schneider K, Hassauer M, Kalberlah F. 1994. Risk assessment of military waste sites. *Umweltwissen schaften und Schadstoff-Forschung* 6:271-276.

8. REFERENCES

*Schut HAJ, Loeb TR, Grimes LA, et al. 1983. Distribution, elimination, and test for carcinogenicity of 2,6-dinitrotoluene after intraperitoneal and oral administration to strain A mice. *J Toxicol Environ Health* 12:659-670.

Schut HAJ, Loeb TR, Stoner GD. 1982. Distribution, elimination, and test for carcinogenicity of 2,4-dinitrotoluene in strain A mice. *Toxicol Appl Pharmacol* 64:213-220.

Schut HAJ, Dixit R, Loeb TR, et al. 1985. In-vivo and in-vitro metabolism of 2,4-dinitrotoluene in strain A mice. *Biochem Pharmacol* 34:969-976.

*Setchell BP, Waites GMH. 1975. The blood testis barrier. In: Creep RO, Astwood EB, Greiger SR, eds. *Handbook of physiology: Endocrinology V*. Washington, DC: American Physiological Society.

*Shackelford WM, Keith LH. 1976. Frequency of organic compounds identified in water. Athens, GA: U.S. Environmental Protection Agency, Environmental Research Laboratory. EPA-600/4-76-062.

Shoji M, Mori M, Kawajiri T, et al. 1987. Metabolism of 2,4-dinitrotoluene, 2,4-dinitrobenzyl alcohol and 2,4-dinitrobenzaldehyde by rat liver microsomal and cytosol fraction. *Chem Pharm Bull* 4:1579-1586.

Shone MGT, Wood AV. 1974. A comparison of the uptake and translocation of some organic herbicides and a systemic fungicide by barley. I. Absorption in relation to physico-chemical properties. *J Exp Bot* 25:390-400.

*Short RD, Lee CC. 1980. Effect of some nitrotoluenes on the biotransformation of xenobiotics in rats. *Experimentia (Basel)* 36:100-101.

Sikora FJ, Behrends LL, Phillips WD et al. 1997. A microcosm study on remediation of explosives-contaminated groundwater using constructed wetland. *Ann N Y Acad Sci* 829:202-218.

*Simini M, Wentsel RS, Checkai RT, et al. 1995. Evaluation of soil toxicity at Joliet Army Munition Plant. *Environ Toxicol Chem* 14:623-630.

*Simmon VF, Kauhanen K, Tardiff RG. 1977. Mutagenic activity of chemicals identified in drinking water. In: Scott S, Bridges BA, Sohels FH, eds. *Progress in genetic toxicology*. 249-258.

*Simmons MS, Zepp RG. 1986. Influence of humic substances on photolysis of nitroaromatic compounds in aqueous systems. *Water Res* 20:899-904.

*Smith EF II, Smith HJ, Kuchar EJ. 1995. Monitoring of dinitrotoluene and its metabolites in urine by spectrophotometry of their coupled aryl diazonium salts. *Am Ind Hyg Assoc J* 56:1175-1179.

Smith KN. 1983. Determination of the reproductive effects in mice of nine selected chemicals. National Institute for Occupational Health and Safety contract no. 210-81-6011.

*Smith RP. 1996. Toxic responses of the blood. In: CD Klassen, MJ Wonsiewicz, LA Sheinis, eds. *Casarett and Doull's toxicology, the basic science of poisons*, 5th ed. New York, NY: Macmillan Publishing Company. 335-354.

8. REFERENCES

*Soares ER, Lock LF. 1980. Lack of indication of mutagenic effects of dinitrotoluenes and diaminotoluenes in mice. *Environ Mutagen* 2: 111-124.

*Sohr J, Janes W, Bongartz A. 1995. TLC analysis of nitro compounds in residual warfare site contamination. *Analysis Magazine* 23:M25-M26.

*Spanggord RJ, Mill T, Chou TW, et al. 1980. Environmental fate studies of certain munitions wastewater constituents. Phase II laboratory studies. Final Report. U.S. Army Medical Research and Development Command, Ft. Detrick, MD. Contract no. DAMD 17-78-8081.

*Spanggord RJ, Gibson BV, Keck RG, et al. 1982a. Effluent analysis of wastewater generated in the manufacture of 2,4,6-trinitrotoluene 1. Characterization study. *Environ Sci Technol* 16:229-232.

*Spanggord RJ, Mortelmans KE, Griffin AF, et al. 1982b. Mutagenicity in *Salmonella typhimurium* and structure-activity relationships of wastewater components emanating from the manufacture of trinitrotoluene. *Environ Mutagen* 4:163-179.

*Spanggord RJ, Suta BE. 1982. Effluent analysis of wastewater generated in the manufacture of 2,4,6-trinitrotoluene 2. Determination of a representative discharge of ether extractable components. *Environ Sci Technol* 16:233-236.

Spiegel K., Welsch T. 1997. Monitoring degradation processes of explosives by HPLC analysis with UV and amperometric detection. *Fresenius J Anal Chem* 357:333-337.

*Staples CA, Werner AF, Hoogheem TJ. 1985. Assessment of priority pollutant concentrations in the United States using STORET database. *Environ Toxicol Chem* 4:131-142.

*Stayner LT, Dannenberg AL, Bloom T, et al. 1993. Excess hepatobiliary cancer mortality among munitions workers exposed to dinitrotoluene. *J Occup Med* 35:291-296.

*Steuckart C, Berger-Preiss E, Levsen K. 1994. Determination of explosives and their biodegradation products in contaminated soil and water from former ammunition plants by automated multiple development high-performance thin-layer chromatography. *Anal Chem* 66:2570-2577.

*Stoner GD, Greisiger EA, Schut AJ, et al. 1984. A comparison of the lung adenoma response in Strain A/J mice after intraperitoneal and oral administration of carcinogens. *Toxicol Appl Pharmacol* 72:3 13-323.

Struijs J, Stoltenkamp J. 1986. Ultimate biodegradation of 2-, 3- and 4nitrotoluene. *Sci Total Environ* 57:161-170.

*Styles JA, Cross MF. 1983. Activity of 2,4,6-trinitrotoluene in an in vitro mammalian gene mutation assay. *Cancer Lett* 20:103-108.

Suen WC, Haigler BE, Spain JC. 1996. 2,4-dinitrotoluene dioxygenase from *Burkholderia* sp. Strain DNT: similarity to naphthalene dioxygenase. *J Bacterial* 178:4926-4934.

Sundaram K., Witorsch RJ. 1995. Toxic effects of the testes. In: Witorsch, RJ, ed. *Target organ toxicology series: Reproductive toxicology*. 2nd Edition. New York, New York: Raven Press, 99-121.

8. REFERENCES

Tas S, Lauwerys R, Lison D. 1996. Occupational hazards for the male reproductive system. *Crit Rev Toxicol* 26:261-307.

Thompson CR, Kats G, Lennox RW. 1979. Phytotoxicity of air pollutants formed by high explosive production. *Environ Sci Technol* 13:1263-1268.

*Tokiwa H, Nakagawa R, Ohnishi Y. 1981. Mutagenic assay of aromatic nitro compounds with *Salmonella typhimurium*. *Mutat Res* 91:321-325.

*Trabalka JR, Garten CT, Jr. 1982. Development of predictive models for xenobiotic bioaccumulation in terrestrial ecosystems. Oak Ridge National Laboratory, Environmental Science Division, Oak Ridge, TN. Publication no. 2037,ORNL-5869.

*TRI94. 1996. Toxic Chemical Release Inventory. National Library of Medicine, National Toxicology Program, Bethesda, MD.

*Turner MJ. 1986. Identification and quantification of urinary metabolites of dinitrotoluenes in occupationally exposed humans. *Chemical Industry Institute of Toxicology Activities* 6:1-5.

*Turner MJ, Jr, Levine RJ, Nystrom DD, et al. 1985. Identification and quantification of urinary metabolites of dinitrotoluenes in occupationally exposed humans. *Toxicol Appl Pharmacol* 80:166-174.

Urbanski T. 1984. Nitro derivatives of benzene, toluene and aromatics. In: *Chemistry and technology of explosives*. Volume 4. Pergamon Press, Oxford. 151-154.

USITC. 1983. Synthetic organic chemicals: United States production and sales, 1982. U.S. International Trade Commission, Washington, DC. Publication 1422.

*USITC. 1987. Synthetic organic chemicals: United States production and sales, 1986. U.S. International Trade Commission, Washington, DC. Publication 2009.

USITC. 1994. Synthetic organic chemicals: United States production and sales, 1992. U.S. International Trade Commission, Washington, DC. Publication 2720.

Valli K, Brock B J, Joshi DK et al. 1992. Degradation of 2,4-dinitrotoluene by the lignin-degrading fungus *Phanerochaete chrysosporium*. *Appl Environ Microbial* 58(1):221-228.

Vanderlaan M, Watkins BE, Stanker L. 1988. Environmental monitoring by immunoassay. *Environ Sci Technol* 22:247-254.

*Vemot EH, MacEwen JD, Haun CC, et al. 1977. Acute toxicity and skin corrosion data for some organic and inorganic compounds and aqueous solutions. *Toxicol Appl Pharmacol* 42:417-423.

*Vieira I, Sonnier M, Cresteil T. 1996. Developmental expression of CYP2E1 in the human liver: Hypermethylation control of gene expression during the neonatal period. *Eur J Biochem* 238:476-483.

*VT NRA. 1998. Hazard ambient air standards. Vermont air regulations

8. REFERENCES

Wallenborg SR, Ma&ides KE, Nyholm L. 1997. Oxidative and reductive amperometric detection of phenolic and nitroaromatic compounds in packed capillary column supercritical fluid chromatography. *J Chromatogr A* 785:121-128.

Warner KA, Capone DG. 1997. Degradation of 2,4-dichlorophenol in polyhaline estuarine sediments under toxic and anoxic conditions [Abstract]. 97th General Meeting.

Waters MD, Stack HF Jackson MA, et al. 1994. The performance of short-term tests in identifying potential germ cell mutagens: A qualitative and quantitative analysis. *Mutat Res* 34 1:109-131.

Weisberg CA, Ellickson ML. 1998. Practical modifications to U.S. EPA method 8330 for the analysis of explosives by HPLC. *Am Lab* 30:32N, 32P-32Q, 32S-32v.

Weiss CF, Glazko AJ, Weston JK. 1960. Chloramphenicol in the newborn infant: A physiologic explanation of its toxicity when given in excessive doses. *N Eng J Med* 262:787-794.

*West JR, Smith HW, Chasis H. 1948. Glomerular filtration rate, effective renal blood flow, and maximal tubular excretory capacity in infancy. *J Ped* 32a:10-18.

*Widdowson EM, Dickerson JWT. 1964. Chemical composition of the body. In: Comar CL, Bronner F, eds. *Mineral metabolism: An advanced treatise, Volume II: The elements part A*. New York, NY: Academic Press.

Wilbourn J, Partensky C, Morgan G. 1996. LARC evaluates printing processes and printing inks, carbon black and some nitro compounds. *Stand J Work Environ Health* 22:154-156.

*Woodruff RC, Mason JM, Valencia R, et al. 1985. Chemical mutagenesis testing in *Drosophila*. V. Results of 53 coded compounds tested for the National Toxicology Program. *Environ Mutagen* 7:677-702.

*Woollen BH, Hall MG, Craig R, et al. 1985. Dinitrotoluene: An assessment of occupational absorption during the manufacture of blasting explosives. *Jnt Arch Occup Environ Health* 55:319-330.

*Working PK, Butterworth BE. 1984. An assay to detect chemically induced DNA repair in rat spermatocytes. *Environ Mutagen* 6:273-286.

Wyrobek AJ. 1986. Application of human sperm parameters for monitoring. In: Sorsa M, Norppa H, eds. *Progress in clinical and biological research. Volume 207. Monitoring of occupational genotoxins*. New York, NY: Alan R Liss Inc, 101-120.

Yinon J. 1989. Metabolic studies of explosives 6. Electron impact and chemical ionization mass spectrometry of metabolites of 2,4-dinitrotoluene. *Biomed Environ Mass Spectrom* 18: 149-156.

*Yinon J. 1996. Trace analysis of explosives in water by gas chromatography-mass spectrometry with a temperature-programmed injector. *J Chromatogr A* 742:205-209.

*Yook KS, Hong SM, Kim JH. 1994. Comparison of liquid-liquid extraction and solid-phase extraction coupled with GC/MS for determination of priority pollutants in water. *Anal Sci Technol* 7:441-453.

8. REFERENCES

*Young WS III, Lietman PS. 1978. Chloramphenicol glucoronyl transferase: Assay, ontogeny and inducibility. *J Pharmacol Exp Ther* 204:203-211.

Zhao YH, Lang PZ. 1996. Evaluation of the partitioning of hydrophobic pollutants between aquatic and solid phases in natural systems. *Sci Total Environ* 177: 1-7.

*Ziegler EE, Edwards BB, Jensen RL, et al. 1978. Absorption and retention of lead by infants. *Pediatr Res* 12:29-34.

9. GLOSSARY

Acute Exposure-Exposure to a chemical for a duration of 14 days or less, as specified in the Toxicological Profiles.

Adsorption Coefficient (K_{oc})-The ratio of the amount of a chemical adsorbed per unit weight of organic carbon in the soil or sediment to the concentration of the chemical in solution at equilibrium.

Adsorption Ratio (K_d)-The amount of a chemical adsorbed by a sediment or soil (i.e., the solid phase) divided by the amount of chemical in the solution phase, which is in equilibrium with the solid phase, at a fixed solid/solution ratio. It is generally expressed in micrograms of chemical sorbed per gram of soil or sediment.

Bioconcentration Factor (BCF)-The quotient of the concentration of a chemical in aquatic organisms at a specific time or during a discrete time period of exposure divided by the concentration in the surrounding water at the same time or during the same period.

Cancer Effect Level (CEL)-The lowest dose of chemical in a study, or group of studies, that produces significant increases in the incidence of cancer (or tumors) between the exposed population and its appropriate control.

Carcinogen-A chemical capable of inducing cancer.

Ceiling Value-A concentration of a substance that should not be exceeded, even instantaneously.

Chronic Exposure-Exposure to a chemical for 365 days or more, as specified in the Toxicological Profiles.

Developmental Toxicity-The occurrence of adverse effects on the developing organism that may result from exposure to a chemical prior to conception (either parent), during prenatal development, or postnatally to the time of sexual maturation. Adverse developmental effects may be detected at any point in the life span of the organism.

Embryotoxicity and Fetotoxicity- Any toxic effect on the conceptus as a result of prenatal exposure to a chemical; the distinguishing feature between the two terms is the stage of development during which the insult occurred. The terms, as used here, include malformations and variations, altered growth, and in utero death.

EPA Health Advisory-An estimate of acceptable drinking water levels for a chemical substance based on health effects information. A health advisory is not a legally enforceable federal standard, but serves as technical guidance to assist federal, state, and local officials.

Immediately Dangerous to Life or Health (IDLH)-The maximum environmental concentration of a contaminant from which one could escape within 30 min without any escape-impairing symptoms or irreversible health effects.

Intermediate Exposure-Exposure to a chemical for a duration of 15-364 days, as specified in the Toxicological Profiles.

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Immunologic Toxicity-The occurrence of adverse effects on the immune system that may result from exposure to environmental agents such as chemicals.

In Vitro-Isolated from the living organism and artificially maintained, as in a test tube.

In Vivo-Occurring within the living organism.

Lethal Concentration_(LO) (LC_{LO})- The lowest concentration of a chemical in air which has been reported to have caused death in humans or animals.

Lethal Concentration₍₅₀₎ (LC₅₀)-A calculated concentration of a chemical in air to which exposure for a specific length of time is expected to cause death in 50% of a defined experimental animal population.

Lethal Dose_(LO) (LD_{LO})-The lowest dose of a chemical introduced by a route other than inhalation that is expected to have caused death in humans or animals.

Lethal Dose₍₅₀₎ (LD₅₀)-The dose of a chemical which has been calculated to cause death in 50% of a defined experimental animal population.

Lethal Time₍₅₀₎ (LT₅₀)-A calculated period of time within which a specific concentration of a chemical is expected to cause death in 50% of a defined experimental animal population.

Lowest-Observed-Adverse-Effect Level (LOAEL)-The lowest dose of chemical in a study, or group of studies, that produces statistically or biologically significant increases in frequency or severity of adverse effects between the exposed population and its appropriate control.

Malformations-Permanent structural changes that may adversely affect survival, development, or function.

Minimal Risk Level-An estimate of daily human exposure to a dose of a chemical that is likely to be without an appreciable risk of adverse noncancerous effects over a specified duration of exposure.

Mutagen-A substance that causes mutations. A mutation is a change in the genetic material in a body cell. Mutations can lead to birth defects, miscarriages, or cancer.

Neurotoxicity--The occurrence of adverse effects on the nervous system following exposure to chemical,

No-Observed-Adverse-Effect Level (NOAEL)-The dose of chemical at which there were no statistically or biologically significant increases in frequency or severity of adverse effects seen between the exposed population and its appropriate control. Effects may be produced at this dose, but they are not considered to be adverse.

Octanol-Water Partition Coefficient (K_{ow})-The equilibrium ratio of the concentrations of a chemical in n-octanol and water, in dilute solution.

Permissible Exposure Limit (PEL)-An allowable exposure level in workplace air averaged over an 8-hour shift.

9. GLOSSARY

q₁*-The upper-bound estimate of the low-dose slope of the dose-response curve as determined by the multistage procedure. The q₁* can be used to calculate an estimate of carcinogenic potency, the incremental excess cancer risk per unit of exposure (usually µg/L for water, mg/kg/day for food, and µg/m³ for air).

Reference Dose (RfD)-An estimate (with uncertainty spanning perhaps an order of magnitude) of the daily exposure of the human population to a potential hazard that is likely to be without risk of deleterious effects during a lifetime. The RfD is operationally derived from the NOAEL (from animal and human studies) by a consistent application of uncertainty factors that reflect various types of data used to estimate RfDs and an additional modifying factor, which is based on a professional judgment of the entire database on the chemical. The RfDs are not applicable to nonthreshold effects such as cancer.

Reportable Quantity (RQ)-The quantity of a hazardous substance that is considered reportable under CERCLA. Reportable quantities are (1) 1 pound or greater or (2) for selected substances, an amount established by regulation either under CERCLA or under Sect. 3 11 of the Clean Water Act. Quantities are measured over a 24-hour period.

Reproductive Toxicity-The occurrence of adverse effects on the reproductive system that may result from exposure to a chemical. The toxicity may be directed to the reproductive organs and/or the related endocrine system. The manifestation of such toxicity may be noted as alterations in sexual behavior, fertility, pregnancy outcomes, or modifications in other functions that are dependent on the integrity of this system.

Short-Term Exposure Limit (STEL)-The maximum concentration to which workers can be exposed for up to 15 min continually. No more than four excursions are allowed per day, and there must be at least 60 min between exposure periods. The daily TLV-TWA may not be exceeded.

Target Organ Toxicity-This term covers a broad range of adverse effects on target organs or physiological systems (e.g., renal, cardiovascular) extending from those arising through a single limited exposure to those assumed over a lifetime of exposure to a chemical.

Teratogen-A chemical that causes structural defects that affect the development of an organism.

Threshold Limit Value (TLV)-A concentration of a substance to which most workers can be exposed without adverse effect. The TLV may be expressed as a TWA, as a STEL, or as a CL.

Time-Weighted Average (TWA)-An allowable exposure concentration averaged over a normal 8-hour workday or 40-hour workweek.

Toxic Dose (TD₅₀)-A calculated dose of a chemical, introduced by a route other than inhalation, which is expected to cause a specific toxic effect in 50% of a defined experimental animal population.

Uncertainty Factor (UF)-A factor used in operationally deriving the RfD from experimental data. UFs are intended to account for (1) the variation in sensitivity among the members of the human population, (2) the uncertainty in extrapolating animal data to the case of human, (3) the uncertainty in extrapolating from data obtained in a study that is of less than lifetime exposure, and (4) the uncertainty in using LOAEL data rather than NOAEL data. Usually each of these factors is set equal to 10.

APPENDIX A

ATSDR MINIMAL RISK LEVELS AND WORKSHEETS

The Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA) [42 U.S.C. 9601 et seq.], as amended by the Super-fund Amendments and Reauthorization Act (SARA) [Pub. L. 99-4991, requires that the Agency for Toxic Substances and Disease Registry (ATSDR) develop jointly with the U.S. Environmental Protection Agency (EPA), in order of priority, a list of hazardous substances most commonly found at facilities on the CERCLA National Priorities List (NPL); prepare toxicological profiles for each substance included on the priority list of hazardous substances; and assure the initiation of a research program to fill identified data needs associated with the substances.

The toxicological profiles include an examination, summary, and interpretation of available toxicological information and epidemiologic evaluations of a hazardous substance. During the development of toxicological profiles, Minimal Risk Levels (MRLs) are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration for a given route of exposure. An MRL is an estimate of the daily human exposure to a hazardous substance that is likely to be without appreciable risk of adverse noncancer health effects over a specified duration of exposure. MRLs are based on noncancer health effects only and are not based on a consideration of cancer effects. These substance-specific estimates, which are intended to serve as screening levels, are used by ATSDR health assessors to identify contaminants and potential health effects that may be of concern at hazardous waste sites. It is important to note that MRLs are not intended to define clean-up or action levels.

MRLs are derived for hazardous substances using the no-observed-adverse-effect level/uncertainty factor approach. They are below levels that might cause adverse health effects in the people most sensitive to such chemical-induced effects. MRLs are derived for acute (1-14 days), intermediate (15-364 days), and chronic (365 days and longer) durations and for the oral and inhalation routes of exposure. Currently, MRLs for the dermal route of exposure are not derived because ATSDR has not yet identified a method suitable for this route of exposure. MRLs are generally based on the most sensitive chemical-induced end point considered to be of relevance to humans. Serious health effects (such as irreparable damage to the liver or kidneys, or birth defects) are not used as a basis for establishing MRLs. Exposure to a level above the MRL does not mean that adverse health effects will occur.

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MRLs are intended only to serve as a screening tool to help public health professionals decide where to look more closely. They may also be viewed as a mechanism to identify those hazardous waste sites that are not expected to cause adverse health effects. Most MRLs contain a degree of uncertainty because of the lack of precise toxicological information on the people who might be most sensitive (e.g., infants, elderly, nutritionally or immunologically compromised) to the effects of hazardous substances. ATSDR uses a conservative (i.e., protective) approach to address this uncertainty consistent with the public health principle of prevention. Although human data are preferred, MRLs often must be based on animal studies because relevant human studies are lacking. In the absence of evidence to the contrary, ATSDR assumes that humans are more sensitive to the effects of hazardous substance than animals and that certain persons may be particularly sensitive. Thus, the resulting MRL may be as much as a hundredfold below levels that have been shown to be nontoxic in laboratory animals.

Proposed MRLs undergo a rigorous review process: Health Effects/MRL Workgroup reviews within the Division of Toxicology, expert panel peer reviews, and agencywide MRL Workgroup reviews, with participation from other federal agencies and comments from the public. They are subject to change as new information becomes available concomitant with updating the toxicological profiles. Thus, h4RLs in the most recent toxicological profiles supersede previously published levels. For additional information regarding MRLs, please contact the Division of Toxicology, Agency for Toxic Substances and Disease Registry, 1600 Clifton Road, Mailstop E-29, Atlanta, Georgia 30333.

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MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical Name: 2,4-Dinitrotoluene

CAS Number: 121-14-2

Date: August 1997

Profile Status: Draft for Public Comment

Route: Inhalation Oral

Duration: Acute Intermediate Chronic

Graph Key: 9

Species: Dog

Minimal Risk Level: 0.05 mg/kg/day ppm

Reference: Ellis et al. 1985; Lee et al. 1978

Experimental design: In a subchronic (13 week) study, groups of 4 male and 4 female beagle dogs were administered 0, 1, 5, or 25 mg/kg/day 2,4-DNT in capsules. The 2,4-DNT was mixed with lactose and capsules were prepared weekly. Dogs were observed daily for behavioral changes and clinical signs. Blood was taken before treatment and at 4, 8, and 13 weeks for hematological and clinical chemistry analyses. When animals were moribund or at study termination, major organs and tissues were weighed and examined for histopathology. Bone marrow and kidney cultures were also maintained and cytogenetic analyses performed.

Effects noted in study and corresponding doses: The dose of 5 mg/kg/day produced no adverse effects. Severe neurotoxic effects were observed at 25 mg/kg/day after 12 days. Neurotoxic effects observed were incoordination and stiffness, with the hind legs being most frequently affected. This caused an abnormal hopping gait. After 22 days, some dogs at this dose were moribund.

Dose and end point used for MRL derivation:

NOAEL LOAEL 5 mg/kg/day was the NOAEL for neurological effects (incoordination, stiffness).

Uncertainty Factors used in MRL derivation:

- 10 for use of a LOAEL
- 10 for extrapolation from animals to humans
- 10 for human variability

Was a conversion used from ppm in food or water to a mg/body weight dose? No

If so, explain:

If an inhalation study in animals, list the conversion factors used in determining human equivalent dose:

Other additional studies or pertinent information which lend support to this MRL: No other acute-duration studies were located in which neurotoxicity was reported after dosing with 2,4-DNT. Slight cyanosis was observed in a dominant lethal study in rats dosed with 60 mg/kg 2,4-DNT for 5 days (Lane et al. 1985). This was the lowest dose administered in that study. Decreased fertility was found in mice dosed with 250 mg/kg 2,4-DNT for 2 days (Soares and Lock 1980). Extramedullary hematopoiesis and/or splenic hemosiderosis has been reported in intermediate-duration studies using 2,4-DNT at dietary intakes of 93 mg/kg/day (males) and 108 mg/kg/day (females) (Lee et al. 1978, 1985). Anemia and paralysis were reported in dogs dosed

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with 25 mg/kg 2,4-DNT for up to 13 weeks (Ellis et al. 1985; Lee et al. 1978). Hematological changes, such as anemia, were reported in rats fed 14-45.3 mg/kg/day in the diet for 1-2 years (Ellis et al. 1979; Hazleton Laboratories 1982; Lee et al. 1978). Methemoglobinemia and Heinz bodies were seen in dogs administered 1.5 mg/kg once a day for 24 months (Ellis et al. 1979, 1985).

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MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical Name: 2,4-Dinitrotoluene

CAS Number: 121-14-2

Date: August 1997

Profile Status: Draft for Public Comment

Route: [] Inhalation [x] Oral

Duration: [] Acute [] Intermediate [x] Chronic

Graph Key: 49

Species: Dog

Minimal Risk Level: 0.002 [x] mg/kg/day [] ppm

Reference: Ellis et al. 1979, 1985

Experimental design: Beagle dogs were administered 0, 0.2, 1.5, or 10 mg/kg 2,4-DNT in capsules for 24 months. Blood samples were taken at 3, 6, 9, 12, 18, and 24 months. After 12 months, one dog/sex/group was necropsied and the treatment for another pair from each group was discontinued for 4 weeks to examine the reversibility of effects. Actively dividing bone marrow and kidney cultures were arrested in metaphase and analyzed for chromosomal aberrations.

Effects noted in study and corresponding doses: Methemoglobinemia and Heinz bodies were observed in dogs fed 1.5 mg/kg. Biliary hyperplasia and neurotoxicity (paralysis and cerebellar lesions) were also noted at this dose. Hematological effects were not observed at the low dose. No testicular degeneration was observed up to 10 mg/kg 2,4-DNT.

Calculations: $0.2 \text{ mg/kg} \times 1/100 (\text{UF}) = 0.002 \text{ mg/kg}$

Dose and end point used for MRL derivation:

[x] NOAEL [] LOAEL 0.2 mg/kg was the NOAEL for hematological effects.

Uncertainty Factors used in MRL derivation:

- [] 10 for use of a LOAEL
- [x] 10 for extrapolation from animals to humans
- [x] 10 for human variability

Was a conversion used from ppm in food or water to a mg/body weight dose? No

If so, explain:

If an inhalation study in animals, list the conversion factors used in determining human equivalent dose:

Other additional studies or pertinent information which lend support to this MRL: Decreased red blood cell count was observed in a 2-year study in which rats were fed 3.9 mg/kg/day 2,4-DNT and anemia was observed at 34.5 mg/kg/day (Ellis et al. 1979; Lee et al. 1978, 1985). However, there were foci of altered or hyperplastic hepatocytes found in the 0.6 mg/kg/day group in this study. Hepatocellular degeneration and foci of cellular alteration were found in rats fed 27 mg/kg/day 2,4-DNT for 52 weeks (Leonard et al. 1987).

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MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical Name: 2,6-Dinitrotoluene

CAS Number: 606-20-2

Date: August 1997

Profile Status: Draft for Public Comment

Route: [] Inhalation [x] Oral

Duration: [] Acute [x] Intermediate [] Chronic

Graph Key: 10

Species: Dog

Minimal Risk Level: 0.004 [x] mg/kg/day [] ppm

Reference: Lee et al. 1976

Experimental design: Beagle dogs (4/sex/group) were administered 0, 4, 20, or 100 mg/kg 2,6-DNT in capsules for up to 13 weeks. Body weights were recorded weekly and blood samples were obtained at 4, 8, 13, and/or 17 weeks. After 4 or 13 weeks of treatment, one animal/sex/group was euthanized and another animal/sex/group had treatment discontinued for another 4 weeks to determine the reversibility of effects. Dogs that received the high dose and were placed on the reversibility study were continued on the reversibility study for 19 weeks (instead of 4) before being euthanized, due to the severity of the symptoms observed.

Effects noted in study and corresponding doses: Treatment-related mortality occurred at 20 and 100 mg/kg 2,6-DNT. No neurological effects were found in the 4-mg/kg group, but at 20 mg/kg, listlessness, incoordination, and lack of balance were found; these effects were rapidly reversible after cessation of treatment. Neurological effects became more severe at 100 mg/kg and progressed to paralysis, occasional tremors, and inability to eat. Body weight loss correlated with food consumption at 20 and 100 mg/kg. Anemia and compensatory reticulocytosis were also found at 20 and 100 mg/kg. Other treatment-related effects observed at mid and/or high dose were thymic involution, bile duct hyperplasia, testicular degeneration, hepatic inflammation, and dilated renal tubules. None of these effects were observed in animals treated with 4 mg/kg. However, after 13 weeks, mild extramedullary erythropoiesis in the spleen and lymphoid depletion were observed at 4 mg/kg; this lesion progressed in severity at higher dose levels.

Calculations: $4 \text{ mg/kg} \times 1/1,000 (\text{UF}) = 0.004 \text{ mg/kg}$

Dose and end point used for MRL derivation:

[] NOAEL [x] LOAEL 4 mg/kg for hematological effects

Uncertainty Factors used in MRL derivation:

- [x] 10 for use of a LOAEL
- [x] 10 for extrapolation from animals to humans
- [x] 10 for human variability

Was a conversion used from ppm in food or water to a mg/body weight dose? No

If so, explain:

If an inhalation study in animals, list the conversion factors used in determining human equivalent dose:

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Other additional studies or pertinent information which lend support to this MRL: After up to 13 weeks, extramedullary hematopoiesis and splenic hemosiderosis were found in rats fed 35 mg/kg/day (but not 7 mg/kg/day) and mice fed 51 mg/kg/day (but not 11 mg/kg/day) in the diet (Lee et al. 1976). Bile duct hyperplasia and decreased body weight gain were found in rats fed 35 mg/kg for up to 13 weeks (Lee et al. 1976). Hepatocytic vacuolation and degeneration were found in rats fed 7 mg/kg/day 2,6-DNT for 52 weeks (Leonard et al. 1987).

APPENDIX B

USER'S GUIDE

Chapter 1

Public Health Statement

This chapter of the profile is a health effects summary written in non-technical language. Its intended audience is the general public especially people living in the vicinity of a hazardous waste site or chemical release. If the Public Health Statement were removed from the rest of the document, it would still communicate to the lay public essential information about the chemical.

The major headings in the Public Health Statement are useful to find specific topics of concern. The topics are written in a question and answer format. The answer to each question includes a sentence that will direct the reader to chapters in the profile that will provide more information on the given topic.

Chapter 2

Tables and Figures for Levels of Significant Exposure (LSE)

Tables (2-1) 2-2, and 2-3) and figures (2-1 and 2-2) are used to summarize health effects and illustrate graphically levels of exposure associated with those effects. These levels cover health effects observed at increasing dose concentrations and durations, differences in response by species, minimal risk levels (MRLs) to humans for noncancer end points, and EPA's estimated range associated with an upper- bound individual lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. Use the LSE tables and figures for a quick review of the health effects and to locate data for a specific exposure scenario. The LSE tables and figures should always be used in conjunction with the text. All entries in these tables and figures represent studies that provide reliable, quantitative estimates of No-Observed-Adverse- Effect Levels (NOAELs), Lowest-Observed-Adverse-Effect Levels (LOAELs), or Cancer Effect Levels (CEls).

The legends presented below demonstrate the application of these tables and figures. Representative examples of LSE Table 2-1 and Figure 2-1 are shown. The numbers in the left column of the legends correspond to the numbers in the example table and figure.

LEGEND

See LSE Table 2-1

(1) Route of Exposure One of the first considerations when reviewing the toxicity of a substance using these tables and figures should be the relevant and appropriate route of exposure. When sufficient data

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exists, three LSE tables and two LSE figures are presented in the document. The three LSE tables present data on the three principal routes of exposure, i.e., inhalation, oral, and dermal (LSE Table 2-1, 2-2, and 2-3, respectively). LSE figures are limited to the inhalation (LSE Figure 2-1) and oral (LSE Figure 2-2) routes. Not all substances will have data on each route of exposure and will not therefore have all five of the tables and figures.

- (2) **Exposure Period** Three exposure periods - acute (less than 15 days), intermediate (15-364 days), and chronic (365 days or more) are presented within each relevant route of exposure. In this example, an inhalation study of intermediate exposure duration is reported. For quick reference to health effects occurring from a known length of exposure, locate the applicable exposure period within the LSE table and figure.
- (3) **Health Effect** The major categories of health effects included in LSE tables and figures are death, systemic, immunological, neurological, developmental, reproductive, and cancer. NOAELs and LOAELs can be reported in the tables and figures for all effects but cancer. Systemic effects are further defined in the “System” column of the LSE table (see key number 18).
- (4) **Key to Figure** Each key number in the LSE table links study information to one or more data points using the same key number in the corresponding LSE figure. In this example, the study represented by key number 18 has been used to derive a NOAEL and a Less Serious LOAEL (also see the 2 “18r” data points in Figure 2-1).
- (5) **Species** The test species, whether animal or human, are identified in this column. Section 2.5, “Relevance to Public Health,” covers the relevance of animal data to human toxicity and Section 2.3, “Toxicokinetics,” contains any available information on comparative toxicokinetics. Although NOAELs and LOAELs are species specific, the levels are extrapolated to equivalent human doses to derive an MRL.
- (6) **Exposure Frequency/Duration** The duration of the study and the weekly and daily exposure regimen are provided in this column. This permits comparison of NOAELs and LOAELs from different studies. In this case (key number 18), rats were exposed to 1,1,2,2-tetrachloroethane via inhalation for 6 hours per day, 5 days per week, for 3 weeks. For a more complete review of the dosing regimen refer to the appropriate sections of the text or the original reference paper, i.e., Nitschke et al. 1981.
- (7) **System** This column further defines the systemic effects. These systems include: respiratory, cardiovascular, gastrointestinal, hematological, musculoskeletal, hepatic, renal, and dermal/ocular. “Other” refers to any systemic effect (e.g., a decrease in body weight) not covered in these systems. In the example of key number 18,1 systemic effect (respiratory) was investigated.
- (8) **NOAEL** A No-Observed-Adverse-Effect Level (NOAEL) is the highest exposure level at which no harmful effects were seen in the organ system studied. Key number 18 reports a NOAEL of 3 ppm for

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the respiratory system which was used to derive an intermediate exposure, inhalation MRL of 0.005 ppm (see footnote “b”).

- (9) **LOAEL** A Lowest-Observed-Adverse-Effect Level (LOAEL) is the lowest dose used in the study that caused a harmful health effect. LOAELs have been classified into “Less Serious” and “Serious” effects. These distinctions help readers identify the levels of exposure at which adverse health effects first appear and the gradation of effects with increasing dose. A brief description of the specific endpoint used to quantify the adverse effect accompanies the LOAEL. The respiratory effect reported in key number 18 (hyperplasia) is a Less serious LOAEL of 10 ppm. Ml&s are not derived from Serious LOAELs.
- (10) **Reference** The complete reference citation is given in chapter 8 of the profile.
- (11) **CEL** A Cancer Effect Level (CEL) is the lowest exposure level associated with the onset of carcinogenesis in experimental or epidemiologic studies. CELs are always considered serious effects. The LSE tables and figures do not contain NOAELs for cancer, but the text may report doses not causing measurable cancer increases.
- (12) **Footnotes** Explanations of abbreviations or reference notes for data in the LSE tables are found in the footnotes. Footnote “b” indicates the NOAEL of 3 ppm in key number 18 was used to derive an MFU of 0.005 ppm.

LEGEND

See Figure 2-1

LSE figures graphically illustrate the data presented in the corresponding LSE tables. Figures help the reader quickly compare health effects according to exposure concentrations for particular exposure periods.

- (13) **Exposure Period** The same exposure periods appear as in the LSE table. In this example, health effects observed within the intermediate and chronic exposure periods are illustrated.
- (14) **Health Effect** These are the categories of health effects for which reliable quantitative data exists. The same health effects appear in the LSE table.
- (15) **Levels of Exposure** concentrations or doses for each health effect in the LSE tables are graphically displayed in the LSE figures. Exposure concentration or dose is measured on the log scale “y” axis. Inhalation exposure is reported in mg/m³ or ppm and oral exposure is reported in mg/kg/day.
- (16) **NOAEL** In this example, 18r NOAEL is the critical endpoint for which an intermediate inhalation exposure MRL is based. As you can see from the LSE figure key, the open-circle symbol indicates to a

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NOAEL for the test species-rat. The key number 18 corresponds to the entry in the LSE table. The dashed descending arrow indicates the extrapolation from the exposure level of 3 ppm (see entry 18 in the Table) to the MRL of 0.005 ppm (see footnote "b" in the LSE table).

- (17) CEL Key number 38r is 1 of 3 studies for which Cancer Effect Levels were derived. The diamond symbol refers to a Cancer Effect Level for the test species-mouse. The number 38 corresponds to the entry in the LSE table.
- (18) Estimated Upper-Bound Human Cancer Risk Levels This is the range associated with the upper-bound for lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. These risk levels are derived from the EPA's Human Health Assessment Group's upper-bound estimates of the slope of the cancer dose response curve at low dose levels (q_1^*).
- (19) Key to LSE Figure The Key explains the abbreviations and symbols used in the figure.

The Relevance to Public Health section provides a health effects summary based on evaluations of existing toxicologic, epidemiologic, and toxicokinetic information. This summary is designed to present interpretive, weight-of-evidence discussions for human health end points by addressing the following questions.

- 1 . What effects are known to occur in humans?
- 2 . What effects observed in animals are likely to be of concern to humans?
- 3 . What exposure conditions are likely to be of concern to humans, especially around hazardous waste sites?

The section covers end points in the same order they appear within the Discussion of Health Effects by Route of Exposure section, by route (inhalation, oral, dermal) and within route by effect. Human data are presented first, then animal data. Both are organized by duration (acute, intermediate, chronic). *In vitro* data and data from parenteral routes (intramuscular, intravenous, subcutaneous, etc.) are also considered in this section. If data are located in the scientific literature, a table of genotoxicity information is included.

The carcinogenic potential of the profiled substance is qualitatively evaluated, when appropriate, using existing toxicokinetic, genotoxic, and carcinogenic data. ATSDR does not currently assess cancer potency or perform cancer risk assessments. Minimal risk levels (MRLs) for noncancer end points (if derived) and the end points from which they were derived are indicated and discussed.

Limitations to existing scientific literature that prevent a satisfactory evaluation of the relevance to public health are identified in the Data Needs section.

SAMPLE

TABLE 2-1. Levels of Significant Exposure to [Chemical x] – Inhalation

Key to figure ^a	Species	Exposure frequency/duration	System	NOAEL (ppm)	LOAEL (effect)		Reference
					Less serious (ppm)	More serious (ppm)	
INTERMEDIATE EXPOSURE							
3	Systemic	↓	↓	↓	↓	↓	↓
4	18	Rat	13 wk 5d/wk 6hr/d	Resp	3 ^b	10 (hyperplasia)	Nitschke et al. 1981
CHRONIC EXPOSURE							
Cancer							
38	Rat	18 mo 5d/wk 7hr/d			20	(CEL, multiple organs)	Wong et al. 1982
39	Rat	89–104 wk 5d/wk 6hr/d			10	(CEL, lung tumors, nasal tumors)	NTP 1982
40	Mouse	79–103 wk 5d/wk 6hr/d			10	(CEL, lung tumors, hemangiosarcomas)	NTP 1982

^a The number corresponds to entries in Figure 2-1.

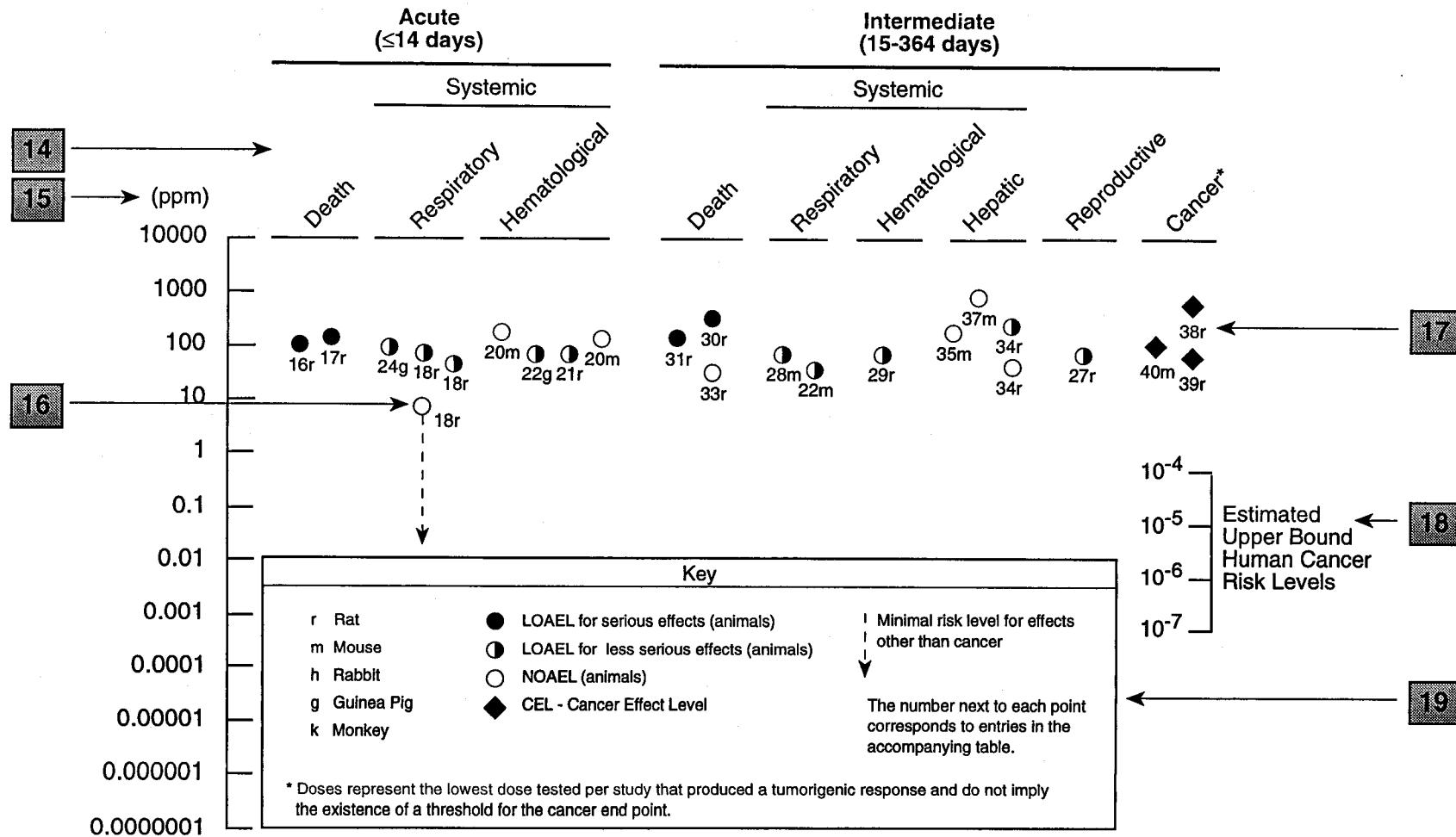
^b

an uncertainty factor of 100 (10 for extrapolation from animal to humans, 10 for human variability).

SAMPLE

13

Figure 2-1. Levels of Significant Exposure to [Chemical X] – Inhalation



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Chapter 2 (Section 2.5)

Relevance to Public Health

Interpretation of Minimal Risk Levels

Where sufficient toxicologic information is available, we have derived minimal risk levels (MRLs) for inhalation and oral routes of entry at each duration of exposure (acute, intermediate, and chronic). These MRLs are not meant to support regulatory action; but to acquaint health professionals with exposure levels at which adverse health effects are not expected to occur in humans. They should help physicians and public health officials determine the safety of a community living near a chemical emission, given the concentration of a contaminant in air or the estimated daily dose in water. MRLs are based largely on toxicological studies in animals and on reports of human occupational exposure.

MRL users should be familiar with the toxicologic information on which the number is based. Chapter 2.5, “Relevance to Public Health,” contains basic information known about the substance. Other sections such as 2.8, “Interactions with Other Substances,” and 2.9, “Populations that are Unusually Susceptible” provide important supplemental information.

MRL users should also understand the MRL derivation methodology. MRLs are derived using a modified version of the risk assessment methodology the Environmental Protection Agency (EPA) provides (Barnes and Dourson 1988) to determine reference doses for lifetime exposure (RfDs).

To derive an MRL, ATSDR generally selects the most sensitive endpoint which, in its best judgement, represents the most sensitive human health effect for a given exposure route and duration. ATSDR cannot make this judgement or derive an MRL unless information (quantitative or qualitative) is available for all potential systemic, neurological, and developmental effects. If this information and reliable quantitative data on the chosen endpoint are available, ATSDR derives an MRL using the most sensitive species (when information from multiple species is available) with the highest NOAEL that does not exceed any adverse effect levels. When a NOAEL is not available, a lowest-observed-adverse-effect level (LOAEL) can be used to derive an MRL, and an uncertainty factor (UP) of 10 must be employed. Additional uncertainty factors of 10 must be used both for human variability to protect sensitive subpopulations (people who are most susceptible to the health effects caused by the substance) and for interspecies variability (extrapolation from animals to humans). In deriving an MRL, these individual uncertainty factors are multiplied together. The product is then divided into the inhalation concentration or oral dosage selected from the study. Uncertainty factors used in developing a substance-specific MRL are provided in the footnotes of the LSE Tables.

APPENDIX C

ACRONYMS, ABBREVIATIONS, AND SYMBOLS

ACGIH	American Conference of Governmental Industrial Hygienists
ADME	Absorption, Distribution, Metabolism, and Excretion
atm	atmosphere
ATSDR	Agency for Toxic Substances and Disease Registry
BCF	bioconcentration factor
BSC	Board of Scientific Counselors
C	Centigrade
CDC	Centers for Disease Control
CEL	Cancer Effect Level
CERCLA	Comprehensive Environmental Response, Compensation, and Liability Act
CFR	Code of Federal Regulations
CLP	Contract Laboratory Program
cm	centimeter
CNS	central nervous system
d	day
DHEW	Department of Health, Education, and Welfare
DHHS	Department of Health and Human Services
DOL	Department of Labor
ECG	electrocardiogram
EEG	electroencephalogram
EPA	Environmental Protection Agency
EKG	see ECG
F	Fahrenheit
F ₁	first filial generation
FAO	Food and Agricultural Organization of the United Nations
FEMA	Federal Emergency Management Agency
FIFRA	Federal Insecticide, Fungicide, and Rodenticide Act
fpm	feet per minute
ft	foot
FR	<i>Federal Register</i>
g	gram
GC	gas chromatography
gen	generation
HPLC	high-performance liquid chromatography
hr	hour
IDLH	Immediately Dangerous to Life and Health

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IARC	International Agency for Research on Cancer
ILO	International Labor Organization
in	inch
Kd	adsorption ratio
kg	kilogram
kgg	metric ton
K _{oc}	organic carbon partition coefficient
K _{ow}	octanol-water partition coefficient
L	liter
LC	liquid chromatography
LC _{Lo}	lethal concentration, low
LC ₅₀	lethal concentration, 50% kill
LD _{Lo}	lethal dose, low
LD ₅₀	lethal dose, 50% kill
LOAEL	lowest-observed-adverse-effect level
LSE	Levels of Significant Exposure
m	meter
mg	milligram
min	minute
mL	milliliter
mm	millimeter
mmHg	millimeters of mercury
mmol	millimole
mo	month
mppcf	millions of particles per cubic foot
MRL	Minimal Risk Level
MS	mass spectrometry
NIEHS	National Institute of Environmental Health Sciences
NIOSH	National Institute for Occupational Safety and Health
NIOSHTIC	NIOSH's Computerized Information Retrieval System
ng	nanogram
nm	nanometer
NHANES	National Health and Nutrition Examination Survey
nmol	nanomole
NOAEL	no-observed-adverse-effect level
NOES	National Occupational Exposure Survey
NOHS	National Occupational Hazard Survey
NPL	National Priorities List
NRC	National Research Council
NTIS	National Technical Information Service
NTP	National Toxicology Program
OSHA	Occupational Safety and Health Administration

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PEL	permissible exposure limit
pg	picogram
pmol	picomole
PHS	Public Health Service
PMR	proportionate mortality ratio
ppb	parts per billion
ppm	parts per million
ppt	parts per trillion
REL	recommended exposure limit
RfD	Reference Dose
RTECS	Registry of Toxic Effects of Chemical Substances
sec	second
SCE	sister chromatid exchange
SIC	Standard Industrial Classification
SMR	standard mortality ratio
STEL	short term exposure limit
STORET	STORAGE and RETRIEVAL
Tg-DNT	technical grade dinitrotoluene
TLV	threshold limit value
TSCA	Toxic Substances Control Act
TRI	Toxics Release Inventory
TWA	time-weighted average
U.S.	United States
UF	uncertainty factor
yr	year
WHO	World Health Organization
wk	week
>	greater than
≥	greater than or equal to
=	equal to
<	less than
≤	less than or equal to
%	percent
α	alpha
β	beta
δ	delta
γ	gamma
μm	micrometer
μg	microgram

